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Butterflies (Lepidoptera, Papilionoidea and Hesperioidea) of the “Baixada Santista” region, coastal São Paulo, southeastern Brazil

Ronaldo Bastos Francini¹, Marcelo Duarte², Olaf Hermann Hendrik Mielke³,
Astrid Caldas⁴ & André Victor Lucci Freitas⁵

¹Curso de Ciências Biológicas, Universidade Católica de Santos, Campus D. Idílio José Soares, Av. Conselheiro Nébias, 300, 11065-902 Santos-SP, Brasil. francini@unisantos.edu.br

²Museu de Zoologia da Universidade de São Paulo, Avenida Nazaré 481, 04263-000 São Paulo-SP, Brasil. mduartes@usp.br

³Departamento de Zoologia, Universidade Federal do Paraná, Caixa Postal 19020, 81531-980 Curitiba-PR, Brasil. omhesp@ufpr.br

⁴Department of Entomology, National Museum of Natural History, Smithsonian Institution, PO Box 37012 NHB Stop 127, Washington, DC, 20013-7012 USA. astridcaldas@gmail.com

⁵Departamento de Biologia Animal and Museu de Zoologia, Instituto de Biologia, Universidade Estadual de Campinas, Caixa Postal 6109, 13083-970 Campinas-SP, Brasil. baku@unicamp.br (corresponding author)

ABSTRACT. Butterflies (Lepidoptera, Papilionoidea and Hesperioidea) of the “Baixada Santista” region, coastal São Paulo, southeastern Brazil. A list with 538 species of butterflies recorded in the Baixada Santista, São Paulo (SE Brazil) is presented. Standard sampling protocols (i.e. with entomological nets) were followed. Baited traps were installed for fruit feeding species. Data from the literature and entomological collections were also considered in the total estimated species richness. The species richness recorded in the Baixada Santista region represents about 16% of the Brazilian butterfly fauna, and 34% of the known butterfly fauna for the state of São Paulo. The present list contains an appreciably higher number of species in comparison to other lists from similar biomes farther south, such as Blumenau in Santa Catarina, and Maquiné in Rio Grande do Sul.

KEYWORDS. Atlantic Forest; conservation; diversity; Neotropical region.

RESUMO. Borboletas (Lepidoptera, Papilionoidea e Hesperioidea) da região da Baixada Santista, litoral de São Paulo (SE Brasil). Uma lista com 538 espécies de borboletas registradas na Baixada Santista é apresentada. Foram seguidos os protocolos amostrais padronizados (i.e. redes entomológicas). Armadilhas com iscas foram instaladas para coleta de espécies frugívoras. Dados de literatura e coleções entomológicas também foram considerados nas estimativas de riqueza de espécies. A riqueza de espécies registrada na Baixada Santista representa cerca de 16% da fauna de borboletas do Brasil, e 34% da fauna de borboletas do Estado de São Paulo. A presente lista contém um número consideravelmente alto de espécies em comparação com outras listas de biomas similares mais ao sul, como Blumenau em Santa Catarina, e Maquiné no Rio Grande do Sul.

PALAVRAS-CHAVE. Conservação; diversidade; Mata Atlântica; região Neotropical.

Interest in the conservation and monitoring of biological diversity has increased dramatically in the past 20 years. Nonetheless, species inventories and lists, arguably one of the most essential tools for any practical decision about species conservation (Lewinsohn & Prado 2002), are still lacking for most groups, including butterflies (Brown & Freitas 1999).

The Brazilian Atlantic Forest is a heterogeneous and endangered biome (Brown & Freitas 2000b), and according to Morellato & Haddad (2000) and Tabarelli *et al.* (2005) less than 10% of it remains. In the past ten years, a number of butterfly inventories in the Atlantic Forest have been published (see Santos *et al.* 2008 for a recent review on this topic), but much more information is needed for a better understanding of the general patterns of butterfly distribution in this biome.

There are about 3,300 species of butterflies in Brazil (Brown 1996; Brown & Freitas 1999; Lewinsohn *et al.* 2005), and more than 2,100 species can be found in the Atlantic Forest (Brown 1992). In the state of São Paulo, where these

insects have been reasonably well sampled (Brown 1992; Brown & Freitas 1999, 2000b), about 1,600 species can be found. But even in São Paulo there are still a variety of habitats, such as the wet forests in the coastal region, that lack adequate information on butterfly richness and faunistic composition (Brown & Freitas 1999).

Butterflies have been considered one of the most appropriate taxonomic groups for assessment of environmental disturbances in many parts of the world (Brown 1991; Freitas *et al.* 2006; Uehara-Prado *et al.* 2007 and references therein). They are excellent indicators in conservation planning (see Caldas & Robbins 2003 for examples) and are seen as valuable biodiversity indicators (McGeoch 1998). They are also an effective “umbrella group” for habitat and diversity conservation (New 1997). However, a first and critical step for using these organisms as indicators is to obtain local and regional lists of species.

In the present study we provide a list of the butterfly species found in the “Baixada Santista” region. Our objective is

not only to improve the knowledge of these insects in the coastal area of the state of São Paulo, but also to offer a tool for conservation and biodiversity decision-making.

For the purpose of this study, the Baixada Santista region includes the municipalities of Bertioga, Cubatão, Guarujá, Mongaguá, Praia Grande, Santos and São Vicente (Fig. 1, but see Kronka *et al.* 2005 for a definition based on hydrological resources). That entire region has been systematically explored since 1530, when the Portuguese colonizers arrived in Brazil, but the indigenous inhabitants probably used its natural resources extensively even before them (Petrone 1965).

The Baixada Santista is characterized by coastal plains, being delimited to the north by a continuous SW-NE mountain range with elevations varying from 800 m to 1,200 m (Serra do Mar), and to the south by the Atlantic Ocean. The regional climate is warm and wet, without a marked dry season (Santos 1965; SIGRH 2005; DAEE 2009). The average annual rainfall is about 2,500 mm, but towards the mountains it can reach values as high as 3,000–4,000 mm in some of the valleys and montane slopes. The average annual temperature is 22°C, with a maximum of 40°C and a minimum of 4°C. Frost have never been recorded in the region (Santos 1965).

According to Veloso *et al.* (1991), at least five primary distinct formations or physiognomies can be found in the region: sub-montane dense rain forest (mountain slopes from 50m to 500m); lowlands rain forest (clay soils, from 5m to 50m); sandy scrub forests (“restingas”) and dunes (sandy soils, from sea level to 10m); mangroves and salt marshes (all estuarine systems); and vegetation growing on rocky shores in the seashore. Besides these formations, the region is extensively covered by many anthropic systems with various levels of disturbance and regeneration.

MATERIAL AND METHODS

Species records were obtained through field collections conducted by RBF (1968–2009) and AVLF (1988–2009). At least one specimen of each species was collected, and deposited in one of the following Brazilian collections (exact numbers of vouchers can be obtained from their curators): Museu de Zoologia da USP (MZSP), São Paulo, São Paulo or Museu de Zoologia da Unicamp (ZUEC), Campinas, São Paulo. Additional data were obtained from specimens deposited in the collections of two institutions in Brazil: Museu de Zoologia da Universidade de São Paulo (MZSP), São Paulo, São Paulo, and Coleção de Entomologia Pe. J. S. Moure (DZUP), Departamento de Zoologia, Universidade Federal do Paraná, Curitiba, Paraná. Species without actual data on geographical distribution or only present in neighboring areas were not included.

A total of 48 sites throughout the entire Baixada Santista were sampled from 1968 to 2009, and they included all physiognomies described above (Fig. 1) that extended from 0 to 500 m above sea level. We had at least one sampling day in each one of these sites, with some sites being more intensively sampled than others. Well sampled sites (a total of 35

with more than five days of sampling) are shown in Fig. 1, and further details on each of these sites are provided on Table I. Among those, the most sampled sites are in the area of Xixová-Japuí State Park (sites 7–10 in Fig. 1), the area of the Rio Quilombo Valley (sites 21–22), the Voturuá Municipal Park (site 14), and the “Poço das Antas” area (sites 2–4), each one with more than 500 hours of sampling (details of the first three regions can be found in Freitas 1993, 1996 and Ramos & Freitas 1999).

Sampling protocols followed Brown (1972), Brown & Freitas (1999), and Freitas *et al.* (2003), with emphasis on diurnal collections made with nets, although butterfly traps were also used several times for sampling fruit feeding species.

The taxonomy follows mostly Lamas (2004), but also Willmott (2003) for the genus *Adelpha*, Mielke (2005) for Hesperioidae, Wahlberg *et al.* (2009) for the higher taxonomic categories for Nymphalidae, Duarte & Robbins (2010) for some hairstreaks (Lycaenidae, Theclinae, Eumaeini), and Hall (2005) for some metalmarks (Riodinidae).

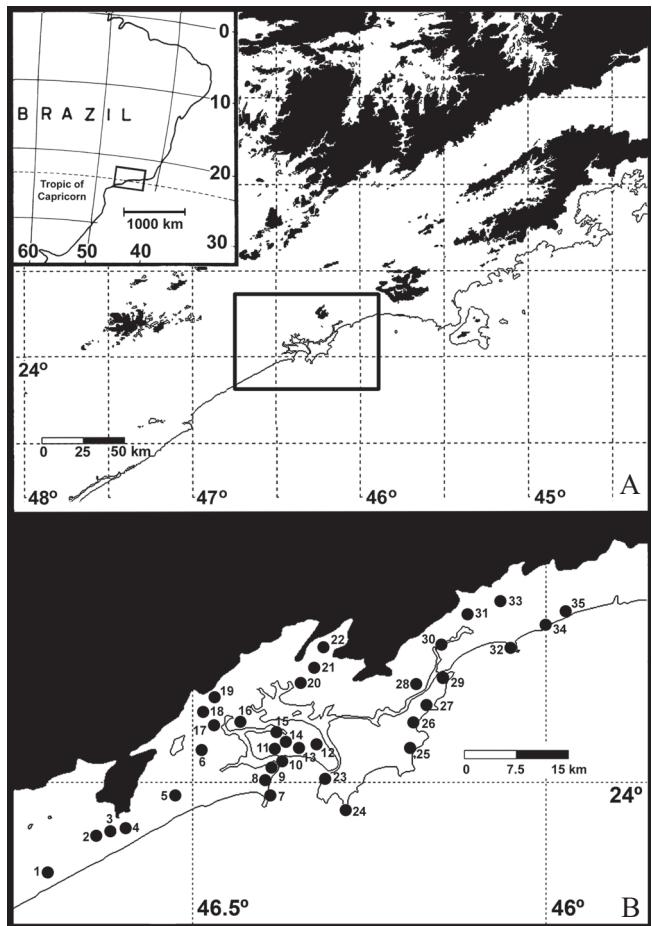


Fig. 1. Location of the Baixada Santista region in Southeastern Brazil. A. Enlarged view of the black rectangle in the South America map (top and left). B. Enlarged view of the black rectangle in A (the Baixada Santista region). Solid circles show the 35 most sampled sites in the Baixada Santista region (for details of each site see Table I). Black areas indicate altitudes above 1000m in A, and above 500m in B. Source: modified from IBGE 2009.

Table I. Descriptors of sampled sites showed in the local map of Fig. 1.

Number	Name	Municipality	Coordinates
1	Balneário Itaóca	Mongaguá	24°06'S; 46°40'W
2	Poço das Antas	Mongaguá	24°05'S; 46°37'W
3	Serra de Mongaguá	Mongaguá	24°05'S; 46°36'W
4	Cidade da Criança	Praia Grande	24°04'S; 46°35'W
5	Restinga da Vila Caiçara	Praia Grande	24°01'S; 46°30'W
6	Rio Branco	São Vicente	23°56'S; 46°28'W
7	Fortaleza Itaipu	Praia Grande	24°01'S; 46°23'W
8	Praia de Itaquitanuva	São Vicente	24°00'S; 46°23'W
9	Morro do Japuí	São Vicente	23°59'S; 46°23'W
10	Praia das Vacas	São Vicente	23°59'S; 46°22'W
11	Morro dos Barbosas	São Vicente	23°58'S; 46°23'W
12	Santos urban area	Santos	23°57'S; 46°18'W
13	Orquidário/Morro da Nova Cintra	Santos	23°58'S; 46°20'W
14	Morro do Voturuá	São Vicente/Santos	23°57'S; 46°21'W
15	Jardim Casqueiro	Santos/Cubatão	23°56'S; 46°23'W
16	Ilha Caraguatá	Cubatão	23°55'S; 46°23'W
17	Manoel da Nóbrega Highway	Cubatão	23°55'S; 46°27'W
18	Rio Cubatão valley	Cubatão	23°55'S; 46°29'W
19	Anchieta Highway	São Vicente/Cubatão	23°53'S; 46°28'W
20	Trapucaia Farm	Santos	23°52'S; 46°20'W
21	Rio Quilombo valley	Santos	23°50'S; 46°19'W
22	Upper Rio Quilombo	Santos	23°49'S; 46°18'W
23	Praia do Góes	Guarujá	24°00'S; 46°18'W
24	Forte dos Andradadas	Guarujá	24°01'S; 46°17'W
25	Praia de Pernambuco	Guarujá	23°57'S; 46°11'W
26	Praia do Iporanga	Guarujá	23°53'S; 46°09'W
27	Praia do Camburi	Guarujá	23°52'S; 46°11'W
28	Caiubura	Bertioga	23°52'S; 46°11'W
29	Morro da Armação	Guarujá	23°51'S; 46°08'W
30	Restinga de Bertioga	Bertioga	23°49'S; 46°09'W
31	Usina Itatinga	Bertioga	23°46'S; 46°06'W
32	Praia do Indaiá	Bertioga	23°49'S; 46°02'W
33	Fazenda Acaraú	Bertioga	23°45'S; 46°02'W
34	Praia de Itaguaré	Bertioga	23°46'S; 45°57'W
35	Praia de Guaratuba	Bertioga	23°45'S; 45°53'W

RESULTS AND DISCUSSION

A total of 538 butterfly species were recorded in the Baixada Santista region (Appendix I) representing about 16% of the Brazilian butterfly fauna, and 34% of the known butterfly fauna for the state of São Paulo (Brown & Freitas 1999). These numbers are in accordance with the expected richness of the region (according to Brown & Freitas 2000b), and considering the large sampling effort, this list can be considered

relatively complete (but see also the next sections). From the total of 538 species, 340 (63%) are Papilionoidea and 198 (37%) are Hesperioidea. The best represented family in the study area is Hesperiidae, with 198 species, followed by Nymphalidae with 179 species (Appendix I). These two families are the richest in all Neotropical sites surveyed to date, followed by Lycaenidae, Riodinidae, Pieridae and Papilionidae, the latter two with much lower richness (see general site lists compiled in Brown & Freitas 1999, 2000b; Brown 2005).

This total of 538 species is appreciably higher than the number of species found in some of the lists from similar biomes farther south, such as Blumenau (376 species) in Santa Catarina, and Maquiné (292 species) in Rio Grande do Sul (Iserhard & Romanowski 2004; OHMM and AVL, unpublished data). Some sites with richness similar to Baixada Santista can be found in the southern sector of the Atlantic Forest in Paraná and Santa Catarina states, such as Joinville with about 757 species already recorded, Jaraguá do Sul with about 540 species, and Curitiba with 528 species (Mielke 1996; Carlos G. C. Mielke, OHMM and AVL, unpublished data). However, the number of species recorded at Baixada Santista is well below the number recorded in more tropical areas in Rio de Janeiro and Espírito Santo states, where some sites can have over 800 species (Brown & Freitas 2000a, b; Brown 2005). The species richness seems to be higher also in some well sampled sites in the seasonal semideciduous forests in the interior, such as the Mata da Santa Genebra (702 species) and Ribeirão Cachoeira (567 species) in São Paulo state, Yacutinga (572 species) and Iguazú (652 species) in Misiones (Northern Argentina), and also in complex mosaic landscapes of deep valleys and high mountains in localities across several southeastern states, such as the Serra do Japi in São Paulo (679 species), Poços de Caldas in Minas Gerais (577 species) and Itatiaia in Rio de Janeiro (914 species) (Zikán & Zikán 1968; Ebert 1969; Brown 1992; Brown & Freitas 2000b, 2002b; Bustos 2008, 2009). It is important to note that none of the species collected in this study has been considered threatened of extinction in the Brazilian Red List (Machado *et al.* 2008).

Species richness. The species richness of the family Nymphalidae at the Baixada Santista is considered very accurate; we expect to have very few additional records to the list. For example, the nymphalids *Eunica eurota dolores* (Prittitz, 1871) (Biblidinae), *Archeuptychia cluena* (Drury, 1782) (Satyrinae), *Cissia myncea* (Cramer, 1780) (Satyrinae), and *Scada karschiana karschiana* (Herbst, 1792) (Danainae) occur in sites less than 20 km north and east of the study region in the same biome (in lowland and submontane rain forests; AVL and RBF, pers. obs.), and could well be present in small populations in the mountain slopes north of Bertioga, including the Usina Itatinga and Fazenda Acaraú (see Fig. 1).

Based on museum collections and field experience of the authors, very few new records would also be expected in Pieridae and Papilionidae. The few additions might include the papilionids *Protagonistus asius* (Fabricius, 1781) and *Mimoides lysithous* (Hübner, 1821), both occurring north and south of Baixada Santista and also recorded in neighbor mountain sites. In the Pieridae, some migratory and widespread species could still be added to the list, including *Anteos clorinde* (Godart, [1824]) and *Phoebis neocypris* (Hübner, [1823]). Montane species could also appear in winter months together with the already recorded *Catasticta bithys* (Hübner, [1831]) and *Pereute antodyca* (Boisduval, 1836). These additional records might include *Theochila maenacte itatiayae* (Foetterle, 1902) (recorded feeding on flowers in Ubatuba,

northern São Paulo, at sea level) and *Hesperocharis erota* (Lucas, 1852), two species commonly found in the mountains above 800m around the study region.

In the remaining three families, additions are expected to be greater. The Lycaenidae may grow up to more than 70 species, and the Riodinidae to near 60 species (see Brown & Freitas 2000b and data in the present work). Examples of species of Riodinidae recorded less than 50 km north and east of the study area includes *Emesis ocypore zelotes* Hewitson, 1872, *E. neemias* Hewitson, 1872, *E. elegia* Stichel, 1929, *Isapis agyrtus abydus* Stichel 1929, *Aricoris constantius* (Fabricius, 1793), *Panaropsis inaria* (Westwood, 1851), *Calospila apotheta* (H. W. Bates, 1868) and *Stalachitis phlegia susanna* (Fabricius, 1787). In the Hesperiidae the increase may also be significant, especially due to the low sampling of the fast flying Pyrrhopyginae for the region, and up to 10 species could be added to the list.

Several studies propose that the local number of species can be predicted based on surrogates of total butterfly richness (Beccaloni & Gaston 1995; Brown & Freitas 2000b). In that sense, the predicted richness for the Baixada Santista region based on the Ithomiini (4.3–4.6% of total fauna) and Nymphalidae (25–29% of total fauna) as surrogates would be between 558 and 660 species, a figure above the observed total of 538 species. If the above predictions of probable additions to each family are confirmed, the list of butterflies for the Baixada Santista would reach approximately 570 species, a number near that predicted based on Ithomiini richness only. However, if we are to make a prediction based on surrogates, and use the Nymphalidae as the best surrogate assuming that this is a well sampled group, and that an entire family is a better surrogate than a tribe, then it would be fair to assert that we have collected only 81.7% of the total butterfly fauna of the Baixada Santista. Therefore, even if our list is considered enough to provide a clear picture of the faunistic composition in the study region, several new records may be expected to be added to the list in all six families (in different proportions) with further field work in some of the less surveyed sites.

Taxonomic composition. In the present study, the diversity of larger butterfly groups follow the pattern reported in the total list of Brazilian butterflies, where Hesperiidae is the richest family, followed by Nymphalidae (Brown & Freitas 1999 and Table II). This pattern was also reported in other relatively complete inventories in the Neotropics, showing Hesperiidae as the best represented butterfly group in any well surveyed locality, followed by the Nymphalidae (Brown & Freitas 2000b). However, in short term studies with low sampling effort there has been a clear numerical dominance of Nymphalidae (Motta 2002; Iserhard & Romanowski 2004; Duarte & Almeida 2006; Marchiori & Romanowski 2006), likely due to the fact that they are easily captured and identified in the field. The Hesperiidae, Lycaenidae and Riodinidae, all including many small and inconspicuous species, require more effort to be adequately sampled. Because species of these three groups are difficult to identify, they are also usu-



Fig. 2. Overview of the main landscapes in the Baixada Santista region. For vegetation description and site details see text and Table I. A. General view of the region showing the large mangroves: hills in the horizon are the Japuí-Xixová State Park; B. Secondary open vegetation and lowland rainforest: Morro do Japuí; C. Restinga vegetation: Bertioga; D. Submontane dense rainforest: slopes near Fazenda Acaraú; E. Dune and restinga vegetation: Praia de Itaguaré; F. Mangroves: Praia de Itaguaré.

ally undersampled, or not even considered in some studies (e.g. Almeida *et al.* 1986; Paz *et al.* 2008). We believe that a higher numerical dominance of the Hesperiidae is expected in all complete or near-complete butterfly surveys.

Concerning the Lycaenidae and Riodinidae, the pattern of richness reported in the present list is not the same of that observed in the total Brazilian list, where the latter is richer than the former (Table II). In a comparative analysis,

Lycaenidae is richer than Riodinidae in most southern Atlantic Forest sites and in the montane forests, with the only remarkable exception being Joinville, in Santa Catarina state (Table II). In fact, the richness of Riodinidae at a given site has been shown to be positively correlated with the mean temperature of the site (Brown 2005). This may explain why this family is better represented in warmer forests of northern Atlantic Forest (Brown & Freitas 2000b and Table II), and in several Amazonian sites (Brown & Freitas 1999, 2002a; Brown 2005).

CONCLUSIONS

The butterfly fauna of the Baixada Santista region deserves attention (i.e. continuing studies and long-range conservation strategies) even if no threatened species have been recorded in the present study. The entire area shows a unique combination of different habitats and geography that includes deep valleys, expansive mangroves, high mountains and lowland plains, which provide conditions for the presence of a rich and diverse fauna combining elements from

Table II. Diversity and community composition of butterflies recorded in Brazil, in São Paulo State, and in 22 sites in the Atlantic Forest domain. All sites are in Brazil except Yacutinga and Iguazú, located in Northern Argentina. Asterisks indicate the richest butterfly family in each site.

	Hesperiidae	Papilionidae	Pieridae	Lycaenidae	Riodinidae	Nymphalidae	Total
Brazil – total list ^{1,2}	1165*	69	65	420	761	788	3268
São Paulo ^{1,2,3}	689*	35	46	206	222	389	1587
Coastal Brazil (southern)							
Baixada Santista ^{4,5}	198*	18	29	65	49	179	538
Joinville ^{4,6}	364*	21	31	68	86	187	757
Jaraguá do Sul ⁷	237*	19	36	47	47	165	551
Blumenau ⁸	112	13	33	26	25	167*	376
Maquiné ⁹	97	13	24	35	19	104*	292
Coastal Brazil (northern)							
Santa Teresa ^{4,10}	322*	17	36	64	86	244	769
Linhares ⁴	376*	19	25	92	105	218	835
Xerém ⁴	131	22	29	43	60	177*	462
Rio de Janeiro ^{4,24,25}	250*	42	31	133	80	168	697
Montane forests							
Itatiaia ^{4,11}	366*	22	38	144	116	228	914
Serra do Japi ^{3,4,12}	253*	19	36	111	52	208	679
Poços de Caldas ^{4,13}	224*	16	35	87	47	168	577
Morro Grande ^{14, 15}	198*	16	35	54	49	188	540
Curitiba ^{6,16}	233*	14	30	53	44	154	528
Semideciduous forests							
Santa Genebra ^{3,4,17}	311*	17	28	91	42	213	702
Ribeirão Cachoeira ^{4,15,17}	245*	14	26	75	38	169	567
Morro do Diabo ^{3,18}	197*	15	19	35	38	156	460
Yacutinga ¹⁹	248*	12	21	68	49	174	572
Iguazú ²⁰	283*	18	30	72	60	189	652
Cerrado áreas							
Brasília ^{21,22}	335*	16	26	112	137	213	839
Belo Horizonte ⁴	170*	14	30	79	35	159	487
Uberlândia ²³	52	8	17	36	28	110*	251

¹Brown & Freitas (1999); ²M. Duarte (unpublished data); ³Brown & Freitas (unpublished data); ⁴Brown & Freitas (2000b); ⁵Present study; ⁶O. H. H. Mielke & C. G. C. Mielke (unpublished); ⁷A. V. L. Freitas unpublished (data compiled from early lists by Fritz Hoffmann); ⁸O. H. H. Mielke & A. V. L. Freitas (unpublished data); ⁹Iserhard & Romanowski (2004); ¹⁰Brown & Freitas (2000a); ¹¹Zikán & Zikán (1968); ¹²Brown (1992); ¹³Ebert (1969); ¹⁴Uehara-Prado *et al.* (2004); ¹⁵Uehara-Prado, Brown & Freitas (unpublished); ¹⁶Mielke (1996); ¹⁷Brown & Freitas (2002b); ¹⁸Mielke & Casagrande (1998); ¹⁹Bustos (2008); ²⁰Bustos (2009); ²¹Emery *et al.* (2006); ²²Mielke *et al.* (2008); ²³Motta (2002); ²⁴Duarte *et al.* (2010); ²⁵Monteiro *et al.* (2010).

both the north and the south of Brazil. This combination can be found again farther south, in the Iguape-Cananéia region, near the border between the States of São Paulo and Paraná, but in that area the temperature drops near freezing levels several times during the winter months, a feature that could explain the absence of several of the tropical species observed in the Baixada Santista region, and that contributes to its uniqueness.

It is clear to us that, even with the large amount of information that exists for the Brazilian Atlantic Forest butterflies (see Brown & Freitas 2000a, b and Santos *et al.* 2008) there are relatively few well sampled sites to allow comparative studies. Concentrated efforts to generate other lists of species are needed to help understand the distribution patterns of butterfly richness along the various environmental gradients of this endangered Biome (see also Duarte *et al.* 2010; Monteiro *et al.* 2010).

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Appendix I. Butterflies (Papilioidea and Hesperioida) from the Baixada Santista region. Number of species are provided within parenthesis for each major taxa. Total: 538 species.

Papilioidea (340)	Lycenidae (65)
Papilionidae (18)	Polyommatainae (3)
Papilioninae (18)	<i>Leptotes cassius cassius</i> (Cramer, 1775)
Leptocircini (4)	<i>Hemiargus hanno hanno</i> (Stoll, 1790)
<i>Eurytides dolicaon deicoon</i> (C. Felder & R. Felder, 1864)	<i>Zizula cyna</i> (W. H. Edwards, 1881)
<i>Protesilaus protesilaus nigricornis</i> (Staudinger, 1884)	Theclinae (62)
<i>Protesilaus telesilaus vitellus</i> (Fruhstorfer, 1907)	<i>Brangas ca. getus</i> (Fabricius, 1787)
<i>Mimoides protodamas</i> (Godart, 1819)	<i>Evenus regalis</i> (Cramer, 1775)
Troidini (7)	<i>Evenus satyroides</i> (Hewitson, 1865)
<i>Battus polydamas polydamas</i> (Linnaeus, 1758)	<i>Atlides cosa</i> (Hewitson, 1867)
<i>Battus polystictus galenus</i> (Fruhstorfer, 1907)	<i>Atlides rustan</i> (Stoll, 1790)
<i>Battus crassus crassus</i> (Cramer, 1777)	<i>Pseudolycaena marsyas</i> (Linnaeus, 1758)
<i>Parides agavus</i> (Drury, 1782)	<i>Theritas triquetra</i> (Hewitson, 1865)
<i>Parides anchises nephalion</i> (Godart, 1819)	<i>Theritas hemon</i> (Cramer, 1775)
<i>Parides tros tros</i> (Fabricius, 1793)	<i>Theritas lisus</i> (Stoll, 1790)
<i>Parides zacynthus zacynthus</i> (Fabricius, 1793)	<i>Thereus</i> sp.
Papilionini (7)	<i>Rekoia meton</i> (Cramer, 1779)
<i>Heraclides anchisiades capys</i> (Hübner, [1809])	<i>Rekoia palegon</i> (Cramer, 1780)
<i>Heraclides androgeus laodocus</i> (Fabricius, 1793)	<i>Rekoia marius</i> (Lucas, 1857)
<i>Heraclides hectorides</i> (Esper, 1794)	<i>Rekoia stagira</i> (Hewitson, 1867)
<i>Heraclides thoas brasiliensis</i> (Rothschild & Jordan, 1906)	<i>Arawacus meliboeus</i> (Fabricius, 1793)
<i>Heraclides torquatus polybius</i> (Swainson, 1823)	<i>Arawacus ellida</i> (Hewitson, 1867)
<i>Pterourus menatius cleotas</i> (Gray, 1832)	<i>Ocaria thales</i> (Fabricius, 1793)
<i>Pterourus scamander grayi</i> (Boisduval, 1836)	<i>Chlorostrymon simaethis</i> (Drury, 1773)
Pieridae (29)	<i>Chlorostrymon telea</i> (Hewitson, 1868)
Dismorphiinae (8)	<i>Magnastigma hirsuta</i> (Prittitz, 1865)
<i>Dismorphia amphione astynome</i> (Dalman, 1823)	<i>Cyanophrys acaste</i> (Prittitz, 1865)
<i>Dismorphia astyocha</i> Hübner, [1831]	<i>Cyanophrys herodotus</i> (Fabricius, 1793)
<i>Dismorphia crisia crisia</i> (Drury, 1782)	<i>Allosmaitia strophius</i> (Godart, [1824])
<i>Dismorphia melia</i> (Godart, [1824])	<i>Laothus phydelta</i> (Hewitson, 1867)
<i>Dismorphia thermesia</i> (Godart, 1819)	<i>Janthecla armilla</i> (H.H. Druce, 1907)
<i>Enantia lina psamathe</i> (Fabricius, 1793)	<i>Janthecla aurora</i> (H.H. Druce, 1907)
<i>Enantia limnorina</i> (C. Felder & R. Felder, 1865)	<i>Janthecla flosculus</i> (H.H. Druce, 1907)
<i>Enantia clarissa</i> (Weymer, 1895)	<i>Badecla badaca</i> (Hewitson, 1868)
Coliadinae (14)	<i>Lamprospilus orcidia</i> (Hewitson, 1874)
<i>Anteos menippe</i> (Hübner, [1818])	<i>Ziegleria hesperitis</i> (Butler & H. Druce, 1872)
<i>Phoebeis argante argante</i> (Fabricius, 1775)	<i>Kisutam syllis</i> (Godman & Salvin, 1887)
<i>Phoebeis philea philea</i> (Linnaeus, 1763)	<i>Electrostrymon endymion</i> (Fabricius, 1775)
<i>Phoebeis sennae marcellina</i> (Cramer 1777)	<i>Calycopis caulonia</i> (Hewitson, 1877)
<i>Rhabdodryas trite banksi</i> (Breyer, 1939)	<i>Calycopis gentilla</i> (Schaus, 1902)
<i>Aphrissa statira statira</i> (Cramer, 1777)	<i>Calycopis janeirica</i> (C. Felder, 1862)
<i>Pyrisitia leuce leuce</i> (Boisduval, 1836)	<i>Calycopis</i> sp.
<i>Pyrisitia nise tenella</i> (Boisduval, 1836)	<i>Strymon mulucha</i> (Hewitson, 1867)
<i>Eurema agave pallida</i> (Chavannes, 1850)	<i>Strymon bazochii</i> (Godart, [1824])
<i>Eurema deva deva</i> (Doubleday, 1847)	<i>Strymon bubastus</i> (Stoll, 1780)
<i>Eurema albula sinoe</i> (Godart, 1819)	<i>Strymon oreala</i> (Hewitson, 1868)
<i>Eurema elathea flavescentia</i> (Chavannes, 1850)	<i>Strymon ziba</i> (Hewitson, 1868)
<i>Eurema phiale paula</i> (Röber, 1909)	<i>Tmolus echion</i> (Linnaeus, 1767)
<i>Leucidia elvina</i> (Godart, 1819)	<i>Nicolaea besidia</i> (Hewitson, 1868)
Pierinae (7)	<i>Ministrymon cruenta</i> (Gosse, 1880)
<i>Archonias brassolis tereas</i> (Godart, 1819)	<i>Ministrymon azia</i> (Hewitson, 1873)
<i>Catasticta bithys</i> (Hübner, [1831])	<i>Ministrymon</i> sp.
<i>Pereute antodyca</i> (Boisduval, 1836)	<i>Gargina caninius</i> (H. H. Druce, 1907)
<i>Melete lycimnia flippantha</i> (Fabricius, 1793)	<i>Theclopsis murex</i> (H. H. Druce, 1907)
<i>Glutophrissa drusilla drusilla</i> (Cramer, 1777)	<i>Ostrinotes ca. empusa</i> (Hewitson, 1867)
<i>Leptophobia aripa balidia</i> (Boisduval, 1836)	<i>Strephonota sphinx</i> (Fabricius, 1775)
<i>Ascia monuste orseis</i> (Godart, 1819)	<i>Pantheodes hebraeus</i> (Hewitson, 1867)
	<i>Thepytus echelta</i> (Hewitson, 1867)
	<i>Oenomaus atesa</i> (Hewitson, 1867)
	<i>Parrhasius polibetes</i> (Stoll, 1781)

- Michaelus jebus* (Godart, [1824])
Aubergina vanessoides (Prittitz, 1865)
Iaspis talayra (Hewitson, 1868)
Celmia celmus (Cramer, 1775)
Celmia conoveria (Scahus, 1902)
Erora sp.
Chalybs hassan (Stoll, 1790)
Symbiopsis strenua (Hewitson, 1877)
- Riodinidae (49)
- Euselasiinae (5)
- Euselasia hygenius occulta* Stichel, 1919
 - Euselasia fervida fervidina* Stichel, 1919
 - Euselasia eucerus* (Hewitson, 1872)
 - Euselasia thucydides thucydides* (Fabricius, 1793)
 - Euselasia clesa* (Hewitson, 1856)
- Riodininae (44)
- Mesosemiini (7)
- Mesosemia odice* (Godart, [1824])
 - Mesosemia rhodia* (Godart, [1824])
 - Mesosemia meeda* Hewitson, 1858
 - Leucochimona icare matatha* (Hewitson, 1873)
 - Voltinia phryxe* (C. Felder & R. Felder, 1865)
 - Voltinia cebrenia* (Hewitson, [1873])
 - Cremania alector* (Geyer, 1837)
- Eurybiini (2)
- Eurybia pergaea* (Geyer, 1832)
 - Eurybia molochina hyacinthina* Stichel, 1910
- Riodinini (12)
- Lyropteryx apollonia apollonia* Westwood, 1851
 - Ancyluris aulestes pandama* (Saunders, 1850)
 - Rhetus periander eleusinus* Stichel, 1910
 - Panara soana soana* Hewitson, 1875
 - Brachyglenis drymo* (Godman & Salvin, 1886)
 - Metacharis ptolomaeus* (Fabricius, 1793)
 - Pheles atricolor* (Butler, 1871)
 - Barbicornis basilis basilis* Godart, [1824]
 - Syrmatia nyx* (Hübner, [1817])
 - Charis cadytis* Hewitson, 1866
 - Lasaia agesilas* (Latrelle, [1809])
 - Riodina lycisca lycisca* (Hewitson, [1863])
- Symmachiiini (4)
- Pirascca arbuscula mandosa* (H. Druce, 1904)
 - Pirascca sagaris satnius* (Dalman, 1823)
 - Pterographium sicora* (Hewitson, 1875)
 - Stichelia bocchoris suavis* (Stichel, 1911)
- Helicopini (2)
- Anteros lectabilis* Stichel, 1909
 - Anteros renaldus notius* Stichel, 1911
- Nymphidiini (11)
- Lemonias zygia zygia* Hübner, [1807]
 - Juditha molpe* (Hübner, [1808])
 - Synargis calyce* (C. Felder & R. Felder, 1862)
 - Synargis phliasus phliasus* (Clerck, 1764)
 - Menander menander nitida* (Butler, 1867)
 - Menander felsina* (Hewitson, 1863)
 - Adelotyta malca* (Schaus, 1902)
 - Nymphidium lisimon lisimon* (Stoll, 1790)
 - Catocyclotis aemulius* (Fabricius, 1793)
 - Theope teramus* (Godart, [1824])
 - Theope lycaenina* H. W. Bates, 1868
- Incertae sedis (6)
- Echydna chaseba* (Hewitson, 1854)
 - Emesis fastidiosa* Ménétriés, 1855
 - Emesis fatimella fatimella* Westwood, 1851
 - Emesis mandana* (Cramer, 1780)
 - Emesis diogenia* Prittitz, 1865
 - Emesis* sp.
- Nymphalidae (179)
- Libytheinae (1)
- Libytheana carinenta* (Cramer, 1777)
- Danainae (29)
- Danaini (5)
- Euploea* (2)
 - Lycorea halia discreta* Haensch, 1909
 - Lycorea ilione ilione* (Cramer, 1775)
- Danaina (3)
- Danaus eresimus plexaure* (Godart, 1819)
 - Danaus erippus* (Cramer, 1775)
 - Danaus gilippus gilippus* (Cramer, 1775)
- Ithomiini (24)
- Tithoreina (1)
- Aeria olena olena* Weymer, 1875
- Melinaeina (2)
- Melinaea ethra* (Godart, 1819)
 - Melinaea ludovica paraiya* Reakirt, 1866
- Mechanitina (3)
- Methona themisto* (Hübner, 1818)
 - Mechanitis lysimnia lysimnia* (Fabricius, 1793)
 - Mechanitis polymnia casabranca* Haensch, 1905
- Napeogenina (2)
- Epityches eupompe* (Geyer, 1832)
 - Hypothyris ninonia daeta* (Boisduval, 1836)
- Ithomiina (4)
- Placidina euryanassa* (C. Felder & R. Felder, 1860)
 - Ithomia agnoscia zikani* D'Almeida, 1940
 - Ithomia drymo* Hübner, 1816
 - Ithomia lichyi lichyi* D'Almeida, 1939
- Olerina (1)
- Oleria aquata* (Weymer, 1875)
- Dirceniina (7)
- Callithomia lenea xantho* (C. Felder & R. Felder, 1860)
 - Dircenna dero* (Hübner, 1823)
 - Episcada clausina striposis* Haensch, 1909
 - Episcada hymenaea hymenaea* (Prittitz, 1865)
 - Episcada philoclea* (Hewitson, [1855])
 - Pteronymia euritea* (Cramer, 1780)
 - Pteronymia carlia* Schaus, 1902
- Godyridina (4)
- Hypoleria adasa adasa* (Hewitson, [1855])
 - Heterosais edessa* (Hewitson, [1855])
 - Pseudoscada acilla acilla* (Hewitson, 1867)
 - Pseudoscada erruca* (Hewitson, 1855)
- Satyrinae (40)
- Morphini (7)
- Antirrhea archaea* Hübner, [1822]
 - Morpho aega* (Hübner, [1822])
 - Morpho anaxibia* (Esper, 1801)
 - Morpho epistrophus* (Fabricius, 1796)
 - Morpho helenor achillaena* (Hübner, [1823])
 - Morpho menelaus* (Linnaeus, 1758)
 - Morpho hercules* (Dalman, 1823)

- Brassolini (11)
- Brassolis astyra astyra* Godart, [1824]
 - Brassolis sophorae sophorae* (Linnaeus, 1758)
 - Caligo beltrao* (Illiger, 1801)
 - Caligo brasiliensis brasiliensis* (C. Felder, 1862)
 - Caligo illioneus illioneus* (Cramer, 1775)
 - Caligo idomeneus ariphron* Fruhstorfer, 1910
 - Catoblepia amphirhoe* (Hübner, [1825])
 - Dasyophtalma creusa creusa* (Hübner, [1821])
 - Eryphanis reevesi reevesi* (Doubleday, [1849])
 - Opsiphanes invirae pseudophilon* Fruhstorfer, 1907
 - Narope cyllastros* Doubleday, [1849]
- Haeterini (2)
- Pierella lamia lamia* (Sulzer, 1776)
 - Pierella nereis* (Drury, 1782)
- Elymniini (1)
- Manataria hercyna* (Hübner, [1821])
- Satyrini (19)
- Pronophilina (3)
 - Eteona tisiphone* (Boisduval, 1836)
 - Praepedalioides amussis* (Thieme, 1905)
 - Praepedalioides phanias* (Hewitson, 1862) - Euptychiina (16)
 - Carminda paeon* (Godart, [1824])
 - Chloreuptychia arnaca* (Fabricius, 1776)
 - Euptychia ernestina* Weymer, 1911
 - Euptychia westwoodi* Butler, 1867
 - Eptychoides castrensis* (Schaus, 1902)
 - Forsterinaria necys* (Godart, [1824])
 - Forsterinaria quantius* (Godart, [1824])
 - Godartiana muscosa* (Butler, 1870)
 - Hermeuptychia hermes* (Fabricius, 1775)
 - Pareuptychia ocirrhoe interjecta* (D'Almeida, 1952)
 - Paryphthimoides phronius* (Godart, 1824)
 - Splendeuptychia doxes* (Godart, [1824])
 - Splendeuptychia hygina* (Butler, 1877)
 - Taygetis laches marginata* Staudinger, [1887]
 - Taygetis rufomarginata* Staudinger, 1888
 - Yphthimoides renata* (Stoll, 1780)

Charaxinae (16)

 - Anaeini (11)
 - Consul fabius drurii* (Butler, 1874)
 - Hypna clytemnestra huebneri* Butler, 1866
 - Siderone galanthis galanthis* (Cramer, 1775)
 - Zaretis strigosus* (Gmelin, [1790])
 - Fountainea ryphea phidile* (Geyer, 1837)
 - Memphis appias* (Hübner, 1825)
 - Memphis leonida editha* (W. P. Comstock, 1961)
 - Memphis moruus stheno* (Prittitz, 1865)
 - Memphis acidalia victoria* (H. Druce, 1877)
 - Memphis otrere* (Hübner, [1825])
 - Memphis philumena corita* (Fruhstorfer, 1916) - Preponini (5)
 - Archaeoprepona amphimachus pseudomeander* (Fruhstorfer, 1906)
 - Archaeoprepona demophon thalpius* (Hübner, [1814])
 - Archaeoprepona demophoon antimache* (Hübner, [1819])
 - Archaeoprepona meander castorina* (E. May, 1932)
 - Prepona pylene pylene* Hewitson, [1854]

Biblidinae (26)

 - Biblis hyperia nectanabis* (Fruhstorfer, 1909)

Catonephele (11)

 - Catonephele acontius acontius* (Linnaeus, 1771)
 - Catonephele numilia penthia* (Hewitson, 1852)
 - Eunica maja maja* (Fabricius, 1775)
 - Eunica sydonia sydonia* (Godart, [1824])
 - Eunica volumna volumna* (Godart, [1824])
 - Myscelia orsis* (Drury, 1782)
 - Ectima thecla thecla* (Fabricius, 1796)
 - Hamadryas amphinome amphinome* (Linnaeus, 1767)
 - Hamadryas arete* (Doubleday, 1847)
 - Hamadryas arinome obnubila* (Fruhstorfer, 1916)
 - Hamadryas februa februa* (Hübner, [1823])
 - Hamadryas feronia feronia* (Linnaeus, 1758)
 - Hamadryas fornax fornax* (Hübner, [1823])
 - Hamadryas epinome* (C. Felder & R. Felder, 1867)
 - Pyrrhogrya neaerea ophni* Butler, 1870
 - Nica flavilla flavilla* (Godart, [1824])
 - Temenis laothoe meridionalis* Ebert, 1965
 - Dynamine agacles* (Dalman, 1823)
 - Dynamine artemisia artemisia* (Fabricius, 1793)
 - Dynamine athemon maeon* (Doubleday, 1849)
 - Dynamine postverta postverta* (Cramer, 1779)
 - Callicore astarte selima* (Guenée, 1872)
 - Callicore pygas eucale* (Fruhstorfer, 1916)
 - Diaeathria clymena meridionalis* (H. W. Bates, 1864)
 - Haematera pyrame pyrame* (Hübner, [1819])

Apaturinae (3)

 - Doxocopa agathina vacuna* (Godart, [1824])
 - Doxocopa laurentia laurentia* (Godart, [1824])
 - Doxocopa kallina* (Staudinger, 1886)

Cyrestinae (3)

 - Marpesia chiron chiron* (Fabricius, 1775)
 - Marpesia petreus petreus* (Cramer, 1776)
 - Marpesia zerynthia* Hübner [1823]

Nymphalinae (22)

 - Coeini (4)
 - Colobura dirce dirce* (Linnaeus, 1758)
 - Historis acheronta* (Fabricius, 1775)
 - Historis odious dious* Lamas, 1995
 - Smyrna blomfildia blomfildia* (Fabricius, 1781) - Nymphalini (4)
 - Hypanartia bella* (Fabricius, 1793)
 - Hypanartia lethe* (Fabricius, 1793)
 - Vanessa braziliensis* (Moore, 1883)
 - Vanessa myrinna* (Doubleday, 1849) - Kallimini (5)
 - Anartia amathea roeselia* (Eschscholtz, 1821)
 - Anartia jatrophae jatrophae* (Linnaeus, 1763)
 - Junonia evarete* (Cramer, 1779)
 - Siproeta epaphus trayja* (Hübner, [1823])
 - Siproeta stelenes meridionalis* (Fruhstorfer, 1909) - Melitacini (9)
 - Chlosyne lacinia saundersi* (Doubleday, [1847])
 - Anthanassa frisia hermas* (Hewitson, 1864)
 - Eresia eunice esora* Hewitson, 1857
 - Eresia lansdorfi* (Godart, 1819)
 - Eresia perna perna* Hewitson, 1852
 - Ortilia ithra* (W. F. Kirby, 1900)
 - Ortilia orticas orticas* (Schaus, 1902)
 - Tegosa claudina* (Eschscholtz, 1821)
 - Telenassa teletusa teletusa* (Godart, [1824])

- Limenitidinae (13)
- Adelpha serpa serpa* (Boisduval, 1836)
 - Adelpha hyas hyas* (Doyère, [1840])
 - Adelpha mythra* (Godart, [1824])
 - Adelpha plesaure plesaure* Hübner, 1823
 - Adelpha iphiclus ephesa* (Ménétriés, 1857)
 - Adelpha melona pseudarete* Fruhstorfer, 1915
 - Adelpha syma* (Godart, [1824])
 - Adelpha cythereaaea* (C. Felder & R. Felder, 1867)
 - Adelpha viola viola* Fruhstorfer, 1913
 - Adelpha capucinus velia* (C. Felder & R. Felder, 1867)
 - Adelpha thesprotia* (C. Felder & R. Felder, 1867)
 - Adelpha lycorias lycorias* (Godart, [1824])
 - Adelpha cocala caninia* Fruhstorfer, 1915
- Heliconiinae (26)
- Argynnini (1)
 - Euptoieta hegesia meridiana* Stichel, 1938
 - Acræini (10)
 - Actinote pellenea pellenea* Hübner, [1821]
 - Actinote brylla* Oberthür, 1917
 - Actinote carycina* Jordan, 1913
 - Actinote pyrrha pyrrha* (Fabricius, 1775)
 - Actinote melanisans* Oberthür, 1917
 - Actinote genitrix* D'Almeida, 1922
 - Actinote parapheles* Jordan, 1913
 - Actinote canutia* (Hopffer, 1874)
 - Actinote discrepans* D'Almeida, 1958
 - Actinote surima surima* (Schaus, 1902)
- Heliconiini (15)
- Agraulis vanillae maculosa* (Stichel, [1908])
 - Dione juno juno* (Cramer, 1779)
 - Dione moneta moneta* (Hübner, [1825])
 - Dryadula phaetusa* (Linnaeus, 1758)
 - Dryas iulia alcionea* (Cramer, 1779)
 - Philaethria dido dido* (Linnaeus, 1763)
 - Philaethria wernickei* (Röber, 1906)
 - Eueides aliphera aliphera* (Godart, 1819)
 - Eueides isabella dianasa* (Hübner, [1806])
 - Eueides pavana* Ménétriés, 1857
 - Heliconius besckei* Ménétriés, 1857
 - Heliconius erato phyllis* (Fabricius, 1775)
 - Heliconius ethilla narcaea* Godart, 1819
 - Heliconius numata robigus* Weymer, 1875
 - Heliconius sara apseudes* (Hübner, [1813])
- Hesperiodea – Hesperiidae (198)
- Pyrrhopyginae (5)
- Myscelus amystis epigona* Herrich-Schäffer, 1869
 - Myscelus santhilarius* (Latreille, [1824])
 - Elbella* sp.
 - Elbella lamprus albociliata* Mielke, 1995
 - Pyrrhopuge charybdis semita* Evans, 1951
- Pyrginae (80)
- Eudaminae (38)
- Aguna asander asander* (Hewitson, 1867)
 - Astraptes creteus siges* (Mabille, 1903)
 - Astraptes chiriquensis oenander* (Hewitson, 1876)
 - Astraptes fulgerator fulgerator* (Walch, 1775)
 - Astraptes talus* (Cramer, 1777)
 - Autochton longipennis* (Plötz, 1882)
 - Autochton neis* (Geyer, 1832)
- Autochton reflexus (Mabille & Boullet, 1912)
- Autochton zarex (Hübner, 1818)
- Bungalotis midas (Cramer, 1775)
- Celaenorhinus eligius punctiger (Burmeister, 1878)
- Celaenorhinus similis Hayward, 1933
- Cephise cephise (Herrich-Schäffer, 1869)
- Chioides catillus catillus (Cramer, 1779)
- Euryphellus euribates polygius (Latreille, [1824])
- Drepanalys phoenicoides (Mabille & Boullet, 1919)
- Epargyreus exadeus exadeus (Cramer, 1779)
- Nascus paulliniae (Sepp, [1842])
- Nascus phocus (Cramer, 1777)
- Phanus australis L. D. Miller, 1965
- Phocides pigmalion pigmalion (Cramer, 1779)
- Phocides polybius phanias (Burmeister, 1880)
- Polygonus leo pallida Röber, 1925
- Polygonus savigny savigny (Latreille, 1824)
- Polythrix caunus (Herrich-Schäffer, 1869)
- Polythrix octomaculata (Sepp, [1844])
- Proteides mercurius mercurius (Fabricius, 1787)
- Salatis salatis (Stoll, 1782)
- Typhedanus undulatus (Hewitson, 1867)
- Urbanus albimargo takuta Evans, 1952
- Urbanus dorantes dorantes (Stoll, 1790)
- Urbanus doryssus albicuspis (Herrich-Schäffer, 1869)
- Urbanus esmeraldus (Butler, 1877)
- Urbanus esta Evans, 1952
- Urbanus procne (Plötz, 1880)
- Urbanus proteus proteus (Linnaeus, 1758)
- Urbanus simplicius (Stoll, 1790)
- Urbanus teleus (Hübner, 1821)
- Pyrgini (42)
- Achyliodes busirus rioja Evans, 1953
 - Achyliodes mithridates thraso (Hübner, [1807])
 - Anastrus obscurus Hübner, [1824]
 - Anastrus sempiternus simplicior (Möschler, 1877)
 - Anastrus tolimus robigus (Plötz 1884)
 - Camptopleura janthinus (Capronnier, 1874)
 - Chiomara asychis autander (Mabille, 1891)
 - Milanion clito (Fabricius, 1787)
 - Cycloglypha tisias (Godman & Salvin, 1896)
 - Cycloglypha thrasibus thrasibus (Fabricius, 1793)
 - Ebrietas anacreon anacreon (Staudinger, 1876)
 - Gesta gesta (Herrich-Schäffer, 1863)
 - Gorgythion plautia (Möschler, 1877)
 - Gorgythion begga begga (Prittitz, 1868)
 - Gorgythion beggina escalophoides Evans, 1953
 - Grais stigmaticus stigmaticus (Mabille, 1833)
 - Helias phalaenoides palpalis (Latreille, [1824])
 - Heliopetes omrina (Butler, 1870)
 - Milanion leucaspis (Mabille, 1878)
 - Mylon maimon (Fabricius, 1775)
 - Nisoniades castolus (Hewitson, 1878)
 - Nisoniades bipuncta (Schaus, 1902)
 - Nisoniades sp.
 - Ouleus accendens accendens (Mabille, 1895)
 - Pellicia costimacula costimacula Herrich-Schäffer, 1870
 - Polyctor polyctor polyctor (Prittitz, 1868)
 - Pyrgus orcyoides (Giacomelli, 1928)
 - Pyrgus orcus (Stoll, 1780)

- Pythonides lancea* (Hewitson, 1868)
Pythonides tullia Evans, 1953
Quadrus cerialis (Stoll, 1782)
Sostrata bifasciata bifasciata (Ménétriés, 1829)
Sostrata cronion (C. Felder & R. Felder, 1867)
Spathilepia clonius (Cramer, 1775)
Spioniades artemides (Stoll, 1782)
Staphylus incisus (Mabille, 1878)
Telemiades antiope antiope (Plötz, 1882)
Telemiades avitus (Stoll, 1781)
Telemiades epicalus Hübner, [1819]
Trina geometrina geometrina (C. R. Felder & R. Felder, 1867)
Xenophanes tryxus (Stoll, 1780)
Zera zera zera (Butler, 1870)
- Heteropterinae (1)
Dalla diraspes (Hewitson, 1877)
- Hesperiinae (112)
Anatrytone mella (Godman, 1900)
Anthoptus epictetus (Fabricius, 1793)
Argon lota (Hewitson, 1877)
Arita arita (Schaus, 1902)
Artines aquilina (Plötz, 1882)
Callimormus saturnus (Herrich-Schäffer, 1869)
Callimormus corades (C. Felder, 1862)
Carystoides basoches (Latreille, [1824])
Carystus phorcus claudianus (Latreille, 1824)
Cobalopsis nero (Herrich-Schäffer, 1869)
Cobalopsis miaba (Schaus, 1902)
Cobalus virbius hersilia (Plötz, 1882)
Conga chydaea (Butler, 1877)
Corticea corticea (Plötz, 1882)
Corticea lysias potex Evans, 1955
Cymaenes tripunctus theogenis (Capronnier, 1874)
Cymaenes uruba uruba (Plötz, 1886)
Cymaenes gisca Evans, 1955
Cyneia bistrigula (Herrich-Schäffer, 1869)
Cyneia diluta (Herrich-Schäffer, 1869)
Cyneia trimaculata (Herrich-Schäffer, 1869)
Cyneia sp.1
Cyneia sp.2
Damas clavus (Herrich-Schäffer, 1869)
Decinea decinea decinea (Hewitson, 1876)
Euphyes leptosema (Mabille, 1891)
Eutychide physcella (Hewitson, 1866)
Eutychide olympia (Plötz, 1882)
Enosis schausi Mielke & Casagrande, 2002
Enosis uza pruinosa (Plötz, 1882)
Eutocus vetulus matildae (Hayward, 1941)
Flaccilla aecas (Stoll, 1781)
Hylephila phyleus phyleus (Drury, 1773)
Justinia kora (Hewitson, 1877)
Lamponia lamponia (Hewitson, 1876)
Ludens ludens (Mabille, 1891)
Lento lento (Mabille, 1878)
Lycas argentea (Hewitson, 1866)
Lychnuchoides ozias ozias (Hewitson, 1878)
Lychnuchus celsus (Fabricius, 1793)
Methionopsis ina (Plötz, 1882)
Metron chrysogastra hypodesma (Plötz, 1882)
Metron oropa (Hewitson, 1877)
- Miltomiges cinnamomea* (Herrich-Schäffer, 1869)
Mnasilus allubita (Butler, 1877)
Moeris striga striga (Geyer, 1832)
Morys compta compta (Butler, 1877)
Morys geisa geisa (Möschler, 1879)
Mucia zygia (Plötz, 1886)
Neoxeniades scipio scipio (Fabricius, 1793)
Niconiades cydia (Hewiston 1876)
Niconiades sp.
Naevolus orius orius (Mabille, 1883)
Nyctelius nyctelius nyctelius (Latreille, [1824])
Onophas columbaria distigma Bell, 1930
Orphe gerasa (Hewitson, 1867)
Orthos orthos hyalinus (Bell, 1930)
Panoquina evadnes (Stoll, 1781)
Panoquina hecebolum (Scudder, 1872)
Panoquina fusina viola Evans, 1955
Panoquina panoquinooides minima De Jong, 1983
Panoquina lucas lucas (Fabricius, 1793)
Panoquina ocola (W. H. Edwards, 1863)
Panoquina sp.
Papias phainis Godman, 1900
Paracarystus hypargyra (Herrich-Schäffer, 1869)
Paracarystus menestries menestries (Latreille, [1824])
Penicula cristatus (Bell, 1930)
Penicula subviridis (Plötz, 1886)
Perichares philetis aurina Evans, 1955
Perichares seneca seneca (Latreille, [1824])
Phanes almoda (Hewitson, 1866)
Phanes rezia (Plötz, 1882)
Pheraeus fastus (Hayward, 1939)
Polites vibex catilina (Plötz, 1886)
Pompeius pompeius (Latreille, [1824])
Psoralis stacara (Schaus, 1902)
Pyrrhopygopsis socrates socrates (Ménétriés, 1855)
Quasimellana nicomedes (Mabille, 1883)
Quinta cannae (Herrich-Schäffer, 1869)
Quinta locutia (Hewitson, 1876)
Repens repens Evans, 1955
Saliana esperi esperi Evans, 1955
Saliana saladin catha Evans, 1955
Saliana salius (Cramer, 1775)
Saliana triangularis (Kaye, 1914)
Saliana mathiolus (Herrich-Schäffer 1869)
Saliana sp.
Saturnus reticulata tiberius (Möschler, 1883)
Sodalia argyropila (Mabille, 1876)
Sodalia coler (Schaus, 1902)
Synale hylaspes (Stoll, 1781)
Synapte malitiosa anistia (Plötz, 1882)
Talides sergestus (Cramer, 1775)
Tisias quadrata quadrata (Herrich-Schäffer 1869)
Tirynthia conflua (Herrich-Schäffer 1869)
Thracides cleanthes cleanthes (Latreille, [1824])
Thracides nanea nanea (Hewitson, 1867)
Thracides phidon (Cramer, 1779)
Turesis complanula (Herrich-Schäffer, 1869)
Vacerra bonfilius (Latreille, [1824])
Vehilius stictomenes stictomenes (Butler, 1877)
Vertica verticalis verticalis (Plötz, 1882)
Vettius artona (Hewitson, 1868)

Vettius diversa diversa (Herrich-Schäffer, 1869)
Vettius lafrenaye lafrenaye (Latreille, [1824])
Vettius phyllus prona Evans, 1955
Vettius richardi (A. G. Weeks, 1906)

Vinius letis (Plötz, 1883)
Vinius tryhana istria Evans, 1955
Virga austrinus (Hayward, 1934)
Zariaspes mys (Hübner, [1808])