

UNIVERSIDADE ESTADUAL DE CAMPINAS  
SISTEMA DE BIBLIOTECAS DA UNICAMP  
REPOSITÓRIO DA PRODUÇÃO CIENTÍFICA E INTELECTUAL DA UNICAMP

**Versão do arquivo anexado / Version of attached file:**

Versão do Editor / Published Version

**Mais informações no site da editora / Further information on publisher's website:**

<https://www.mdpi.com/1424-2818/15/7/861>

**DOI:** <https://doi.org/10.3390/d15070861>

**Direitos autorais / Publisher's copyright statement:**

©2023 by MDPI. All rights reserved.

DIRETORIA DE TRATAMENTO DA INFORMAÇÃO

Cidade Universitária Zeferino Vaz Barão Geraldo




CEP 13083-970 – Campinas SP

Fone: (19) 3521-6493

<http://www.repositorio.unicamp.br>

## Article

# Morphometric Analysis of a Trapdoor Spider (Araneae, Idiopidae) across Different Brazilian Biomes Reveals the Geographic Variation of Spiders from the Caatinga Biome

Rafael Fonseca-Ferreira <sup>1,\*</sup> , Millke Jasmine A. Morales <sup>2</sup>, Leonardo S. Carvalho <sup>3</sup>   
and José Paulo L. Guadanucci <sup>4</sup> 

- <sup>1</sup> Pós-graduação em Ecologia, Evolução e Biodiversidade, Instituto de Biociências, Universidade Estadual Paulista, Avenida 24A, 1515, Rio Claro 131506-900, SP, Brazil
  - <sup>2</sup> Departamento de Genética, Evolução, Microbiologia e Imunologia, Instituto de Biologia, Universidade Estadual de Campinas, Campinas 13083-970, SP, Brazil; millke@unicamp.br
  - <sup>3</sup> Campus Amílcar Ferreira Sobral, Universidade Federal do Piauí, BR 343, KM 3.5, Bairro Meladão, CEP, Floriano 64808-660, PI, Brazil; carvalho@ufpi.edu.br
  - <sup>4</sup> Departamento de Biodiversidade, Instituto de Biociências, Universidade Estadual Paulista, Avenida 24A, 1515, Rio Claro 131506-900, SP, Brazil; jose.guadanucci@unesp.br
- \* Correspondence: rafaelbioferreira@gmail.com

**Abstract:** Widespread species are exposed to different environmental drivers and can consequently present variations in body shape and/or size. Trapdoor spiders of the genus *Idiops* are generally associated with a sedentary lifestyle and limited dispersion. However, the species *Idiops pirassununguensis* has a wide distribution, occurring in a diverse range of distinct environmental conditions, with their presence recorded in the Amazon, Caatinga, and Cerrado Brazilian biomes. We investigated how their morphological variation is structured regarding the biomes in which they occur through a morphometric analysis of the linear measurements and morphogeometric shapes of 64 specimens. Combining different methods proved to be a valuable approach to understanding how the spider's morphology varies in different environments. The results were congruent and complementary, indicating intraspecific geographic variation, with the Caatinga specimens being distinct from their biome conspecifics. In Caatinga, a biome with periods of severe drought and warm climate, *I. pirassununguensis* specimens were found to be smaller, in addition to having narrower and elongated sternums and shorter legs. The morphological structuring herein is consistent with the results found comparing animals from Caatinga and other Brazilian biomes. Despite differences in their non-sexual structures, the specimens share a set of diagnostic sexual characteristics for the species, allowing all individuals to be classified as belonging to the same species.

**Keywords:** morphometry; *Idiops*; Mygalomorphae; idiopinae; landmarks; Fourier analysis; discriminant analysis



**Citation:** Fonseca-Ferreira, R.; Morales, M.J.A.; Carvalho, L.S.; Guadanucci, J.P.L. Morphometric Analysis of a Trapdoor Spider (Araneae, Idiopidae) across Different Brazilian Biomes Reveals the Geographic Variation of Spiders from the Caatinga Biome. *Diversity* **2023**, *15*, 861. <https://doi.org/10.3390/d15070861>

Academic Editors: Vera Opatova and Michael Wink

Received: 4 February 2023

Revised: 11 July 2023

Accepted: 14 July 2023

Published: 16 July 2023



**Copyright:** © 2023 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (<https://creativecommons.org/licenses/by/4.0/>).

## 1. Introduction

The existence of morphological, ecological, and/or behavioral differences between populations distributed throughout a species' natural range is known as geographic variation [1–3]. Therefore, individuals of the same species that are exposed to different environmental drivers may present variations in body shape and/or size, which are often a reflection of these organisms adapting to different environments [2,4]. Due to its large dimensions, Brazil encompasses a wide range of environmental conditions, from grasslands (e.g., Pampa) and savannas (i.e., Cerrado), to dry (i.e., Caatinga) and humid (Amazon and Atlantic) forests. These biomes are shaped by both abiotic (e.g., temperature, precipitation, aridity, and relief and soil characteristics) and biotic (e.g., vegetation cover and the availability of organic resources) characteristics [5,6]. These heterogeneous environments might

influence geographic variation both at an intraspecific level, especially in the case of widely distributed species, and at an interspecific level [1].

When assessing the geographic variation within a species, the most common tools require measurements of body sizes and shapes, the core of morphometric analysis [7]. These analyses are generally performed using two main types of analytical methods, which are directly related to the nature of the analyzed structures. The first is traditional or linear morphometry, based on the analysis of linear measurements, such as the length, width, and height of the body, and other morphological traits [8]. The second, more recent, is geometric morphometry, which aims to analyze and quantify the differences between complex morphological shapes, through multivariate analysis and the visualization of variation in a morphospace [8,9]. Based on these morphometric tools, it is possible to explore several biological factors, ranging from the recognition and delimitation of possible cryptic or morphologically related species [10,11] to the exploration of patterns of ecogeographic variation [12–14].

Based on a combined perspective, which involves measurements of both linear and geometric structures, few studies have investigated the possible patterns of geographic variation between different environments [15–17]. These tools have been used to assess several Brazilian taxa, with conflicting results. A study carried out with bee populations throughout the country did not find evidence of strong morphological variation between the analyzed populations [18]. However, similar studies with grasshoppers [19], beetles [20], and bats [21] found geographically structured populations.

In spiders, few studies have explored morphometric variation throughout the range of a species [22–28]. The majority of the existing morphometric studies are focused on species delimitation and on the investigation of sexual dimorphism [22–24,28–34]. There are also some studies that focus on habitat preference and foraging ecology [35,36], and others that investigate the morphological variation of specific structures in a phylogenetic context [36–39]. Despite the diversity of approaches, morphometric analyses based on multivariate and morphogeometric data have only been applied to Mygalomorph spiders on a few occasions [28,29,38–40].

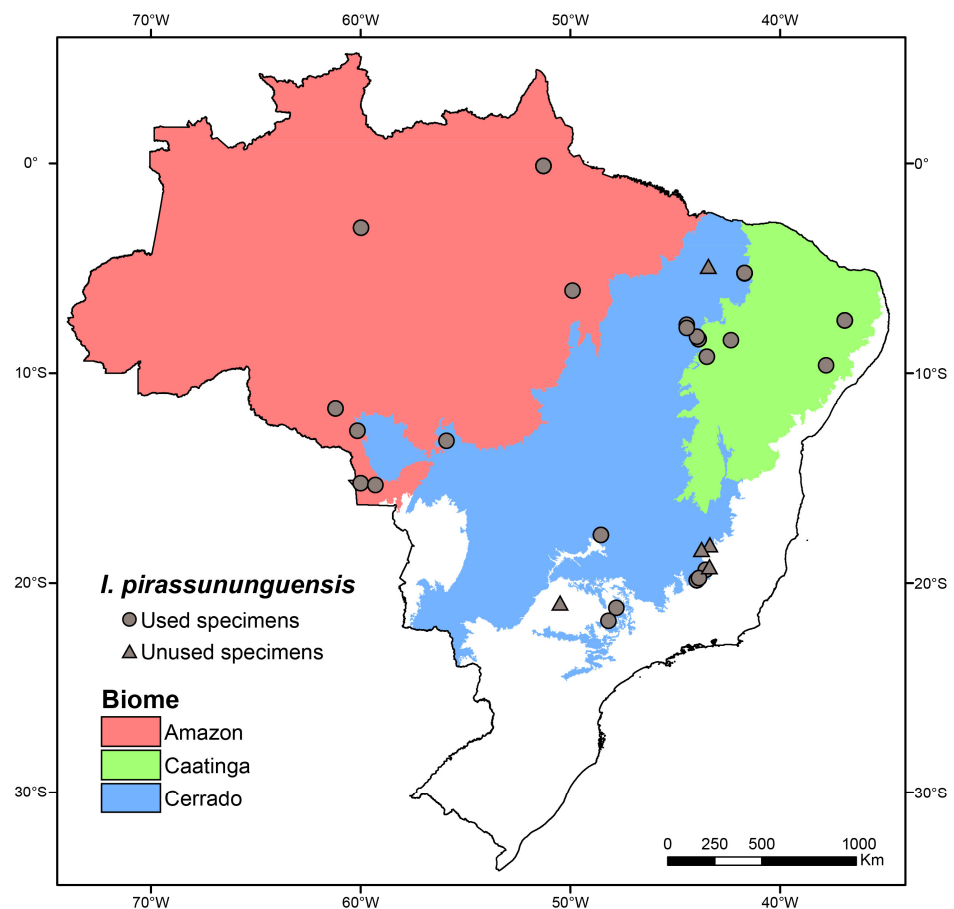
Mygalomorph trapdoor spiders generally show marked morphological homogeneity (i.e., a reduced number of relevant taxonomic characteristics) and are typically associated with a sedentary lifestyle, limited dispersal capabilities, narrow geographic distribution, and high levels of environmental specificity [30,41–45]. These characteristics suggest that trapdoor spiders are excellent models for morphometric studies [30,39–41]. Here, through a broad analysis based on linear and geometric morphometric methods, we investigated the patterns of geographic variation of the trapdoor spiders of the species *Idiops pirassununguensis* Fukami and Lucas, 2008 (Idiopidae). The sampled populations are from different biomes in Brazil, including a tropical rainforest (Amazon), a savanna (Cerrado), and a seasonally dry tropical forest (Caatinga) [46].

Considering the species' wide distribution, which is unusual among trapdoor spiders, the presence of records of the target species in significantly different biomes creates an uncertain expectation regarding its interpopulation differences in body size. The size of adult spiders might be smaller in warmer biomes (i.e., Caatinga), following the temperature-size rule [47–49] or the starvation resistance hypothesis [50]. On the other hand, the water conservation hypothesis suggests that higher desiccation rates might be observed in smaller individuals [51,52], resulting in the expectation of larger spiders in warmer biomes. Therefore, we tested these hypotheses by morphometrically comparing the differences among the specimens of *I. pirassununguensis* from the Amazon, Caatinga, and Cerrado biomes, a wet–dry cline.

## 2. Materials and Methods

For the morphometric analysis, we selected 64 specimens of *I. pirassununguensis*, previously identified during the review of neotropical species of the genus *Idiops* [46]. Only males were used in the analysis, as they represent more than 90% of the material available

for examination. As such, the number of available females would not be sufficient for the statistical analysis. Additionally, some specimens were damaged owing to the burning of the Butantan Institute's arachnid collection (see Kumar, 2010 [53]) and were thus excluded from the present study. Specimens from 24 of the 34 localities with records for the species from Brazil were included (Table S1), covering its entire distribution range: the Amazon ( $n$ : 20), Caatinga ( $n$ : 18) and Cerrado ( $n$ : 26) biomes (Figure 1). The definition of the biomes of the localities with records for the species was based on the official Brazilian biomes' limits, provided by the Instituto Brasileiro de Geografia e Estatística [54]. To perform the morphometric analysis, the specimens were photographed and measured using a Leica M205C stereomicroscope equipped with a Leica DFC295 digital camera.

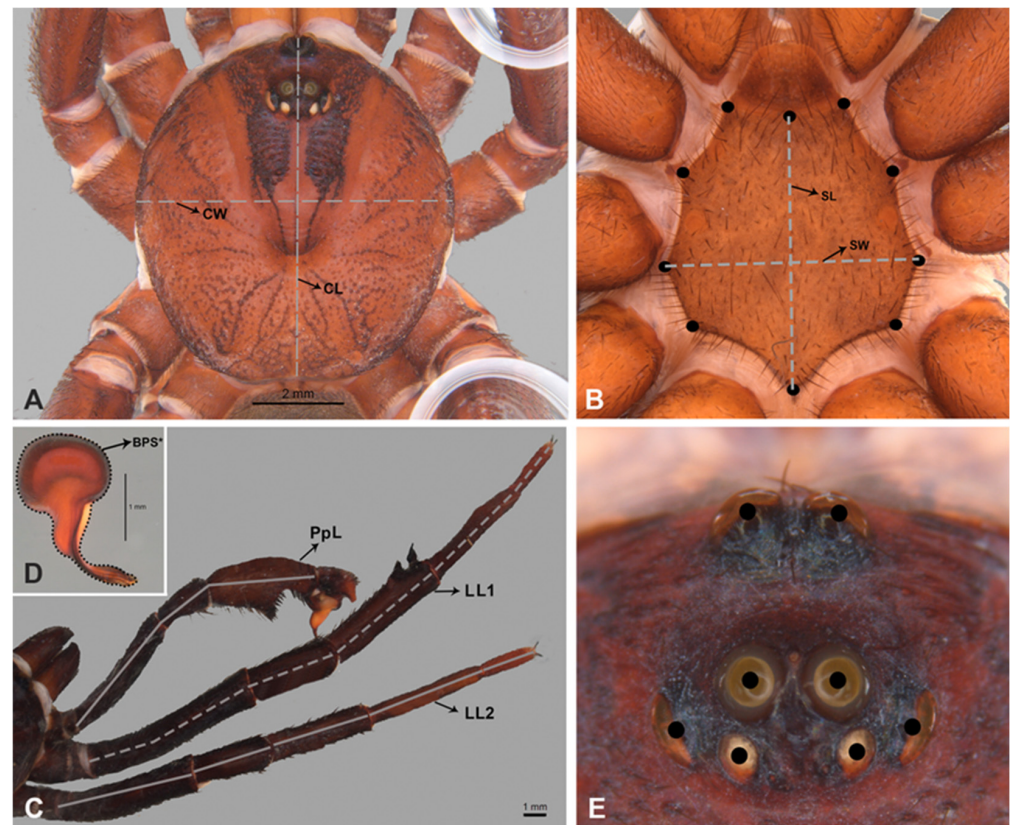


**Figure 1.** Known geographic distribution of *I. pirassununguensis*, highlighted by Brazilian biomes, categorized by specimens used (circles) and not used (triangles) in the statistical analyses.

The material examined is deposited in the following Brazilian zoological collections: Coleção Aracnológica Diamantina, Rio Claro, São Paulo (CAD; curator J. P. L. Guadanucci); Coleção de História Natural, Universidade Federal do Piauí, Florianó (CHNUFPI; curator J.F. Vilela); Instituto Butantan, São Paulo (IBSP; curator A.D. Brescovit); Instituto Nacional de Pesquisas da Amazônia, Manaus (INPA; curator M. L. Oliveira); Museu de Ciências e Tecnologia, Pontifícia Universidade Católica, Porto Alegre (MCTP; curator R. Teixeira); Museu Paraense Emílio Goeldi, Belém (MPEG; curator A.B. Bonaldo); Museu de Zoologia, Universidade de São Paulo, São Paulo (MZSP; curator R. Pinto-da-Rocha); Centro de Coleções Taxonômicas, Universidade Federal de Minas Gerais, Belo Horizonte (UFMG; curator A.J. Santos); and Coleção de Arachnida, Universidade Federal do Mato Grosso, Cuiabá (UFMT; A. Chagas Junior).

### 2.1. Definition and Measurement of Morphometric Variables

To explore the variation in the linear structures in relation to each biome, nine measurements of different body parts were taken: the carapace length (CL), carapace width (CW), sternum length (SL), sternum width (SW), leg 1 length (LL1), leg 2 length (LL2), leg 3 length (LL3), leg 4 length (LL4), and the pedipalp length (PpL) (Figure 2A–C). The definition of the linear measurements used was based on the structures that showed morphological variation among different groups of spiders in previous morphometric studies [28,35,36].



**Figure 2.** Morphometric variables of *I. pirassununguensis*. (A) Carapace length and width. (B) Length and width of the sternum; sternum landmarks. (C) Length of legs and pedipalp (legs 3 and 4 not visible in the image). (D) Shape and representation of the contour of the copulatory bulb. (E) Eye landmarks.

In addition to linear measurements, to detect geometrically significant variations of specimens in relation to biomes, geometric morphometric analysis was performed based on images of the dorsal copulatory bulb (Figure 2D), the sternum (Figure 2B), and the eye arrangement (Figure 2E). These structures were analyzed according to their biological characteristics: contours for the copulatory bulb and landmarks for the sternum and eye arrangement.

### 2.2. Linear Morphometric Analysis

First, the mean, the standard deviation, and the minimum and maximum values were generated for each linear measurement obtained. To perform the morphometric analysis of the linear measurements, seven relative measurements were generated from the ratio between the length and width of the carapace and sternum and between the length of each of the legs and the pedipalp by the width of the carapace. These ratios were used because they better characterize the proportional morphological variation among different groups, effectively demonstrating the adaptation of the specimens to different environments [35].

Then, the relative variables were subjected to a series of multivariate analyses. To explore and visualize possible patterns of similarity among the specimens, the relative vari-



ables were subjected to a principal coordinate analysis (PCoA) calculated with the Gower coefficient of similarity. Finally, to investigate whether there is structuring of the specimens in relation to the biomes, two complementary analyses were carried out. First, a canonical variate analysis (CVA) was performed to visualize the structuring patterns of the specimens among the biomes (i.e., Amazon, Caatinga and Cerrado). This analysis maximizes the variation among the previously defined groups by comparing the samples classified a priori and calculating the probability of individuals belonging to a different group [55]. Then, a permutational multivariate analysis of variance (PERMANOVA) was used to test whether there are significant differences among the analyzed groups [56]. The PERMANOVA was based on the Gower similarity index, with 10,000 permutations. *p*-values were corrected for pairwise comparisons using Bonferroni's correction; *p*-values < 0.01 were considered statistically significant. All multivariate analyses of linear measurements were conducted using PAST (Paleontological Statistics) version 4.03 [57].

### 2.3. Geometric Morphometric Analysis

#### 2.3.1. Geometric Morphometric Analysis Based on Outlines

To investigate the variation in the shape of the copulatory bulb (Figure 2D), which has a curvilinear shape that is difficult to replicate using landmarks, Fourier elliptic functions were used. These analyses describe and characterize the outlines of the analyzed structures in a quantifiable way [58]. To perform the analysis, initially, the images of the copulatory bulbs were converted into silhouettes, using Photoshop CC 2018 software. The following analyses were performed on the RStudio platform (R software v. 4.2).

First, using the R package Momocs v. 1.3.0, a series of procedures necessary to perform the elliptic Fourier analysis was performed [59]. For this, the contours of the copulatory bulb images were extracted and smoothed. Then, from the function 'calibrate\_harmonicpower\_efourier', a series of harmonics were generated; these are functions resulting from the accumulation of sine and cosine functions, detecting an increasing form of outline details. These harmonics predict the number of contours needed to achieve 95% to 99% of the shape variation, which is called Fourier power. With the number of harmonics defined, the Fourier elliptics were analyzed using the 'eFourier' function, which normalizes all images with respect to rotation, size, and orientation [59].

The generated coefficients were submitted to principal component analysis (PCA), to visualize the distribution of the copulatory bulb outlines in morphospace (the R base 'PCA' function) and characterize their variation, based on the values of the first two principal components (R base 'PCcontrib' function). Finally, as a way of visualizing the variation in the shape of the copulatory bulb in relation to the biome and calculating the hit rate in relation to the groups defined a priori, a CVA was performed using the R package MASS ('lda' function).

#### 2.3.2. Geometric Morphometric Analysis Based in Landmarks

To investigate the variation in the shape of the sternum (Figure 2B) and eye arrangements (Figure 2E), which present well-defined geometric structures and allow for the insertion of replicable reference points, morphometric analyses based on landmarks were performed.

First, image libraries were built using the TPSUtil software. Then, the landmarks were inserted, with the help of the TPSDig2 v. 1.31 [60]. Ten landmarks were defined for the sternum (Figure 2B) and eight landmarks for the ocular arrangement, referring to the central position of each eye (Figure 2E).

The following morphometric and statistical analysis were performed using MorphoJ v. 1.06 [61]. First, information regarding shape was extracted through generalized procrustes analysis (GPA). This analysis removes information related to size, orientation, and position, keeping only information regarding the shape (i.e., the Procrustes coordinates); it standardizes each specimen to a single unit centroid size [8,59]. Then, for each structure, a PCA was performed, based on the covariance matrix of the Procrustes forms, followed by a CVA. The significance of the CVA was evaluated through the values of the Procrustes and Maha-

lanobis distances, as well as the respective  $p$ -values, with 10,000 permutation replicates;  $p$  values  $< 0.01$  were considered statistically significant. Finally, to investigate the influence of size on shape (the allometric effect) and the size variation, a multivariate regression was performed between the centroid size (the independent variable) and the Procrustes coordinates (the dependent variable), followed by a permutation test of 10,000 interactions.

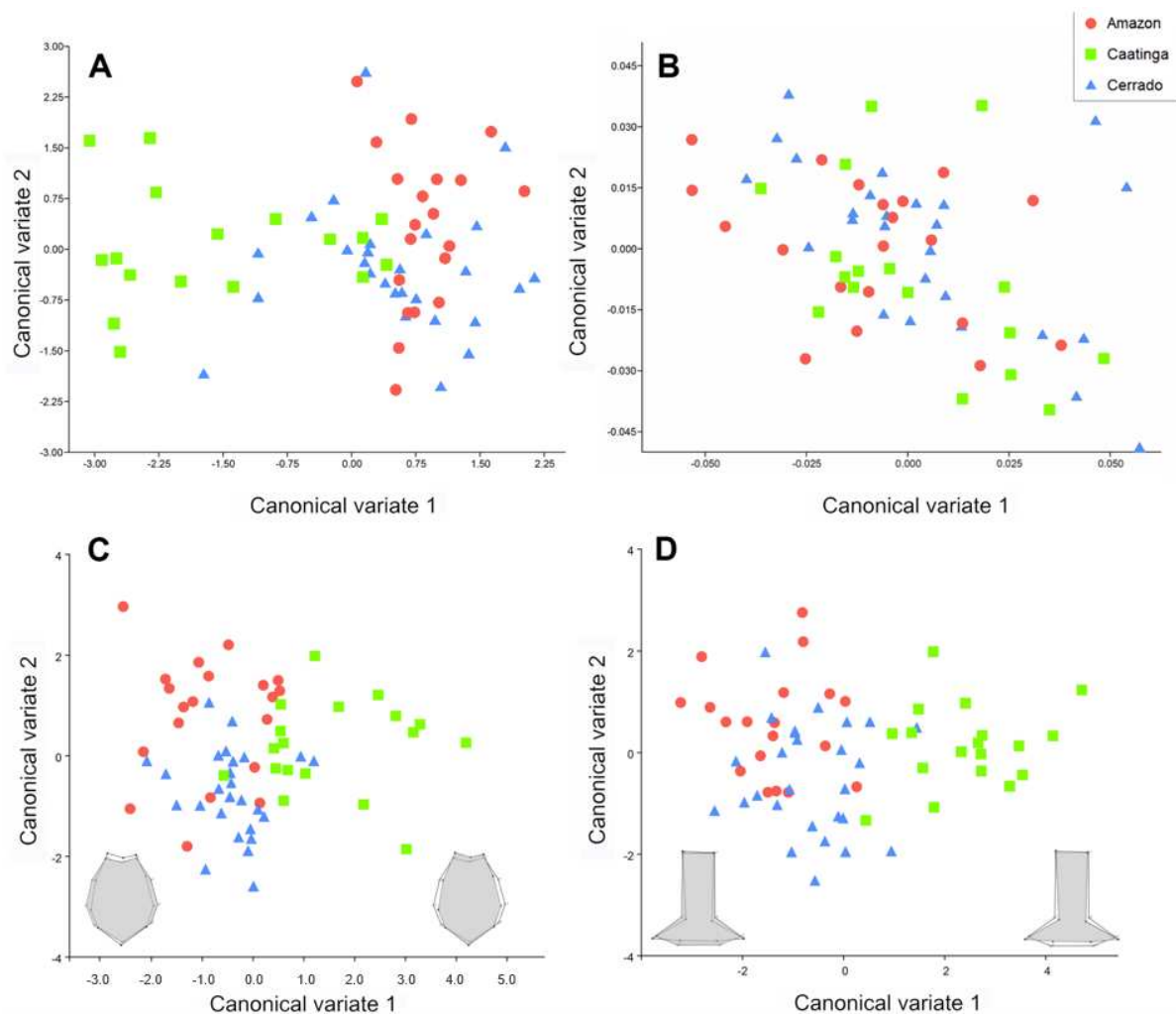
### 3. Results

#### 3.1. Linear Morphometry

The total variation among the specimens for each linear morphometric variable and the variation among biomes are presented in Table 1. In relation to the multivariate analysis of traditional relative measures, the PCoA resulted in seven axes, of which the first two described 59.8% of the variation (Axis 1: 50.1%, Axis 2: 9.7%; Figure S1, Table S2A). Regarding the CVA, the scatter plot (Figure 3A) also shows the differentiation of the Caatinga specimens, which are separated from the others and distributed along the left portion of the graph, while the Amazon and Cerrado specimens are superimposed in the right portion of the graph. The differentiation among the groups was confirmed by the general PERMANOVA ( $F = 11.55$ ,  $p < 0.01$ ). The separation of the Caatinga specimens in relation to specimens from other biomes was evidenced by a pairwise PERMANOVA, both between the Caatinga specimens and the Amazon specimens ( $F = 14.74$ ,  $p < 0.01$ ) and between the Caatinga specimens and the Cerrado specimens ( $F = 14.6$ ,  $p < 0.01$ ). There was no significant difference between specimens from the Amazon and those from Cerrado (Table 2a).

**Table 1.** Mean, standard deviation, and minimum and maximum values for each linear measurements of *I. pirassununguensis*, according to the biome and the total variation.

Measurements	Amazon (n: 20)	Caatinga (n: 18)	Cerrado (n: 26)	Total (n: 64)
Carapace length	6.96 ± 0.14 (5.53–8.04)	7 ± 0.11 (5.94–8.12)	7.08 ± 0.12 (5.64–8.2)	7 ± 0.07 (5.53–8.2)
Carapace width	6.51 ± 0.16 (4.97–7.56)	6.5 ± 0.11 (5.39–7.31)	6.65 ± 0.16 (5.07–8)	6.56 ± 0.08 (4.97–8)
Sternum length	4 ± 0.09 (3.10–4.60)	4.09 ± 0.07 (3.35–4.73)	4.02 ± 0.07 (3.45–4.8)	4.07 ± 0.04 (3.10–4.8)
Sternum width	3.68 ± 0.09 (2.66–4.34)	3.53 ± 0.07 (2.82–4.09)	3.62 ± 0.07 (2.94–4.4)	3.61 ± 0.05 (2.66–4.4)
Leg 1 length	23.4 ± 0.58 (18.36–27.1)	23 ± 0.27 (21.17–25.63)	23.52 ± 0.5 (17.89–27.08)	23.34 ± 0.28 (17.89–27.08)
Leg 2 length	20 ± 0.5 (15.51–23.13)	17.5 ± 0.6 (13.92–21.54)	20 ± 0.43 (14.48–23.1)	19.31 ± 0.3 (13.92–23.13)
Leg 3 length	17.6 ± 0.46 (12.68–20.6)	15.37 ± 0.58 (12.23–20.46)	17.47 ± 0.36 (13.32–20.45)	16.93 ± 0.28 (12.23–20.6)
Leg 4 length	23.95 ± 0.62 (18.26–27.96)	20.56 ± 0.72 (16.62–26.84)	23.67 ± 0.5 (18.15–28.15)	22.88 ± 0.38 (16.62–28.15)
Pedipalp length	12.96 ± 0.27 (10.25–14.9)	11.08 ± 0.42 (9.01–14.85)	12.7 ± 0.25 (10.48–15.16)	12.33 ± 0.2 (9.01–15.16)



**Figure 3.** Canonical variate analysis (CVA) scatterplots of different morphometric variables of *I. pirasununguensis* in relation to the biome of occurrence. (A) Linear measurements. (B) Bulb copulatory. (C) Sternum shape (transformation grids at the bottom of the scatterplot). (D) Eye arrangement (transformation grids at the bottom of the scatterplot). The different biomes are represented by the following symbols and colors: Amazon (red dots), Caatinga (green squares), Cerrado (blue triangles).

### 3.2. Geometric Morphometry

#### 3.2.1. Copulatory Bulb

The Fourier elliptic analysis required seven harmonics, which explained 99% of the variation in the dorsal shape of the copulatory bulb. The PCA resulted in 28 PC axes, of which the first 5 contributed 90.6% of the variation (Table S2B). The first two PCs contributed 75.8% of the total shape variation (PC1: 52.6%, PC2: 23.2%; Figure S2), with the variation in the first principal component (PC1) mainly related to the tegulum width and the variation in the second principal component (PC2) mainly related to the angulation of the embolus.

In relation to the CVA (Figure 3B), the scatter plot does not indicate a clear separation of specimens related to biome, as the plots belonging to the three biomes are mostly overlapping. The average hit rate per biome was high (>70%), and only the Cerrado values showed relevant percentages of specimens being incorrectly hit (22% for Caatinga; 25% for the Amazon; see Table 2b).



**Table 2.** Statistical results of the discriminant analysis in relation to the biomes of occurrence of *I. pirassununguensis*. **(a)** Linear measurements (pairwise PERMANOVA). **(b)** Bulb copulatory (contingency table). **(c)** Sternum (Mahalanobis distance). **(d)** Sternum (Procrustes distance). **(e)** Eye arrangement (Mahalanobis distance). **(f)** Eye arrangement (Procrustes distance). For A, the values on the bottom of the diagonal represent F values while those on the upper represent *p* values. For C and E, the Mahalanobis distance values are on the bottom of the diagonal and their respective *p*-values are on the upper part. The same goes for D and F in relation to the values of the Procrustes distances. Significant values (*p*-value < 0.01) were highlighted with an asterisk.

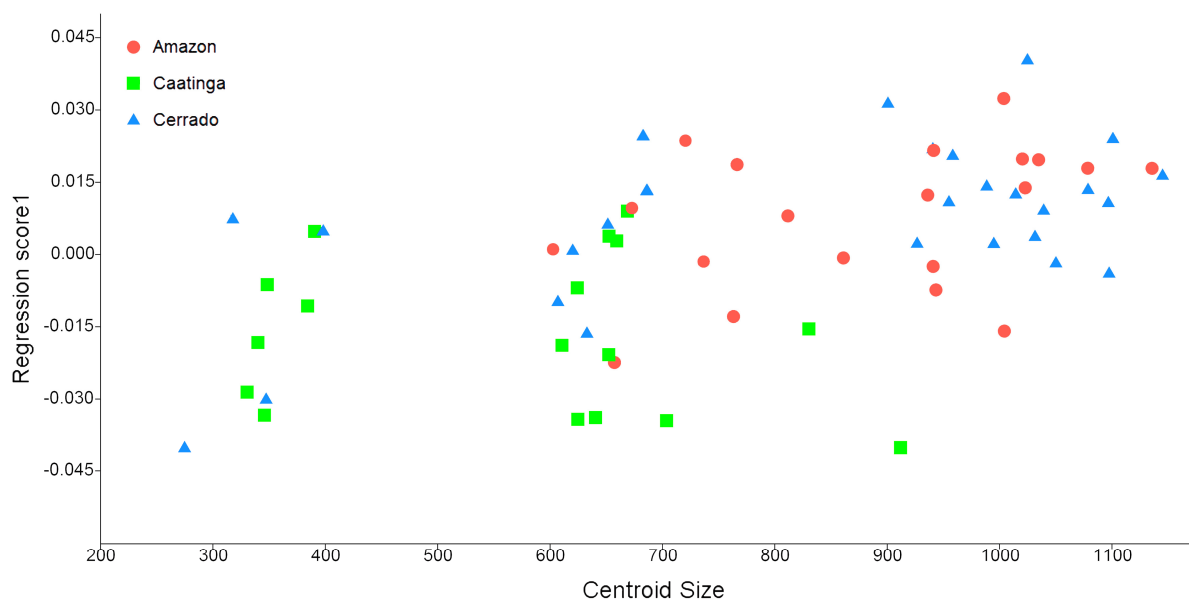
<b>a. Linear measurements (PERMANOVA)</b>				<b>b. Bulb copulatory (contingency table)</b>			
Biome	Amazon	Caatinga	Cerrado	Biome	Amazon	Caatinga	Cerrado
Amazon	-	<0.01	0.914	Amazon	65%	10%	25%
Caatinga	14.74 *	-	<0.01	Caatinga	0%	78%	22%
Cerrado	1.16	14.16 *	-	Cerrado	11.50%	11.50%	77%
<b>c. Sternum (Mahalanobis distance)</b>				<b>d. Sternum (Procrustes distance)</b>			
Biome	Amazon	Caatinga	Cerrado	Biome	Amazon	Caatinga	Cerrado
Amazon	-	<0.01	0.0135	Amazon	-	<0.01	0.583
Caatinga	2.49 *	-	<0.01	Caatinga	0.03 *	-	<0.01
Cerrado	1.56	2.2 *	-	Cerrado	0	0.03 *	-
<b>e. Eye arrangement (Mahalanobis distance)</b>				<b>f. Eye arrangement (Procrustes distance)</b>			
Biome	Amazon	Caatinga	Cerrado	Biome	Amazon	Caatinga	Cerrado
Amazon	-	<0.01	0.0912	Amazon	-	<0.01	0.324
Caatinga	3.86 *	-	<0.01	Caatinga	0.03 *	-	<0.01
Cerrado	1.3	3.17 *	-	Cerrado	0.02	0.03 *	-

### 3.2.2. Sternum

The PCA resulted in 16 PC axes, of which the first 5 contributed 76.8% of the sternum shape variation (Table S2C). The first two PCs contributed 54.5% of the total shape variation (PC1: 44.4%, PC2: 10.1%; Figure S3), with the variation in the first principal component (PC1) mainly related to sternum width and length and the variation in the second principal component (PC2) related to the shape of the anterior portion of the sternum.

According to the CVA scatter plot (Figure 3C), specimens from the Caatinga group differ from those from the Amazon and Cerrado, which are more similar to each other. CV1 explained 70.9% of the variation, which was related to a greater narrowing and elongation of the sternum in individuals from the Caatinga group compared to the others. When compared with the Amazon and Cerrado groups, the Caatinga specimens showed significantly higher values of Procrustes distances and Mahalanobis distances (Table 2c,d), which evidence the distinction between these groups.

The multiple regression showed that, despite the allometry effect having a significant permutation value ( $p < 0.01$ ), size contributed only 8.8% of the shape variation. Specimens from the Caatinga area tend to be smaller than their conspecifics from the other analyzed biomes (Figure 4).



**Figure 4.** Multivariate regression of sternum shape on the sternum centroid size of *I. pirassununguensis*. The different biomes are represented by the following symbols and colors: Amazon (red dots), Caatinga (green squares), Cerrado (blue triangles).

### 3.2.3. Eye Arrangement

Regarding the shape of the eye arrangement, PCA resulted in 12 PC axes, of which the first 5 contributed to 80.3% of the shape variation (Table S2D). The first two PCs contributed 51.6% of the total shape variation (PC1: 28.7%, PC2: 22.9%; Figure S4). The first principal component (PC1) is mainly related to the length of the eye tubercle and the distance of the eye tubercle in relation to the anterior lateral eyes. The second principal component (PC2) explains the part of the variation related to the arrangement of the anterior median eyes and the posterior lateral eyes.

According to the CVA results (Figure 3D), the Caatinga specimens are separated from those from the Amazon and the Cerrado, as observed for the sternum shape (Figure 3C). These differences are statistically significant ( $p < 0.01$ ) for the Mahalanobis and Procrustes distances (Table 2d,e). CV1 explained 92.1% of the variation, which was related to a greater narrowing of the eye tubercle and the distance between the anterior lateral eyes of Caatinga individuals when compared to specimens from the other biomes.

Although the multiple regression analysis indicates that 3% of the size variation is related to the shape, this analysis did not provide significant support in the permutation test ( $p$ -value = 0.06).

## 4. Discussion

This study constitutes the first morphometric investigation of an Idiopidae trapdoor spider species. The results of the analysis, which was based on the traditional morphometry of linear measurements and the geometric morphometry of the sternum and the eye arrangement, were congruent and complementary. The specimens from the Caatinga are morphologically distinct from their conspecifics from the Amazon and the Cerrado, which showed no significant variation from each other. Regarding the results of the morphogeometric analysis of the copulatory bulb, there was no separation of the specimens in relation to the biomes.

Individuals from the Caatinga tend to be smaller when compared to specimens from the Amazon and Cerrado. Those specimens shared a narrower and more elongated sternum, shorter legs and pedipalps, a narrower eye tubercle, and anterior lateral eyes that were furthest from the eye tubercle. This differentiation of the Caatinga specimens supports the temperature–size rule [47–49] or the starvation resistance hypothesis [50]. First, the

temperature–size rule postulates that warmer environments harbor smaller species and/or specimens [47–49]. As such, the observed differences may be a byproduct of morphological and physiological adaptations related to the thermoregulation and water balance of spiders due to the restrictions imposed by the arid and semi-arid environment [62,63]. This would be congruent with the Caatinga climatic conditions. This biome is a nucleus of the South American seasonally dry tropical forests (SDTF) [64], being characterized by low levels of annual precipitation, high seasonality (precipitation concentrated in a few consecutive months), high average annual temperatures, low annual thermal amplitude, and low relative humidity [65,66]. The second hypothesis (i.e., starvation resistance) relates to ecological limitations, which are associated with the lower availability of food resources, water resources, and shelter, common in arid and open environments affected by drought; such circumstances are typical of the Caatinga [65,67]. In any case, it is not possible to make a distinction between these two hypotheses with the currently available data.

Our results corroborate those of a recent study of Brazilian scorpions from a temperature cline zone (i.e., Caatinga and Atlantic Forest sites), in which high temperatures showed a significantly negative correlation with body size [49]. In an intraspecific context, Lira et al. [48] suggested that the reduction in the size of the specimens from Caatinga may be related to the molting process and the physiological restrictions imposed by it. Similarly, specimens of the trunk-dwelling lizard *Gonatodes humeralis* (Guichenot, 1855) from the Caatinga exhibited smaller body sizes when compared to Amazonian populations [68]. This variation was interpreted as a result of both abiotic (e.g., temperature and humidity; the so-called temperature–size rule), and biotic (e.g., the availability of food resources; the so-called starvation hypothesis) factors, and their interactions [68]. Finally, laboratory-raised *Acheta domesticus* (Linnaeus, 1758) crickets that were subjected lower hydration conditions were significantly smaller than those submitted to high hydration conditions [69].

However, the smaller body size in arid conditions is not a rule of thumb. For example, gymnophthalmid lizards that inhabit more arid regions are generally more elongated [70]. This result was generally thought to be a byproduct of the association between the habitat structure, lizard behavior, environmental conditions, and lower resource availability [70]. Although the target species of our study is a trapdoor spider, which does not build orb webs, body elongation may be related to heat transfer, since males are wanderers and spend part of their lives outside of burrows looking for females, occasionally being exposed to the sun. Because the environments of the Caatinga biome are characterized by open, seasonally dry, and deciduous tropical phytophysognomies, this might have resulted in specimens with more elongated bodies, as observed in the sternums of specimens from the Caatinga, possibly reducing desiccation.

Some of these morphological variations, which are mainly associated with arid and semi-arid environments, may also be related to the evolutionary and biogeographic history of *I. pirassununguensis* [45]. Phylogeographic studies of different taxa from the Caatinga showed the genetic structuring associated with the Caatinga, the Cerrado, and the Chaco [5,71], biomes that are biogeographically and climatically related. Studies of spiders [72], lizards [73,74], amphibians [75], and birds [76,77] found similar patterns of diversification and structuring, mainly related to the Pliocene and Pleistocene periods, showing genetic lineages with occurrence restricted to the Caatinga.

The current distribution of *I. pirassununguensis* over a large geographic area shows an unusual pattern that is distinct from that observed for other trapdoor spider species, which, in general, have a restricted range and strong geographic and genetic structuring, as observed, for example, in spiders from the families Euctenizidae [30,41,78], Antrodiaetidae [42,79,80], Migidae [43,81,82], Halonoproctidae [83], and even Idiopidae [84]. It is remarkable that, even though *I. pirassununguensis* has a wide distribution across different biomes and presents significant variation in relation to the size and shape of non-sexual structures, the specimens share a series of diagnostic sexual characteristics for the species, related to the copulatory bulb, pedipalp, and tibial apophysis [46]. This finding supports

the hypothesis that, based on various species of insects and spiders [85], male genitalia show less variation than other body parts, indicating a case of intraspecific variation.

The use of different morphometric methods herein demonstrates the importance of combined analysis in quantifying variation and identifying morphologically distinct groups. Our results indicate distinctions in the size and shape of the Caatinga specimens when compared to their conspecifics from the Amazon and Cerrado. These results provide support for future investigations involving an integrative taxonomy using time-calibrated molecular data and species distribution modeling. This approach can lead to a deeper understanding of the diversification patterns of *I. pirassununguensis* across Brazilian biomes and yield insights into possible adaptive processes associated with the evolution of these spiders in semi-arid environments.

**Supplementary Materials:** The following supporting information can be downloaded at: <https://www.mdpi.com/article/10.3390/d15070861/s1>, Figure S1: Linear measurements PCoA; Figure S2: Bulb copulatory PCA; Figure S3: Sternum PCA; Figure S4: Eye arrangement PCA; Table S1: Sample information; Table S2: Table S2: Variance percentage PCoA-PCA.

**Author Contributions:** Conceptualization, R.F.-F., M.J.A.M. and J.P.L.G.; methodology, R.F.-F. and M.J.A.M.; writing—original draft preparation, R.F.-F.; writing—review and editing, R.F.-F., M.J.A.M., L.S.C. and J.P.L.G.; visualization, R.F.-F.; supervision, M.J.A.M. and J.P.L.G.; funding acquisition, R.F.-F. and J.P.L.G. All authors have read and agreed to the published version of the manuscript.

**Funding:** This research was funded by Fundação de Amparo à Pesquisa do Estado de São Paulo, grant number FAPESP 2018/02842-2 to R.F.F. and grant number FAPESP 2017/11985-9 to J.P.L.G.

**Institutional Review Board Statement:** Not applicable.

**Data Availability Statement:** All results generated in this study are included in this article and in supplementary material files.

**Acknowledgments:** We wish to thank all the curators for the loan of specimens. We also thank Ana Lúcia Tourinho, Arthur Galleti Lima, Facundo Labarque, Felipe Garcia, Guilherme H. F. Azevedo, Juliano Zardetto, Milton Ribeiro, Pedro H. Martins, Rafael P. Indicatti, and Tadeu Siqueira for their help with and critical suggestions for this work. We thank the editor Vera Opatova and the anonymous reviewers for their constructive criticism.

**Conflicts of Interest:** The authors declare no conflict of interest.

## References

1. Mayr, E. *Populations, Species, and Evolution: An Abridgment of Animal Species and Evolution*; Harvard University Press: Cambridge, MA, USA, 1970; Volume 19.
2. Gould, S.J.; Johnston, R.F. Geographic variation. *Annu. Rev. Ecol. Syst.* **1972**, *3*, 457–498. [\[CrossRef\]](#)
3. Endler, J.A. *Geographic Variation, Speciation and Clines*; MPB-10; Princeton University Press: Princeton, NJ, USA, 1977; Volume 10.
4. Chown, S.L.; Gaston, K.J. Body size variation in insects: A macroecological perspective. *Biol. Rev.* **2010**, *85*, 139–169. [\[CrossRef\]](#)
5. Werneck, F.P. The diversification of eastern South American open vegetation biomes: Historical biogeography and perspectives. *Quat. Sci. Rev.* **2011**, *30*, 1630–1648. [\[CrossRef\]](#)
6. Rull, V.; Carnaval, A.C. (Eds.) *Neotropical Diversification: Patterns and Processes*; Springer: Berlin/Heidelberg, Germany, 2020.
7. Reyment, R.A. Morphometrics: An historical essay. In *Morphometrics for Nonmorphometricians*; Elewa, A., Ed.; Springer: Berlin/Heidelberg, Germany, 2010; pp. 9–24. [\[CrossRef\]](#)
8. Zelditch, M.L.; Swiderski, D.L.; Sheets, H.D. *Geometric Morphometrics for Biologists: A Primer*; Academic Press: Cambridge, MA, USA, 2012.
9. Klingenberg, C.P. Evolution and development of shape: Integrating quantitative approaches. *Nat. Rev. Genet.* **2010**, *11*, 623–635. [\[CrossRef\]](#)
10. Zúniga-Reinoso, Á.; Benítez, H.A. The overrated use of the morphological cryptic species concept: An example with *Nyctelia* darkbeetles (Coleoptera: Tenebrionidae) using geometric morphometrics. *Zool. Anz.—J. Comp. Zool.* **2015**, *255*, 47–53. [\[CrossRef\]](#)
11. Struck, T.H.; Feder, J.L.; Bendiksy, M.; Birkeland, S.; Cerca, J.; Gusarov, V.I.; Kistenich, S.; Larsson, K.H.; Liow, L.H.; Nowak, M.D.; et al. Finding evolutionary processes hidden in cryptic species. *Trends Ecol. Evol.* **2018**, *33*, 153–163. [\[CrossRef\]](#)
12. Recoder, R.S.; Ribeiro, M.C.; Rodrigues, M.T. Spatial variation in morphometry in *Vanzosaura rubricauda* (Squamata, Gymnophthalmidae) from open habitats of South America and its environmental correlates. *S. Am. J. Herpetol.* **2013**, *8*, 186–197. [\[CrossRef\]](#)

13. Álvarez-Varas, R.; Véliz, D.; Vélez-Rubio, G.M.; Fallabrino, A.; Zárate, P.; Heidemeyer, M.; Godoy, D.A.; Benítez, H.A. Identifying genetic lineages through shape: An example in a cosmopolitan marine turtle species using geometric morphometrics. *PLoS ONE* **2019**, *14*, e0223587. [[CrossRef](#)] [[PubMed](#)]
14. Vilaseca, C.; Méndez, M.A.; Pinto, C.F.; Lemic, D.; Benítez, H.A. Unraveling the morphological variation of *Triatoma infestans* in the peridomestic habitats of Chuquisaca Bolivia: A Geometric Morphometric Approach. *Insects* **2021**, *12*, 185. [[CrossRef](#)]
15. Nattero, J.; Piccinali, R.V.; Macedo Lopes, C.; Hernández, M.L.; Abrahan, L.; Lobbia, P.A.; Rodríguez, C.S.; Carbajal de la Fuente, A.L. Morphometric variability among the species of the *Sordida* subcomplex (Hemiptera: Reduviidae: Triatominae): Evidence for differentiation across the distribution range of *Triatoma sordida*. *Parasites Vectors* **2017**, *10*, 4124. [[CrossRef](#)]
16. Smith, C.H.; Johnson, N.A.; Inoue, K.; Doyle, R.D.; Randklev, C.R. Integrative taxonomy reveals a new species of freshwater mussel, *Potamilus streckersoni* sp. nov. (Bivalvia: Unionidae): Implications for conservation and management. *Syst. Biodivers.* **2019**, *17*, 331–348. [[CrossRef](#)]
17. Aglagane, A.; Tofilski, A.; Er-Rguibi, O.; Laghzaoui, E.M.; Kimdil, L.; El Mouden, E.H.; Fuchs, S.; Oleksa, A.; Aamiri, A.; Aourir, M. Geographical variation of honey bee (*Apis mellifera* L. 1758) populations in South-Eastern Morocco: A geometric morphometric analysis. *Insects* **2022**, *13*, 288. [[CrossRef](#)]
18. Ferreira, V.S.; Aguiar CM, L.; Costa, M.A.; Silva, J.G. Morphometric analysis of populations of *Centris aenea lepeletier* (Hymenoptera: Apidae) from Northeastern Brazil. *Neotrop. Entomol.* **2011**, *40*, 97–102. [[CrossRef](#)]
19. da Silva AC, S.; Nunes, L.A.; de Lima Batista, W.; Lhano, M.G. Morphometric variation among males of *Orphulella punctata* (De Geer, 1773)(Acrididae: Gomphocerinae) from different biomes in Brazil. *J. Orthoptera Res.* **2018**, *27*, 163–171. [[CrossRef](#)]
20. Regueira, J.C.; Damasceno, E.M.; Iannuzzi, L. Shape variation of *Cydianerus latruncularius* (Coleoptera, Curculionidae) across biomes and sexes. *Zool. Anz.* **2020**, *289*, 96–107. [[CrossRef](#)]
21. Cordeiro-Estrela, P.; Baylac, M.; Denys, C.; Marinho-Filho, J. Interspecific patterns of skull variation between sympatric Brazilian vesper mice: Geometric morphometrics assessment. *J. Mammal.* **2006**, *87*, 1270–1279. [[CrossRef](#)]
22. Coyle, F.A. Systematics of the trapdoor spider genus *Aliatypus* (Araneae: Antrodiaetidae). *Psyche* **1974**, *81*, 431–500. [[CrossRef](#)]
23. Coyle, F.A. A revision of the funnelweb mygalomorph spider subfamily Ischnothelinae (Araneae, Dipluridae). In *Bulletin of the American Museum of Natural History*; No. 226; American Museum of Natural History: New York, NY, USA, 1995.
24. Crews, S.C.; Hedin, M. Studies of morphological and molecular phylogenetic divergence in spiders (Araneae: Homalonychus) from the American southwest, including divergence along the Baja California Peninsula. *Mol. Phylogenet. Evol.* **2006**, *38*, 470–487. [[CrossRef](#)] [[PubMed](#)]
25. Entling, W.; Schmidt-Entling, M.H.; Bacher, S.; Brandl, R.; Nentwig, W. Body size–climate relationships of European spiders. *J. Biogeogr.* **2010**, *37*, 477–485. [[CrossRef](#)]
26. Puzin, C.; Leroy, B.; Pétilon, J. Intra-and inter-specific variation in size and habitus of two sibling spider species (Araneae: Lycosidae): Taxonomic and biogeographic insights from sampling across Europe. *Biol. J. Linn. Soc.* **2014**, *113*, 85–96. [[CrossRef](#)]
27. Krehenwinkel, H.; Graze, M.; Rödder, D.; Tanaka, K.; Baba, Y.G.; Muster, C.; Uhl, G. A phylogeographical survey of a highly dispersive spider reveals eastern Asia as a major glacial refugium for Palaearctic fauna. *J. Biogeogr.* **2016**, *43*, 1583–1594. [[CrossRef](#)]
28. Wong, M.K.; Woodman, J.D.; Rowell, D.M. Short-range phenotypic divergence among genetically distinct parapatric populations of an Australian funnel-web spider. *Ecol. Evol.* **2017**, *7*, 5094–5102. [[CrossRef](#)] [[PubMed](#)]
29. Korba, J.; Opatova, V.; Calatayud-Mascarell, A.; Enguídanos, A.; Bellvert, A.; Adrián, S.; Sánchez-Vialas, A.; Arnedo, M.A. Systematics and phylogeography of western Mediterranean tarantulas (Araneae: Theraphosidae). *Zool. J. Linn. Soc.* **2022**, *196*, 845–884. [[CrossRef](#)]
30. Bond, J.E.; Stockman, A.K. An integrative method for delimiting cohesion species: Finding the population-species interface in a group of Californian trapdoor spiders with extreme genetic divergence and geographic structuring. *Syst. Biol.* **2008**, *57*, 628–646. [[CrossRef](#)]
31. Costa-Schmidt, L.E.; de Araújo, A.M. Genitalic variation and taxonomic discrimination in the semi-aquatic spider genus *Paratrechalea* (Araneae: Trechaleidae). *J. Arachnol.* **2010**, *38*, 242–249. [[CrossRef](#)]
32. Fernández-Montraveta, C.; Marugan-Lobon, J. Geometric morphometrics reveals sex-differential shape allometry in a spider. *PeerJ* **2017**, *5*, e3617. [[CrossRef](#)]
33. Valdez-Mondragón, A.; Navarro-Rodríguez, C.I.; Solís-Catalán, K.P.; Cortez-Roldán, M.R.; Juárez-Sánchez, A.R. Under an integrative taxonomic approach: The description of a new species of the genus *Loxosceles* (Araneae, Sicariidae) from Mexico City. *ZooKeys* **2019**, *892*, 93. [[CrossRef](#)]
34. Wilson, J.D.; Zapata, L.V.; Barone, M.L.; Cotoras, D.D.; Poy, D.; Ramírez, M.J. Geometric morphometrics reveal sister species in sympatry and a cline in genital morphology in a ghost spider genus. *Zool. Scr.* **2021**, *50*, 485–499. [[CrossRef](#)]
35. Lapinski, W.; Walther, P.; Tschapka, M. Morphology reflects microhabitat preferences in an assemblage of neotropical wandering spiders. *Zoomorphology* **2015**, *134*, 219–236. [[CrossRef](#)]
36. Wolff, J.O.; Wierucka, K.; Paterno, G.B.; Coddington, J.A.; Hormiga, G.; Kelly, M.B.; Herberstein, M.E.; Ramírez, M.J. Stabilized morphological evolution of spiders despite mosaic changes in foraging ecology. *Syst. Biol.* **2022**, *71*, 1487–1503. [[CrossRef](#)]
37. Kallal, R.J.; Moore, A.J.; Hormiga, G. The shape of weaver: Investigating shape disparity in orb-weaving spiders (Araneae, Araneidae) using geometric morphometrics. *Evol. Biol.* **2019**, *46*, 317–331. [[CrossRef](#)]
38. Bond, J.E.; Beamer, D.A. A morphometric analysis of mygalomorph spider carapace shape and its efficacy as a phylogenetic character (Araneae). *Invertebr. Syst.* **2006**, *20*, 1–7. [[CrossRef](#)]



39. Brandt, S.; Sole, C.; Lyle, R.; Pirk, C. Geometric Morphometric Analysis of Ocular Patterns as a Species Identifier in the South African Endemic Trapdoor Spider Genus *Stasimopus* Simon, 1892 (Araneae, Mygalomorphae, Stasimopidae). *Evol. Biol.* **2023**, 1–15. [\[CrossRef\]](#)
40. Ríos-Tamayo, D. Variación intra-específica en machos de *Actinopus* sp. (Mygalomorphae: Actinopodidae) del norte argentino. *Acta Zoológica Lilloana* **2016**, 60, 78–88.
41. Bond, J.E.; Hedin, M.C.; Ramirez, M.G.; Opell, B.D. Deep molecular divergence in the absence of morphological and ecological change in the Californian coastal dune endemic trapdoor spider *Aptostichus simus*. *Mol. Ecol.* **2001**, 10, 899–910. [\[CrossRef\]](#)
42. Hendrixson, B.E.; Bond, J.E. Testing species boundaries in the *Antrodiaetus unicolor* complex (Araneae: Mygalomorphae: Antrodiaetidae): “paraphyly” and cryptic diversity. *Mol. Phylogenet. Evol.* **2005**, 36, 405–416. [\[CrossRef\]](#)
43. Cooper, S.J.; Harvey, M.S.; Saint, K.M.; Main, B.Y. Deep phylogeographic structuring of populations of the trapdoor spider *Moggridgea tingle* (Migidae) from southwestern Australia: Evidence for long-term refugia within refugia. *Mol. Ecol.* **2011**, 20, 3219–3236. [\[CrossRef\]](#) [\[PubMed\]](#)
44. Mason, L.D.; Wardell-Johnson, G.; Main, B.Y. The longest-lived spider: Mygalomorphs dig deep, and persevere. *Pac. Conserv. Biol.* **2018**, 24, 203–206. [\[CrossRef\]](#)
45. Opatova, V.; Hamilton, C.A.; Hedin, M.; De Oca, L.M.; Král, J.; Bond, J.E. Phylogenetic systematics and evolution of the spider infraorder Mygalomorphae using genomic scale data. *Syst. Biol.* **2020**, 69, 671–707. [\[CrossRef\]](#)
46. Fonseca-Ferreira, R.; Guadanucci JP, L.; Yamamoto, F.U.; Brescovit, A.D. Taxonomic revision of the Neotropical spiders of the genus *Idiops* Perty, 1833 (Araneae, Idiopidae), with description of four new species. *Eur. J. Taxon.* **2021**, 780, 1–71. [\[CrossRef\]](#)
47. Atkinson, D. Temperature and organism size: A biological law for ectotherms. *Adv. Ecol. Res.* **1994**, 25, 1–58. [\[CrossRef\]](#)
48. Horne, C.R.; Hirst, A.G.; Atkinson, D. Temperature-size responses match latitudinal-size clines in arthropods, revealing critical differences between aquatic and terrestrial species. *Ecol. Lett.* **2015**, 18, 327–335. [\[CrossRef\]](#) [\[PubMed\]](#)
49. Lira, A.F.; Foerster, S.I.; Albuquerque, C.M.; Moura, G.J. Contrasting patterns at interspecific and intraspecific levels in scorpion body size across a climatic gradient from rainforest to dryland vegetation. *Zoology* **2021**, 146, 125908. [\[CrossRef\]](#)
50. Cushman, J.H.; Lawton, J.H.; Manly BF, J. Latitudinal patterns in European ant assemblages: Variation in species richness and body size. *Oecologia* **1993**, 95, 30–37. [\[CrossRef\]](#) [\[PubMed\]](#)
51. Remmert, H. Body size of terrestrial arthropods and biomass of their populations in relation to the abiotic parameters of their milieu. *Oecologia* **1981**, 50, 12–13. [\[CrossRef\]](#)
52. Gouveia, S.F.; Correia, I. Geographical clines of body size in terrestrial amphibians: Water conservation hypothesis revisited. *J. Biogeogr.* **2016**, 43, 2075–2084. [\[CrossRef\]](#)
53. Kumar, A. A tragic loss: Fire at Instituto Butantan, Brazil. *Toxicon Off. J. Int. Soc. Toxinology* **2010**, 56, 1528–1529. [\[CrossRef\]](#)
54. IBGE. *Biomass e Sistema Costeiro-Marinho do Brasil: Compatível com a Escala 1: 250 000*; Série Relatórios Metodológicos; v. 45; IBGE: Rio de Janeiro, Brazil, 2019.
55. Strauss, R.E. Discriminating groups of organisms. In *Morphometrics for Nonmorphometricians*; Springer: Berlin/Heidelberg, Germany, 2010; pp. 73–91. [\[CrossRef\]](#)
56. Anderson, M.J. A new method for non-parametric multivariate analysis of variance. *Austral Ecol.* **2001**, 26, 32–46. [\[CrossRef\]](#)
57. Hammer, Ø.; Harper, D.A.; Ryan, P.D. PAST: Paleontological statistics software package for education and data analysis. *Palaeontol. Electron.* **2001**, 4, 9.
58. Kuhl, F.P.; Giardina, C.R. Elliptic Fourier features of a closed contour. *Comput. Graph. Image Process.* **1982**, 18, 236–258. [\[CrossRef\]](#)
59. Bonhomme, V.; Picq, S.; Gaucherel, C.; Claude, J. Momocs: Outline analysis using R. *J. Stat. Softw.* **2014**, 56, 1–24. [\[CrossRef\]](#)
60. Rohlf, F.J. tpsDig. 2001. Available online: <https://life2.bio.sunysb.edu/ee/rohlf/software.html> (accessed on 13 July 2022).
61. Klingenberg, C.P. MorphoJ: An integrated software package for geometric morphometrics. *Mol. Ecol. Resour.* **2011**, 11, 353–357. [\[CrossRef\]](#) [\[PubMed\]](#)
62. Main, B.Y. Adaptations to arid habitats by mygalomorph spiders. In *Evolution of the Flora and Fauna of Arid Australia*; Barker, W.R., Greenslade, P.J., Eds.; Australian Systematic Botany Society: Frewville, Australia, 1982.
63. Canals, M.; Veloso, C.; Solís, R. Adaptation of the spiders to the environment: The case of some Chilean species. *Front. Physiol.* **2015**, 6, 220. [\[CrossRef\]](#) [\[PubMed\]](#)
64. Werneck, F.P.; Costa, G.C.; Colli, G.R.; Prado, D.E.; Sites, J.W., Jr. Revisiting the historical distribution of Seasonally Dry Tropical Forests: New insights based on palaeodistribution modelling and palynological evidence. *Glob. Ecol. Biogeogr.* **2011**, 20, 272–288. [\[CrossRef\]](#)
65. Bucher, E.H. Chaco and Caatinga—South American arid savannas, woodlands and thickets. In *Ecology of Tropical Savannas*; Springer: Berlin/Heidelberg, Germany, 1982; pp. 48–79. [\[CrossRef\]](#)
66. Prado, D.E. As caatingas da América do Sul. *Ecol. Conserv. Caatinga* **2003**, 2, 3–74.
67. Silva, J.M.C.D.; Barbosa, L.C.F.; Leal, I.R.; Tabarelli, M. The Caatinga: Understanding the challenges. In *Caatinga*; Springer: Cham, Switzerland, 2017; pp. 3–19. [\[CrossRef\]](#)
68. de Oliveira, F.R.C.; Passos, D.C.; Borges-Nojosa, D.M. Ecology of the lizard *Gonatodes humeralis* (Sphaerodactylidae) in a coastal area of the Brazilian semi-arid: What differs from the Amazonian populations? *J. Arid. Environ.* **2021**, 190, 104506. [\[CrossRef\]](#)
69. McCluney, K.E.; Date, R.C. The effects of hydration on growth of the house cricket, *Acheta domesticus*. *J. Insect Sci.* **2008**, 8, 32. [\[CrossRef\]](#)

70. Grizante, M.B.; Brandt, R.; Kohlsdorf, T. Evolution of body elongation in gymnophthalmid lizards: Relationships with climate. *PLoS ONE* **2012**, *7*, e49772. [[CrossRef](#)]
71. Vanzolini, P.E. Problemas faunísticos do Cerrado. *Simpósio Sobre o Cerrado* **1963**, *307*, 1.
72. Magalhaes, I.L.; Oliveira, U.; Santos, F.R.; Vidigal, T.H.; Brescovit, A.D.; Santos, A.J. Strong spatial structure, Pliocene diversification and cryptic diversity in the Neotropical dry forest spider *Sicarius cariri*. *Mol. Ecol.* **2014**, *23*, 5323–5336. [[CrossRef](#)]
73. Werneck, F.P.; Gamble, T.; Colli, G.R.; Rodrigues, M.T.; Sites, J.W., Jr. Deep diversification and long-term persistence in the South American ‘dry diagonal’: Integrating continent-wide phylogeography and distribution modeling of geckos. *Evol. Int. J. Org. Evol.* **2012**, *66*, 3014–3034. [[CrossRef](#)] [[PubMed](#)]
74. Fonseca, E.M.; Gehara, M.; Werneck, F.P.; Lanna, F.M.; Colli, G.R.; Sites, J.W., Jr.; Rodrigues, M.T.; Garda, A.A. Diversification with gene flow and niche divergence in a lizard species along the South American “diagonal of open formations”. *J. Biogeogr.* **2018**, *45*, 1688–1700. [[CrossRef](#)]
75. Thome MT, C.; Carstens, B.C.; Rodrigues, M.T.; Alexandrino, J.; Haddad, C.F. Genomic data from the Brazilian sibilator frog reveal contrasting pleistocene dynamics and regionalism in two South American dry biomes. *J. Biogeogr.* **2021**, *48*, 1112–1123. [[CrossRef](#)]
76. Moura CC, D.M.; Araujo HF, D.; Aleixo, A.; Wink, M.; Fernandes, A.M. The role of landscape change and paleoclimatic events in shaping the evolutionary history of the *Poliophtila* gnatcatchers (Passeriformes, Poliophtilidae) with emphasis on species associated with open habitats. *J. Avian Biol.* **2018**, *49*, jav-012409. [[CrossRef](#)]
77. Rocha, A.V.; Cabanne, G.S.; Aleixo, A.; Silveira, L.F.; Tubaro, P.; Caparroz, R. Pleistocene climatic oscillations associated with landscape heterogeneity of the South American dry diagonal explains the phylogeographic structure of the narrow-billed woodcreeper (*Lepidocolaptes angustirostris*, Dendrocolaptidae). *J. Avian Biol.* **2020**, *51*, 1–13. [[CrossRef](#)]
78. Newton, L.G.; Starrett, J.; Jochim, E.E.; Bond, J.E. Phylogeography and cohesion species delimitation of California endemic trapdoor spiders within the *Aptostichus icenoglei* sibling species complex (Araneae: Mygalomorphae: Euctenizidae). *Ecol. Evol.* **2023**, *13*, e10025. [[CrossRef](#)]
79. Newton, L.G.; Starrett, J.; Hendrixson, B.E.; Derkarabetian, S.; Bond, J.E. Integrative species delimitation reveals cryptic diversity in the southern Appalachian *Antrodiaetus unicolor* (Araneae: Antrodiaetidae) species complex. *Mol. Ecol.* **2020**, *29*, 2269–2287. [[CrossRef](#)]
80. Satler, J.D.; Carstens, B.C.; Hedin, M. Multilocus species delimitation in a complex of morphologically conserved trapdoor spiders (Mygalomorphae, Antrodiaetidae, Aliatypus). *Syst. Biol.* **2013**, *62*, 805–823. [[CrossRef](#)]
81. Harvey, M.S.; Main, B.Y.; Rix, M.G.; Cooper, S.J. Refugia within refugia: In situ speciation and conservation of threatened *Bertmainius* (Araneae: Migidae), a new genus of relictual trapdoor spiders endemic to the mesic zone of south-western Australia. *Invertebr. Syst.* **2015**, *29*, 511–553. [[CrossRef](#)]
82. Ferretti, N.E.; Soresi, D.S.; Gonzalez, A.; Arnedo, M. An integrative approach unveils speciation within the threatened spider *Calathotarsus simoni* (Araneae: Mygalomorphae: Migidae). *Syst. Biodivers.* **2019**, *17*, 439–457. [[CrossRef](#)]
83. Opatova, V.; Bond, J.E.; Arnedo, M.A. Uncovering the role of the Western Mediterranean tectonics in shaping the diversity and distribution of the trap-door spider genus *Ummidia* (Araneae, Ctenizidae). *J. Biogeogr.* **2016**, *43*, 1955–1966. [[CrossRef](#)]
84. Opatova, V.; Arnedo, M.A. Spiders on a hot volcanic roof: Colonisation pathways and phylogeography of the Canary Islands endemic trap-door spider *Titanidiops canariensis* (Araneae, Idiopidae). *PLoS ONE* **2014**, *9*, e115078. [[CrossRef](#)] [[PubMed](#)]
85. Eberhard, W.G.; Huber, B.A.; Briceño, R.D.; Salas, I.; Rodriguez, V. One size fits all? Relationships between the size and degree of variation in genitalia and other body parts in twenty species of insects and spiders. *Evolution* **1998**, *52*, 415–431. [[CrossRef](#)] [[PubMed](#)]

**Disclaimer/Publisher’s Note:** The statements, opinions and data contained in all publications are solely those of the individual author(s) and contributor(s) and not of MDPI and/or the editor(s). MDPI and/or the editor(s) disclaim responsibility for any injury to people or property resulting from any ideas, methods, instructions or products referred to in the content.