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Comparison of automatic and visual methods used for image segmentation in Endodontics: a microCT study

Abstract

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To calculate root canal volume and surface area in microCT images, an image segmentation by selecting threshold values is required, which can be determined by visual or automatic methods. Visual determination is influenced by the operator's visual acuity, while the automatic method is done entirely by computer algorithms. Objective: To compare between visual and automatic segmentation, and to determine the influence of the operator's visual acuity on the reproducibility of root canal volume and area measurements. Material and methods: Images from 31 extracted human anterior teeth were scanned with a µCT scanner. Three experienced examiners performed visual image segmentation, and threshold values were recorded. Automatic segmentation was done using the "Automatic Threshold Tool" available in the dedicated software provided by the scanner's manufacturer. Volume and area measurements were performed using the threshold values determined both visually and automatically. Results: The paired Student's t-test showed no significant difference between visual and automatic segmentation methods regarding root canal volume measurements (p=0.93) and root canal surface (p=0.79). Conclusion: Although visual and automatic segmentation methods can be used to determine the threshold and calculate root canal volume and surface, the automatic method may be the most suitable for ensuring the reproducibility of threshold determination.

Keywords: Dental pulp cavity. Threshold limit values. X-ray microtomography.

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Introduction

In Endodontics, it is important for some studies to have a morphometric analysis of teeth for evaluating aspects such as the shaping ability of endodontic instruments, simulated root canal abnormalities, decalcification and sectioning techniques. However, some techniques can have limitations that do not fulfill the requirements for some researchers²⁵.

Microtomography is an imaging modality with increasing application in dental research due to its non-destructive technology that enables visualization of anatomical structures at the micrometer level⁷. In Endodontics, microtomography allows for qualitative and quantitative three-dimensional analyses of root canals while maintaining root integrity^{12,17,21,22,28}. Also, the results obtained with this modality can be as good as those obtained with histological images for endodontic analyses^{8,32}. Since the knowledge of the root canals' anatomical subtleties is essential in Endodontics^{6,31}, there have been several studies measuring root canal surface and volume by microtomography^{1,3,10,12,13,21,25,27}.

To calculate root canal volume and surface area, one needs to begin with image segmentation. First, thresholding is applied to images, resulting in binarization (black and white). It is essential that the grey value thresholds for dental hard tissues and root canal spaces are carefully determined, as inadequacies in this step may result in an over- or underestimation of measurements²⁰.

The correct inclusion of the region of interest in the binarized image is influenced by the choice of the segmentation threshold values used⁸. These values will determine what will be considered white and thus included in the analysis, or black and thus excluded from the analysis.

Thresholds can be determined by visual or automatic methods. Visual determination is influenced by the operator's visual acuity, which results in a subjectivity bias. To overcome this problem, an automatic threshold determination has been proposed. However, it is unclear whether differences between these segmentation methods are indeed significant in the context of microtomography studies. Studies generally use the visual method of image segmentation; however, the software in which the analysis is performed allows automatic segmentation, which is an option not commonly used to perform this step of the analysis of microtomographic images. Thus, this study was set to compare visual and automatic segmentation methods and to determine the influence of the operator's visual acuity on the reproducibility of root canal volume and area measurements.

Material and methods

This research was approved by the local ethics committee (14905013.8.0000.5441/290.975).

Thirty-one extracted anterior and single-rooted maxillary and mandibular teeth with complete root formation, similar size and without any intracanal filling comprised the sample of this study. After chemical disinfection in a 2% glutaraldehyde solution for two hours, tooth crowns were sectioned near the cementoenamel junction, using a carborundum disc coupled to a metallographic cutter Isomet 1000[®] (Buehler Ltd, Lake Bluff, IL, USA). A wax base was made as a support for each tooth.

Images were captured with a SkyScan 1174 microCT unit (Bruker, Kontich, Belgium) and the scanning parameters are shown in Figure 1. After capturing, NRecon version 1.6.6.0 software (Bruker, Kontich, Belgium) was used for image reconstruction, applying a ring artefact correction (set at 4) and a 30% beam hardening correction.

Morphometric parameters were calculated with the CT-Analyzer software (Bruker, Kontich, Belgium). Slices starting from slightly coronal to the cementoenamel junction thru the apex were used to obtain measurements. Three experienced examiners, doctoral students in oral radiology with three years of experience in microtomography, after a calibration session for this analysis, performed visual image segmentation, and all threshold values were recorded. After 15 days, the images were re-evaluated, for manual and automatic methods. Automatic segmentation was done using the "Automatic Threshold Tool" available on CT-Analyzer as suggested by Otsu¹⁵ (1979). Threedimensional analyses were performed using the values

Output	50 kV
Eletrical current	800 µA
Voxel size	33.21 µm
Thick aluminum filter	0.5 mm
Rotation	360°
Rotation step	0.4°

Figure 1- Scanning parameters

determined with the visual and the automatic method.

Both visual and automatic segmentation (Figure 2) were applied to the root's dentin, since it is not possible to directly segment the root canal so that the canal is individualized as an unfilled cavity (Figure 3); instead, it maintains a shade of grey similar to that of the background. The same slices interval has been predetermined for each tooth, in which the first axial cut was determined immediately after the cementoenamel junction, and the last axial view of the tooth apex was determined as the last slice. After determination of the threshold limits for the root's dentin visually and automatically, it is necessary to use advanced tools (i.e., custom processing) before proceeding with the measurements of canal volume and area. The seven steps of the sequence used for custom processing were: 1- Reload the image; 2- Threshold (fill in the threshold values already determined); 3- Despeckle (Remove pores - By image borders - 2D - Image); 4- Bitwise operations (Region of interest - Copy - Image); 5- Reload the image; 6- Threshold (same as step 2); 7- Bitwise operations (Image = Region of interest - Sub - Image).

These steps were recorded and rerun for the other images. After processing, the resultant image should be representative of the morphology of the canal. However, other steps may be necessary with atresic canals. To create the 3D models (Figure 4), the processed images must be saved and loaded into the CTVox software (Bruker, Kontich, Belgium).

Volume and area can be determined by loading the saved images into CT-Analyzer, then proceeding with segmentation of the canal's space; since the image is already binarized at this point, subjectivity is not an issue. After binarization, the images had only two colors: white, which is included in the analysis, and black (background), which is excluded. After that, one can perform the 3D analysis and obtain the volume and area of root canals.

Root canals' volume and area for the thresholding values of each evaluator and for the automatic method were obtained for each tooth. The mean volume and area for the three evaluators were calculated, composing the visual method, and used for comparison with the automatic method. A paired Student's t-test was used to assess the influence of the methods on threshold determination and to identify any existing significant differences among the measurements obtained. The null hypothesis was that the method of threshold determination does not affect the measurement of root canal volume, considering an α =0.05. The Intraclass Correlation Coefficient (ICC) was calculated to evaluate intra- and inter-examiner agreement. Analyses were performed using MedCalc 15.8 (MedCalc Software, Ostend, Belgium).



Figure 2- Axial slices of the root selected by different segmentation methods. A: automatic method; B: visual method – Examiner # 1; C: visual method – Examiner # 2; D: visual method – Examiner # 3



Figure 3- Axial slices of the unfilled cavity of root canal obtained by different segmentation methods. A: automatic method; B: visual method – Examiner # 1; C: visual method – Examiner # 2; D: visual method – Examiner # 3



Figure 4- Image of a 3D model of the root canal after automatic segmentation

Results

The ICC for intra- and inter-examiner agreement for the visual method ranged from 0.47 to 0.914 and from 0.699 to 0.847, respectively, which is considered fair to excellent, according to Cicchetti² (1994). For the automatic method, the ICC was of 1.0, showing a perfect correlation between the analyses.

Mean and standard deviation of canal volume and canal surface using different segmentation methods are shown in Table 1.

There were no statistically significant differences between visual and automatic segmentation methods regarding root canal volume measurements (p=0.93) and root canal surface (p=0.79).

Discussion

In Endodontics, microtomography has been extensively used as a research tool and a gold standard to test and compare endodontic files and to assess the anatomy of root canals. This imaging modality is used to evaluate three-dimensional images^{18,28}, perform measurements^{5,14,19}, allow for image segmentation to

Table 1- Mean and standard deviation of canal volume and canal surface using different segmentation methods

	Canal volume (mm ³)	Canal surface (mm ²)
Automatic	2.85 (±1.29) ^a	26.43 (±6.19) ^b
Visual	2.88 (±1.26) ^a	26.95 (±8.26) ^b

Values followed by different letters (vertical) are significantly different (p<0.05)

evaluate the structures of interest^{11,23} or to determine root canal volume $^{4,16,24,26,29,30}.$

Dedicated software provided by the manufacturer of the microtomography unit is normally used to analyze the acquired images and determine the volume and area of root canals. However, several steps are necessary to obtain that information; among them, image segmentation, which can be achieved by visual or automatic methods, are indeed significant in the context of microtomography studies, because it is unclear whether there are differences between these segmentation methods. The image segmentation is a crucial phase for proper calculation of the volume and surface of a root canal. As microCT is a validated research method for root canal analyses^{8,32}, this study proposed to evaluate if there are differences in the methods used to perform image segmentation.

Since no study carried out to compare the two methods of image segmentation was found in the literature, a direct comparison with our results was not possible. Researchers have done segmentation by visual and automatic methods without knowing if there are differences between them in the evaluation of the images. While some studies employed visual segmentation prior to root canal volume measurement^{4,9,11,16,23}, others presented only the measurements without any description as to how image segmentation was performed^{10,25,29,30}. Alternatively, few researchers used automatic segmentation before calculating root canal volume²⁶.

The visual segmentation could be dependent on the examiner's sight and/or experience with image processing; thus, it is open to subjectivity. Still, the visual method allows the evaluator to control the segmentation process by determining the threshold from values of the segmented areas. Thus, it may be that for segmentation of more complex structures (for example, images of structures that present metal and, consequently, artifact), the visual method might allow a more accurate segmentation. However, in this study there were no differences in relation to automatic segmentation.

The automatic method is based on threshold determination from image histograms. It is carried out by the microtomography analysis software, which enhances reproducibility by leaving subjectivity out of the picture. Yet, it is worth mentioning that intraand inter-examiner agreement levels were lower when applying visual segmentation in comparison with the automatic method, which showed a perfect reproducibility. It is known that reproducibility represents the constancy of the method and is an important advantage of the technique. To determine agreement, we used measurements for volume and area with reasonable results, given that working with low values may overemphasize small differences among datasets. In addition, it might be easier and demand less time, since it is an automatic method. Thus, the automatic method is the most suitable for ensuring the reproducibility of threshold determination in endodontic studies. Nevertheless, further studies are needed to evaluate the reproducibility of segmentation using automatic threshold in other image segmentation tasks.

Conclusion

Visual and automatic segmentation methods can be used to determine the threshold and to calculate root canal volume and surface; however, the automatic method may be the most suitable for ensuring the reproducibility of threshold determination. It is up to the evaluator to choose the preferred method according to their experience and the available time to perform image segmentation.

References

1- Bjørndal L, Carlsen O, Thuesen G, Darvann T, Kreiborg S. External and internal macromorphology in 3D-reconstructed maxillary molars using computerized X-ray microtomography. Int Endod J. 1999;32(1):3-9.

2- Cicchetti DV. Guidelines, criteria, and rules of thumb for evaluating normed and standardized assessment instruments in psychology. Psychol Assess. 1994;6(4):284-90.

3- Dowker SE, Davis GR, Elliott JC. X-ray microtomography: nondestructive three-dimensional imaging for *in vitro* endodontic studies. Oral Surg Oral Med Oral Pathol Oral Radiol Endod. 1997;83(4):510-6.

4- ElAyouti A, Dima E, Judenhofer MS, Löst C, Pichler BJ. Increased apical enlargement contributes to excessive dentin removal in curved root canals: a stepwise microcomputed tomography study. J Endod. 2011;37(11):1580-4.

5- Freire LG, Gavini G, Branco-Barletta F, Sanches-Cunha R, Santos M. Microscopic computerized tomographic evaluation of root canal transportation prepared with twisted or ground nickel-titanium rotary instruments. Oral Surg Oral Med Oral Pathol Oral Radiol Endod. 2011;112(6):e143-8.

6- Grande NM, Plotino G, Gambarini G, Testarelli L, D'Ambrosio F, Pecci R, et al. Present and future in the use of micro-CT scanner 3D analysis for the study of dental and root canal morphology. Ann Ist Super Sanita. 2012;48(1):26-34. 7- Hamba H, Nikaido T, Sadr A, Nakashima S, Tagami J. Enamel lesion parameter correlations between polychromatic micro-CT and TMR. J Dent Res. 2012;91(6):586-91.

8- Jung M, Lommel D, Klimek J. The imaging of root canal obturation using micro-CT. Int Endod J. 2005;38(9):617-26.

9- Kim Y, Perinpanayagam H, Lee J-K, Yoo Y-J, Oh S, Gu Y, et al. Comparison of mandibular first molar mesial root canal morphology using micro-computed tomography and clearing technique. Acta Odontol Scand. 2015;73(6):427-32.

10- Lloyd A, Uhles JP, Clement DJ, Garcia-Godoy F. Elimination of intracanal tissue and debris through a novel laser-activated system assessed using high-resolution micro-computed tomography: a pilot study. J Endod. 2014;40(4):584-7.

11- Marending M, Schicht OO, Paqué F. Initial apical fit of K-files versus LightSpeed LSX instruments assessed by micro-computed tomography. Int Endod J. 2012;45(2):169-76.

12- Nielsen RB, Alyassin AM, Peters DD, Carnes DL, Lancaster J. Microcomputed tomography: an advanced system for detailed endodontic research. J Endod. 1995;21(11):561-8.

13- Oi T, Saka H, Ide Y. Three-dimensional observation of pulp cavities in the maxillary first premolar tooth using micro-CT. Int Endod J. 2004;37(1):46-51.

14- Oliveira MA, Alves LD, Pereira AG, Raposo LH, Biffi JC. Influence of flexion angle of files on the decentralization of oval canals during instrumentation. Braz Oral Res. 2015;29(1):1-6.

15- Otsu N. A threshold selection method from gray-level histograms. IEEE Trans Syst Man Cybern. 1979;9(1):62-6.

16- Ounsi HF, Franciosi G, Paragliola R, Al Hezaimi K, Salameh Z, Tay FR, et al. Comparison of two techniques for assessing the shaping efficacy of repeatedly used nickel-titanium rotary instruments. J Endod. 2011;37(6):847-50.

17- Paqué F, Boessler C, Zehnder M. Accumulated hard tissue debris levels in mesial roots of mandibular molars after sequential irrigation steps. Int Endod J. 2011;44(2):148-53.

18- Park JW, Lee JK, Ha BH, Choi JH, Perinpanayagam H. Threedimensional analysis of maxillary first molar mesiobuccal root canal configuration and curvature using micro-computed tomography. Oral Surg Oral Med Oral Pathol Oral Radiol Endod. 2009;108(3):437-42.

19- Pasqualini D, Bianchi CC, Paolino DS, Mancini L, Cemenasco A, Cantatore G, et al. Computed micro-tomographic evaluation of glide path with nickel-titanium rotary PathFile in maxillary first molars curved canals. J Endod. 2012;38(3):389-93.

20- Pauwels R, Jacobs R, Singer SR, Mupparapu M. CBCT-based bone quality assessment: are Hounsfield units applicable? Dentomaxillofacial Radiol. 2015;44(1):20140238.

21- Peters OA, Laib A, Rüegsegger P, Barbakow F. Three-dimensional analysis of root canal geometry by high-resolution computed tomography. J Dent Res. 2000;79(6):1405-9.

22- Peters OA, Paqué F. Root canal preparation of maxillary molars with the self-adjusting file: a micro-computed tomography study. J Endod. 2011;37(1):53-7.

23- Rödig T, Hausdörfer T, Konietschke F, Dullin C, Hahn W, Hülsmann M. Efficacy of D-RaCe and ProTaper Universal Retreatment NiTi instruments and hand files in removing gutta-percha from curved root canals - a micro-computed tomography study. Int Endod J. 2012;45(6):580-9. 24- Solomonov M, Paqué F, Kaya S, Adigüzel Ö, Kfir A, Yiğit-Özer S. Self-adjusting files in retreatment: a high-resolution micro-computed tomography study. J Endod. 2012;38(9):1283-7.

25- Stavileci M, Hoxha V, Görduysus Ö, Tatar I, Laperre K, Hostens J, et al. Evaluation of root canal preparation using rotary system and hand instruments assessed by micro-computed tomography. Med Sci Monit Basic Res. 2015;21:123-30.

26- Stern S, Patel S, Foschi F, Sherriff M, Mannocci F. Changes in centring and shaping ability using three nickel-titanium instrumentation techniques analysed by micro-computed tomography (μ CT). Int Endod J. 2012;45(6):514-23.

27- Suto K, Saka H, Ide Y. Three dimensional observation of the orifice portion of root canal in the maxillary deciduous second molars using micro-CT. Jap J Ped Dent. 2002;40:541-8.

28- Verma P, Love RM. A micro CT study of the mesiobuccal root canal morphology of the maxillary first molar tooth. Int Endod J. 2011;44(3):210-7.

29- Versiani MA, Pécora JD, Sousa-Neto MD. The anatomy of two-rooted mandibular canines determined using micro-computed tomography. Int Endod J. 2011;44(7):682-7.

30- Versiani MA, Pécora JD, Sousa-Neto MD. Root and root canal morphology of four-rooted maxillary second molars: a micro-computed tomography study. J Endod. 2012;38(7):977-82.

31- Vier-Pelisser FV, Dummer PMH, Bryant S, Marca C, Só MV, Figueiredo JA. The anatomy of the root canal system of three-rooted maxillary premolars analysed using high-resolution computed tomography. Int Endod J. 2010;43(12):1122-31.

32- Zaslansky P, Fratzl P, Rack A, Wu MK, Wesselink PR, Shemesh H. Identification of root filling interfaces by microscopy and tomography methods. Int Endod J. 2011;44(5):395-401.