

UNIVERSIDADE ESTADUAL DE CAMPINAS



INSTITUTO DE BIOLOGIA

RENATO SIMÕES CORDEIRO

**“Expressão de Receptores Androgênicos no Lóbulo Ventral
da Próstata do Gerbilo da Mongólia”**

Este exemplar corresponde à redação final
da tese defendida pelo(a) candidato (a)
Renato Simões Cordeiro
e aprovada pela Comissão Julgadora.

Tese apresentada ao Instituto de Biologia para obtenção do Título de Doutor em Biologia Celular e Estrutural, na área de Biologia Celular.

Orientador: Prof. Dr. Sebastião Roberto Taboga

Campinas, 2007

**FICHA CATALOGRÁFICA ELABORADA PELA
BIBLIOTECA DO INSTITUTO DE BIOLOGIA – UNICAMP**

C811e Cordeiro, Renato Simões
Expressão de receptores androgênicos no lóbulo ventral
da próstata do gerbilo da Mongólia / Renato Simões
Cordeiro. – Campinas, SP: [s.n.], 2007.

Orientador: Sebastião Roberto Taboga.
Tese (doutorado) – Universidade Estadual de
Campinas, Instituto de Biologia.

1. Próstata. 2. Receptores de andrógenos. 3.
Antiandrogênicos. 4. Imunohistoquímica. I. Taboga,
Sebastião Roberto. II. Universidade Estadual de
Campinas. Instituto de Biologia. III. Título.

(rcdt/ib)

Título em inglês: Androgen receptors expression in the Mongolian gerbil ventral prostate.

Palavras-chave em inglês: Prostate, Androgen receptors, Antiandrogens, Immunohistochemical.

Área de concentração: Biologia Celular.

Titulação: Doutor em Biologia Celular e Estrutural.

Banca examinadora: Sebastião Roberto Taboga, Monica Levy Andersen, Alan Peres Ferraz de Melo, Wilson de Melo Júnior, Sérgio Luis Felisbino.

Data da defesa: 30/07/2007.

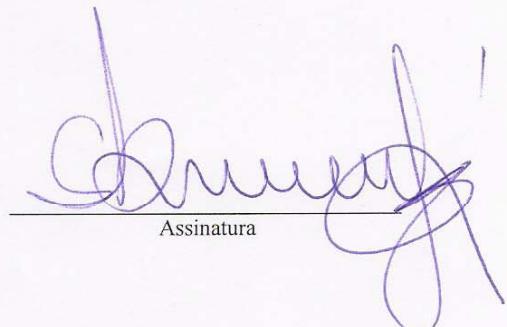
Programa de Pós-Graduação: Biologia Celular e Estrutural.

Campinas, 30 de julho de 2007.

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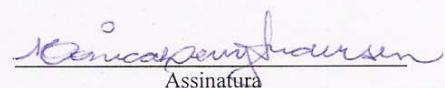


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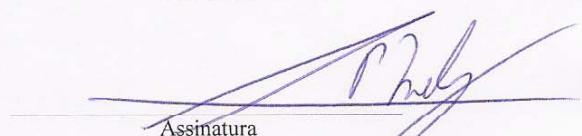
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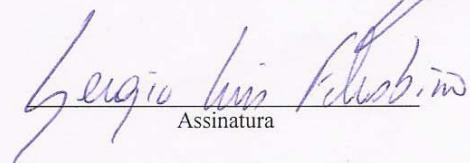
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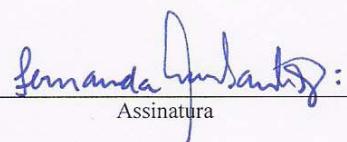
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DEDICATÓRIA

A Deus, por ter me capacitado, me dado forças, ânimo e coragem e principalmente pelo Seu eterno amor e abundante graça. A Ele seja toda glória, honra e louvores por mais esta vitória em minha vida.

Aos meus pais, pelo exemplo de humildade, amor, carinho e dedicação em minha vida. Obrigado por tudo, por todos os momentos terem me ajudado e incentivado a chegar até aqui. À vocês o meu eterno amor e admiração.

A minha irmã Ana Paula, a minha avó Antônia, a minha tia Jacira e tio Humberto, por terem me apoiado em todos os momentos, pelo vosso carinho e companheirismo.

A minha esposa, pela sua amizade, companheirismo, dedicação, carinho e amor e principalmente pelas suas eternas orações a meu favor. Obrigado por tudo que recebi de você todos estes anos, esta é mais uma vitória do nosso eterno amor.

A minha filhinha Débora, um presente de Deus em nossas vidas.

AGRADECIMENTOS ESPECIAIS

A Deus, por ter me escolhido e me resgatados aos seus braços, pelas ricas bênçãos derramadas sobre mim, pela Sua destra poderosa estar sobre a minha vida, pela vida eterna em Jesus Cristo e pela Sua eterna presença e graças pelo Seu Espírito em todos os momentos de sua vida. “E sabemos que todas as coisas contribuem juntamente para o bem daqueles que amam a Deus, daqueles que são chamados por seu decreto” Rm 8:28.

Aos meus pais, que em todos os momentos transmitiram segurança e confiaram na minha capacidade de chegar até aqui. A vocês, que muitas vezes, renunciaram aos seus sonhos para que realizássemos os nossos... A vocês, que apesar da distância, nos momentos mais difíceis sempre se fizeram presentes... A vocês, esta conquista, afinal, não é só minha, mas nossa.

A minha esposa e companheira Aline pelo seu eterno amor, pelo seu incentivo e preciosas orações que proporcionaram a obter as grandes vitórias em nossas vidas. A nossa filhinha Débora, herança do nosso eterno amor.

A minha irmã Ana Paula, tia Jacira e tio Humberto e a avó Antônia, por sempre estarem ao meu lado, dando apoio e carinho para o crescimento de minha vida.

À minha cunhada Alessandra e seu esposo Jorge, por terem me recebido com carinho e terem me amparado e apoiado em Campinas. Obrigado pelas suas amizades e companheirismo em minha vida.

Ao amigo, professor e orientador Dr. Sebastião Roberto Taboga,

Pela orientação, dedicação, paciência e persistência, nestes anos de aprendizado e luta. A você que apesar das inúmeras dificuldades encontradas, dispôs-se a me ajudar, caminhando por itinerários nunca dantes traçados, o meu muito obrigado.

Ao curso de Pós-Graduação em Biologia Celular e Estrutural do Instituto de Biologia, da Universidade Estadual de Campinas – Unicamp – pela oportunidade de incorporar-me e desenvolver-me, não só profissionalmente, em um dos melhores e mais bem conceituados cursos dessa categoria no país.

Antecipadamente, aos membros da Banca Examinadora Profa. Dra. Monica Levy Andersen, Prof. Dr. Alan Peres Ferraz de Melo, Profa. Dra. Fernanda C. Alcântara dos Santos e ao Prof. Dr. Sérgio Luis Felisbino, pela disponibilidade, análises e sugestões.

Aos docentes do curso e outras instituições, os quais compartilharam, humildemente comigo e outros colegas, seus conhecimentos, firmando meu aprimoramento acadêmico-profissional.

Ao Departamento de Biologia do IBILCE-UNESP de São José do Rio Preto, pela permissividade da realização dos meus trabalhos práticos de pós-graduação, assistência docente e participação em atividades.

A Liliam Alves Senne Panagio, pela amizade, presteza e, principalmente, pela paciência dedicada para o meu entendimento e resolução de problemas de ordem burocrática.

Aos antigos e novos amigos de São José do Rio Preto e do Laboratório de Microscopia e Microanálise, pelas inúmeras experiências vividas, momentos de risos, lágrimas, ansiedade, partilha de conhecimentos teórico-práticos específicos, auxílio em muitas atividades, e, sobretudo amigos de aprendizado e luta diária.... Ana Maria, Chicão, Fernanda, Manoel, Wellerson, Silvana, Lara, Cristiane, Maê e Ricardo. Ao técnico do Laboratório de Microscopia e Microanálise do Departamento de Biologia do IBILCE-UNESP de São José do Rio Preto, Luís Roberto, pelos auxílios na execução e aprendizado de técnicas e amizade.

A CAPES, pelo apoio financeiro destinado a esse trabalho e a outros que se fizeram necessários e complementares para meu aprimoramento profissional durante esse período.

Enfim, aqueles por vezes não citados, mas que, de alguma maneira, contribuíram para a minha formação, não necessariamente quanto profissional, mas, sobretudo como ser humano, que particularmente, julgo ser o essencial na vida....

Meu muito obrigado!

*“As coisas que o olho não viu, e o ouvido não ouviu,
e não subiram ao coração do homem
são as que Deus preparou para os que o amam”*

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I. RESUMO

O crescimento normal, a diferenciação e a manutenção da integridade morfológica da glândula prostática são dependentes das interações de concentrações constantes de andrógenos com seus receptores. A necessidade de se estudar esta glândula em resposta aos hormônios e o efeito do bloqueio destes, deve-se ao fato da próstata humana ser o sítio de um grande número de doenças relacionadas à idade, sendo que as de maior importância clínica são o câncer prostático e a hiperplasia prostática benigna, as quais podem ser tratadas por estratégias de remoção de andrógenos. Este estudo teve por objetivo a análise imuno-histoquímica do grau de expressão do receptor androgênico (RA) no lóbulo ventral prostático do gerbilo após terapias de bloqueios androgênicos. Setenta e cinco gerbilos machos foram distribuídos, aleatoriamente, em 3 grupos de 25 animais, cada grupo representando uma fase do desenvolvimento pós-natal: jovem, adulto e senil. Em cada fase foi realizada uma análise morfológica e estereológica dos compartimentos prostáticos, bem como a análise imuno-histoquímica da expressão do RA. Além disso, estabeleceu-se a dosagem hormonal das concentrações séricas de testosterona, como método para verificar a relação da quantidade desse andrógeno com a expressão dos RA. Os resultados demonstraram haver um padrão heterogêneo de distribuição dos RA no lóbulo ventral ao longo do desenvolvimento pós-natal, em que quanto mais jovem for o animal maior a interação de andrógenos estimulando a expressão de RA nos compartimentos prostáticos. As terapias de bloqueios androgênicos diminuíram a expressão de RA no lóbulo ventral e a reposição androgênica após esses bloqueios não apresentou o mesmo grau de intensidade de expressão de RA próximo às condições fisiológicas normais. A regulação e a distribuição do RA nos tecidos prostáticos do gerbilo são mecanismos complexos e que, provavelmente, são geneticamente regulados por andrógenos antes do nascimento ou por outros fatores ainda desconhecidos. O gerbilo parece ser um modelo valioso na tentativa de melhorar o conhecimento do comportamento morfofisiológico e patológico dessa importante glândula em humanos ao longo do envelhecimento e da formulação de novas idéias de terapias de combate ao câncer de próstata.

II. ABSTRACT

The normal growth, differentiation and maintenance of the morphofunctional integrity of the prostate gland are dependent on the interaction of constant levels of androgens with their receptors. The need to study the responses to hormones under several conditions and the effect of their blockage is due to the fact that the human prostate is the site of a great number of age-related diseases, and the ones with a major medical importance are prostate cancer and benign prostatic hyperplasia, which can both be treated with androgen suppression. The aim of this study was to analyze immunohistochemical degree of expression of androgen receptor (AR) of the ventral lobe of the gerbil prostate during different phases of the postnatal development employing different treatments for androgen blocking. Seventy-five male gerbils were distributed, randomly, into 3 groups of 25 animals each, where each group corresponded to one phase of postnatal development: young, adult and aged phase. In each phase, it was possible to morphologically and stereologically analyze the compartments of prostatic ventral lobe, as well as to immunohistochemically analyze the degree of expression of androgen receptor. In addition, it was possible to establish the hormonal dosage of serum testosterone concentrations given the comparative approach of the expression of androgen receptors. There is a heterogeneous pattern of AR distribution in the prostatic ventral lobe throughout postnatal development, in which the younger animal is the higher, the interaction of circulating androgens that stimulate the AR expression in the compartments prostatics. The androgen blockage therapies decreased AR expression in the ventral lobe, but the androgen reposition after these blockages was not showed the same the degree of expression of androgen receptor near normal physiological conditions. The regulation and distribution of AR along the gerbil prostatic tissues are complex mechanisms that are likely to be genetically regulated by androgens prenatally or by other factors that are still unknown. The gerbil seems to be a valuable model in the attempt to improve the understanding of the morphophysiological and pathological behavior of this important gland in humans throughout aging and to stimulate new therapeutic ideas to fight prostate cancer.

III. INTRODUÇÃO

Aspectos gerais do desenvolvimento e da fisiologia prostática

A próstata é uma glândula acessória do trato reprodutor masculino responsável pela produção de nutrientes, gradientes iônicos e pH ótimo para os espermatozóides no fluido seminal (Untergasser *et al.*, 2005).

A próstata origina-se a partir do epitélio do seio urogenital, servindo como glândula sexual e uretral (Dounjacour e Cunha, 1988, Cooke *et al.*, 1991, Marker *et al.*, 2003, Untergasser *et al.*, 2005). Ela é uma glândula túbulo-alveolar composta com atividade secretora principalmente ligada à sua parte alveolar (Reese *et al.*, 1986). O epitélio prostático apresenta ao menos três tipos celulares distintos, os quais podem ser diferenciados por suas características morfológicas e funcionais em células: luminais secretoras, basais e neuroendócrinas (Fig.1) (Abate-Shen e Shen, 2000; Garraway *et al.*, 2003).

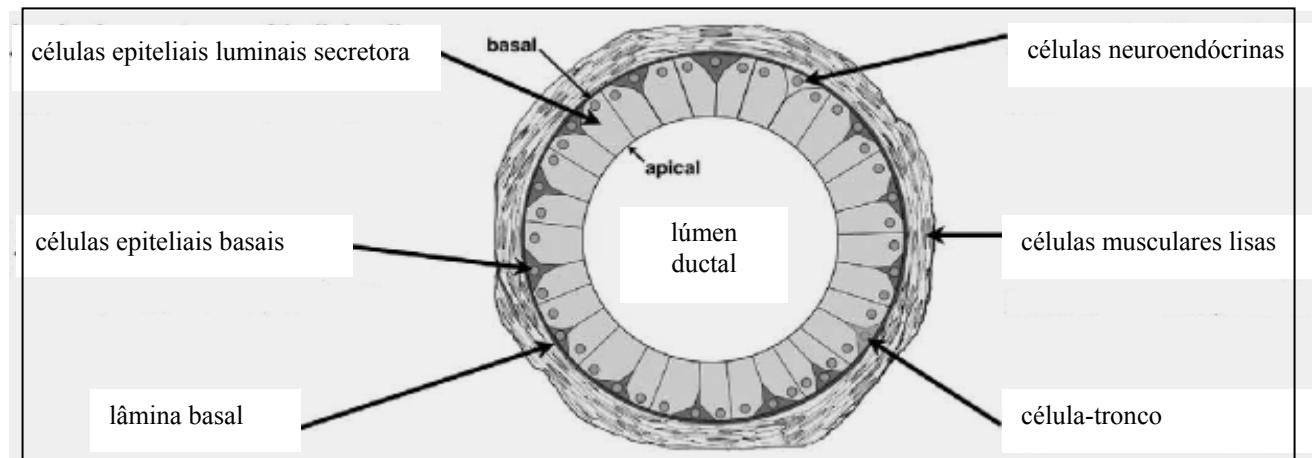


Fig. 1 – Esquema de um ducto prostático em corte transversal, indicando os diferentes tipos celulares (Marker *et al.*, 2003).

Entremeando as partes glandulares existe um estroma ricamente vascularizado, com esparsas fibras conjuntivas, células musculares lisas que têm papel contrátil durante a ejaculação, fibroblastos, macrófagos fixos, além de nervos e terminações nervosas, cada

qual com seu papel importante e específico na viabilidade e função secretora do tecido como um todo. Envolvendo este órgão tem-se uma fina cápsula fibromuscular, que confere diferentes formas ao órgão nos diversos mamíferos já estudados (Carvalho e Line, 1996 e Tuxhorn *et al.*, 2001).

Segundo Slayter e colaboradores (1994), no que se refere à homologia das partes da próstata humana com os lóbulos prostáticos de ratos, o lóbulo ventral em rato corresponde à zona mediana da próstata humana enquanto o lóbulo dorsal corresponde à zona posterior. Partindo dessas homologias morfo-funcionais pré-estabelecidas pode-se, então, buscar condições experimentais, em que a análise das respostas do epitélio e dos componentes do estroma são avaliadas. Assim, existem na literatura trabalhos relacionados à ação de agentes químicos (carcinógenos) na próstata, como os trabalhos de Pollard e Luckert (1987) e ação de agentes endógenos (hormonais) como os trabalhos desenvolvidos por Carvalho *et al.*, (1996, 1997a,b) e Scarano *et al.*, 2006, em que os resultados sugerem similaridade com a espécie humana.

O crescimento normal, a diferenciação e a manutenção da integridade funcional (secretora) e estrutural da próstata e dos demais órgãos acessórios do sistema reprodutor masculino são dependentes de concentrações constantes de andrógenos circulantes e ocorrem através de interações recíprocas entre o mesênquima e o epitélio (Cunha, 1976, Cunha *et al.*, 1996, Hayward *et al.*, 1997, Thomson *et al.*, 1997, Banerjee *et al.*, 2001; Taplin e Ho, 2001; Debes e Tindall, 2002).

Durante a vida fetal, existe um período em que a dependência de andrógenos é absoluta, sendo que sua remoção neste período suprime a formação da próstata (Wells, 1954, Sugimura *et al.*, 1986, Dounjacour e Cunha, 1988).

Nemeth e Lee (1996) e Sugimura *et al* (1986) sugerem que o estroma seja o primeiro alvo da ação androgênica, sendo a reação do epitélio mediada por fatores estromais. Certos efeitos do estroma sobre o epitélio podem ser simplesmente uma função da matriz extracelular produzida em grande parte pelas células estromais.

A necessidade de se estudar as respostas aos hormônios, sob várias condições e o efeito do bloqueio destes, deve-se ao fato de ser a glândula prostática em humanos, o sítio de um grande número de doenças relacionadas à idade, sendo que as de maior importância

clínica são o câncer prostático (CaP) e a hiperplasia prostática benigna (HPB) que se instalaram em resposta as descompensações hormonais. Os hormônios androgênicos e seus receptores, entre outros fatores, exercem papel na etiologia dessas lesões, as quais podem ser tratadas por estratégias de remoção de andrógenos (Droller, 1997, Rauch *et al.*, 1997, Cordeiro *et al.*, 2004, Corradi *et al.*, 2004; Oliveira, 2005).

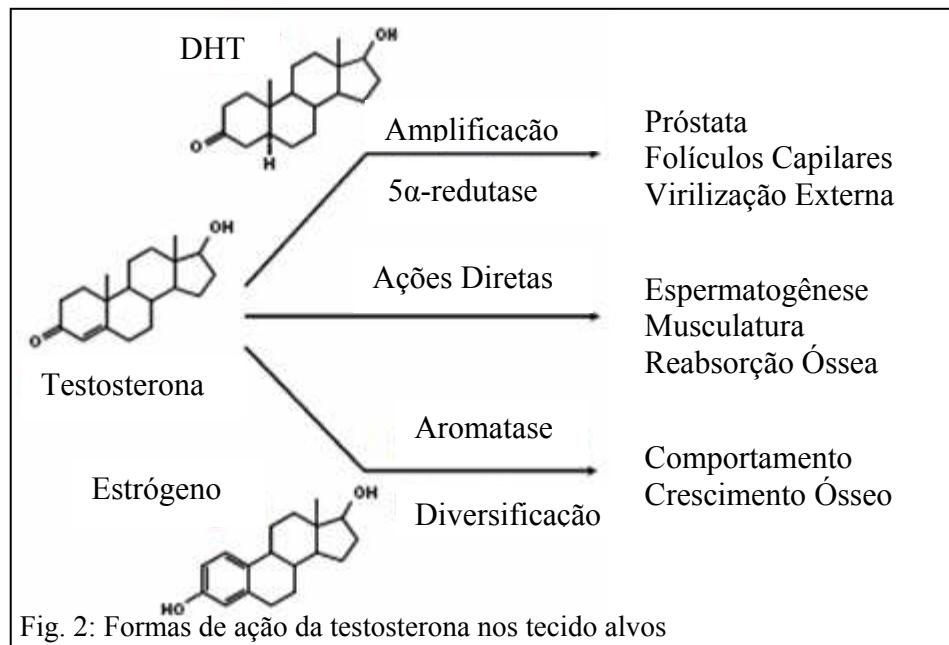
Ação dos andrógenos e anti-androgênicos

Os andrógenos regulam o crescimento, a diferenciação e a morte celular programada em células alvo via interação com o receptor androgênico (RA), os quais são fatores de transcrição que regulam a expressão de genes-alvo por mecanismo dependente do ligante (Cunha *et al.*, 1986; Nevalainen, 1997; Brum *et al.*, 2005). A produção de andrógenos é regulada pelo eixo hipotalâmico-hipofisário-gonadal, sendo que, o principal andrógeno é a testosterona, com as células de Leydig dos testículos produzindo mais de 95% e a glândula adrenal menos de 5% desse (Debes e Tindall, 2002; Hsing *et al.*, 2002).

Na próstata, a testosterona é convertida pela ação da enzima 5- α -redutase tipo 2, em 5- α -di-hidrotestosterona (DHT), a forma androgênica mais ativa, tendo uma afinidade maior pelo RA do que pela testosterona (Galbraith e Duchesne, 1997; Hsing *et al.*, 2002; Dehm e Tindall, 2006). Caso haja um bloqueio na conversão de testosterona para DHT durante o desenvolvimento prostático há uma acentuada redução da morfogênese e crescimento do órgão ou até mesmo importantes modificações no estroma dos indivíduos adultos quando tratados com o inibidor desta enzima (Marker *et al.*, 2003, Corradi *et al.*, 2004).

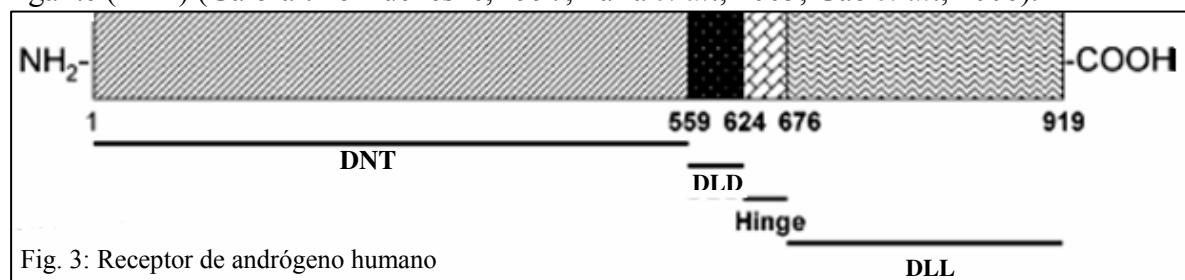
Segundo Dehm e Tindall (2006), existem três formas de ação da testosterona nos tecidos alvos (Fig. 2):

- 1º- Pode difundir através da membrana plasmática das células-alvo e se ligar diretamente ao RA ativando-os;
- 2º- Se converter em DHT pela enzima 5 α -redutase antes de se ligar ao RA;
- 3º- Ou ser aromatizada para estrógeno e atuar através dos receptores de estrógeno.



As células alvos para andrógenos estão localizadas em muitas partes do corpo, com altas concentrações de RA nos órgãos acessórios masculinos, como por exemplo, folículo capilar, pele genital, epidídimos, glândula seminal, próstata, bem como no hipotálamo (Brinkmann *et al.*, 1999, Santos *et al.*, 2004, Farla *et al.*, 2005, Dehm e Tindall, 2006).

O RA é uma fosfoproteína solúvel de 110 kDa, pertencente a uma superfamília de receptores esteróides que funciona como fator de transcrição intracelular. Estruturalmente o RA contém três domínios funcionais (Fig. 3): Domínio N-Terminal (DNT), Domínio Central de Ligação ao DNA (DLD) e Domínio C-Terminal de ligação ao ligante (DLL) (Galbraith e Duchesne, 1997, Farla *et al.*, 2005, Gao *et al.*, 2006).



Em estágio desligado ou inativo, o RA existe no citoplasma formando um complexo que inclui chaperonas e co-chaperonas (família de proteínas *heat shock*, Hsp 90, Hsp70 e Hsp 56). Após a ligação com andrógeno ocorrem alterações na conformação e

composição deste complexo, o receptor então sofre dissociação das chaperonas, dimerização e fosforilação. Em seguida, o RA é translocado para núcleo dentro de 15-60 minutos da entrada do andrógeno na célula, ligando-se aos elementos de resposta androgênica do DNA, promovendo ou estimulando a expressão de genes alvos (Fig. 4) (Hughes *et al.*, 2001; Heinlein e Chang, 2004; Farla *et al.*, 2005). A entrada dessa proteína-RA no núcleo pode ocorrer através de difusão simples pelos poros nucleares ou pode ligar-se a sinais de localização nuclear que resultará na ativação do transporte (Jenster *et al.*, 1993).

A expressão gênica do RA pode ser modulada em múltiplos níveis por mecanismos de transcrição, pós-transcrição e pós-tradução. Além disso, complexos formados por co-ativadores ou co-repressores, podem atuar como moléculas adaptadoras, ligando-se diretamente aos receptores nucleares, recrutando proteínas adicionais e interagindo com a maquinaria transcrecional basal para aumentar a transcrição de genes-alvo (Brum *et al.*, 2005).

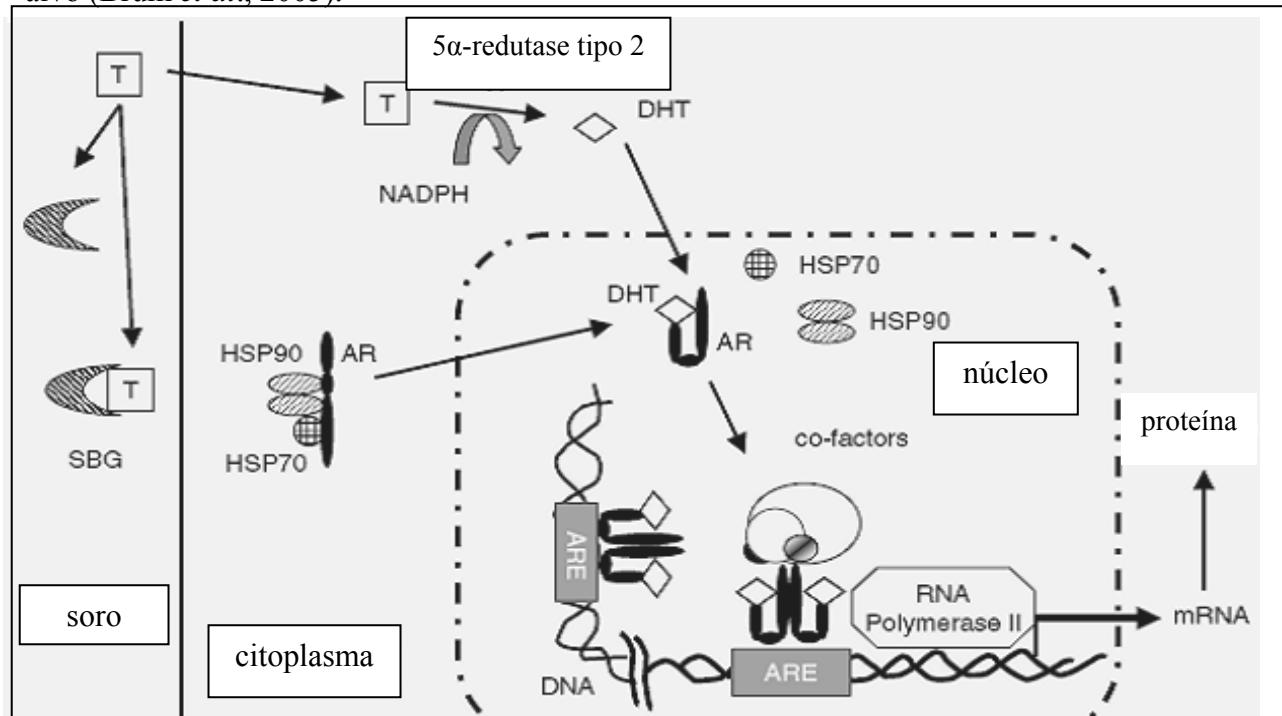


Fig. 4. Mecanismo de ligação do RA. ARE: elemento de resposta androgênica; DHT: di-hidrotestosterona; HSP: protein heat shock; SBG: globulina de ligação a hormônios sexuais; T: testosterona.

Embora a função dos andrógenos seja importante, esta é insuficiente para manter a homeostase prostática. Esse processo também requer interações entre fatores de crescimento peptídicos que são secretados por células epiteliais e estromais de maneira autócrina ou parácrina, atuando no controle de proliferação e morte celular, em ambos os compartimentos celulares (Galbraith e Duchesne, 1997; Untergasser *et al.*, 2001; Lee e Chang, 2003).

Em células epiteliais da próstata humana há uma baixa, porém balanceada, taxa de proliferação e morte celular (<0,20% dia), o que resulta em um estado estável do órgão sem crescimento líquido, com as células sendo renovadas continuamente (Berges *et al.*, 1995). Há um grande interesse em compreender melhor a biologia prostática devido à sua alta propensão em desenvolver tumores malignos, estando entre as mais comuns, as neoplasias que acometem o homem (Bonkhoff e Remberger, 1996; Abate-Shen e Shen, 2000).

A próstata humana é dividida em três discretas regiões, definidas pela sua localização em relação à uretra. As células de cada região variam significativamente no desenvolvimento de HPB e CaP. O CaP é considerado uma doença com grande heterogeneidade, englobando a existência simultânea de entidades patológicas múltiplas como neoplasia intra-epitelial (PIN), adenocarcinomas dependentes e independentes de andrógenos e lesões metastáticas em um mesmo tecido (Droller, 1997; Tang e Porter, 1997; Lucia *et al.*, 1998; Bostwick *et al.*, 2000). Enquanto muitos carcinomas prostáticos retêm um padrão de crescimento baixo, aproximadamente um terço torna-se invasivo localmente ou produz metástases, expandindo-se para os linfonodos locais e órgãos distantes como ossos, fígado e pulmão (Shulz *et al.*, 2003).

No início, o crescimento e a sobrevivência das células alteradas dependem da ação androgênica, em que a castração e/ou a administração de uma droga anti-androgênica seria a melhor alternativa como terapia. Inicialmente o bloqueio androgênico resulta da inibição dos RA, indicado pela redução da expressão de determinados genes alvos e consequentemente redução do tumor (Feldman e Feldman, 2001; Dehm e Tindall, 2006).

Contudo, o CaP pode reincidir em uma forma que é resistente às manipulações hormonais de tratamentos, sendo referido como uma doença andrógeno-independente.

Embora o CaP seja resistente à tentativa de ação do bloqueio androgênico, o RA permanece um fator crítico para crescimento e sobrevida da maioria dos tumores e que, portanto o eixo hormonal de expressão do RA permanece um valioso alvo à terapia (Buchanan *et al.*, 2001).

A HPB é a doença mais prevalente da próstata, com aproximadamente 50% dos homens apresentando evidências histológicas aos 50 anos e 90% aos 80 anos de idade. É considerada uma doença progressiva, definida como o crescimento contínuo da próstata, caracterizando-se por uma predominante proliferação estromal e, embora um aumento substancial do epitélio também ocorra, a integridade regional da glândula é mantida. Esta doença leva à intensificação de sintomas e ao aumento do risco de complicações ao longo do tempo, como a retenção urinária aguda e necessidade de procedimentos cirúrgicos.

As ações androgênicas têm sido amplamente demonstradas na próstata humana, seja por sua ação na morfogênese, diferenciação, proliferação celular e secreções da glândula, ou pela resposta ao tratamento hormonal do CaP e HPB. Contudo, os mecanismos moleculares de transformação neoplásica e carcinogênese ainda não estão bem estabelecidos (Krieg *et al.*, 1993; Bonkhoff e Remberger, 1996; Raghaw *et al.*, 2000; Banerjee *et al.*, 2001; Leav *et al.*, 2001; Brum *et al.*, 2005).

Alterações nas concentrações endógenas de hormônios esteróides relacionados ao envelhecimento, também promovem alterações prostáticas em outras espécies de mamíferos. Em ratos *Brown Norway* senis o crescimento espontâneo da próstata é confinado aos lóbulos laterais e dorsais, sendo, portanto, lóbulo-específico (Banerjee *et al.*, 2001). Em cães também há uma propensão correlacionada com o envelhecimento ao aparecimento de lesões proliferativas espontâneas na próstata (Leav *et al.*, 2001), bem como em primatas (McEntee *et al.*, 1996).

Os andrógenos têm papel fundamental na carcinogênese prostática. Partindo desse princípio, a inibição de vias hormonais permanece como a primeira medida para a prevenção dessa lesão (Lee e Chang, 2003). A supressão das concentrações séricas de testosterona e DHT pode ser realizada por orquiectomia (castração cirúrgica) e/ou administração de drogas antiandrogênicas (Ruijter *et al.*, 1999; Corradi *et al.*, 2004; Cordeiro, *et al.*, 2004).

Após a orquiectomia bilateral, as concentrações séricas de testosterona diminuem de 90 a 95% (Heinlein e Chang, 2004). Em ratos, o lóbulo ventral prostático e a glândula seminal involuem, perdendo aproximadamente 90% de suas células após 21 dias de castração (Meeker *et al.*, 1996; Carvalho *et al.*, 1997a,b). A regressão da glândula ocorre devido à ativação do programa de morte celular apoptótica, a qual afeta, principalmente, células epiteliais glandulares. As células epiteliais basais e estromais são menos sensíveis a supressão androgênica e são mantidas (Bruyninx *et al.*, 1999; Balk, 2002).

Sugimura e colaboradores (1986) demonstraram que a administração de andrógenos pós-castração promove a restauração da estrutura ductal, semelhante à condição normal. Em ratos *Brown Norway*, a apoptose induzida pela castração é lóbulo-específica e uma diminuição nesse processo celular foi observada com o envelhecimento, sugerindo uma evolução para independência a andrógeno relacionada com a senescência nos quatro lóbulos prostáticos (Banerjee *et al.*, 2001). Esses eventos indicam que há uma variação quanto à dependência androgênica nas regiões prostáticas, bem como, entre as células epiteliais, podendo haver outros fatores que permitam a sobrevivência de grupos de células independentes de hormônio.

Para Scher e colaboradores (2004), o sucesso do tratamento anti-androgênico está vinculado à dependência das células à DHT, embora as concentrações hormonais sejam baixas no sangue de pacientes com tumores em progressão pós-castração, a concentração androgênica intratumoral pode ser suficiente para manutenção da lesão. Desta maneira, tumores prostáticos, aparentemente, não se encontram em um ambiente completamente independente de andrógenos após castração (Mohler *et al.*, 2004).

Embora, as concentrações plasmáticas de testosterona declinem após a castração, vários estudos indicam que a produção intraprostática de DHT continua a ocorrer, por meio da conversão de andrógenos adrenais. Desta forma, a DHT está presente mesmo após supressão androgênica, sendo necessários procedimentos terapêuticos adicionais para que o bloqueio hormonal seja efetivo (Nishiyama *et al.*, 2004).

A utilização de medicamentos chamados anti-androgênicos esteroidais e não-esteroidais (castração química) no tratamento de lesões prostáticas têm sido muito indicada,

principalmente nas lesões proliferativas adenocarcinomatosas que dependem desses hormônios, e, além disso, parece ser mais eficiente que a orquiectomia, uma vez que mesmo após a depleção hormonal, a glândula adrenal ainda produz cerca de 5% de hormônios androgênicos (Brooks *et al.*, 1991).

As drogas anti-androgênicas inibem ou diminuem a síntese e/ou a liberação de hormônios hipotalâmicos e de seus fatores de liberação e de hormônios da hipófise anterior (gonadotropinas), inibindo assim a biossíntese ou a secreção de andrógenos e, consequentemente, suas ações sobre os tecidos alvos e, além disso, essas drogas atuam na inibição da união dos andrógenos aos respectivos receptores celulares (Galbraith e Duchesne, 1997; Taplin e Ho, 2001; Heinlein e Chang, 2004).

Uma característica particular da farmacocinética dos anti-androgênicos não-esteroidais, quando comparado com os esteroidais, é que eles proporcionam uma melhor eficiência no bloqueio androgênico para tratamento de CaP, além disso, estimulam a liberação de hormônio luteinizante (LH) e consequentemente há um aumento das concentrações de testosterona testiculares e por esta razão, muitos pacientes permanecem sexualmente potentes durante os tratamentos terapêuticos com anti-androgênicos não-esteroidais (Pompeu *et al.*, 1998; Singh *et al.*, 2000; Goto *et al.*, 2004; Foster e Harris, 2005).

A flutamida é um dos mais conhecidos anti-androgênicos não-esteroidais e atua como inibidor competitivo de testosterona e DHT na ligação com RA no tecido prostático (Fig. 5). Essa droga quando absorvida pelo trato gastrointestinal sofre alterações conformacionais, formando a 2-hidroxyflutamida, metabólito mais potente antiandrogênico *in vivo* e com alta afinidade de ligação para RA do que a flutamida. Esse metabólito inibe a ação da testosterona endógena e exógena, responsáveis pelo crescimento da próstata, atuando principalmente em nível citoplasmático por inibição competitiva da ligação da DHT em seu receptor, bem como possível bloqueio de translocação do complexo receptor-DHT para interior do núcleo inibindo assim, a síntese de DNA prostático estimulada pela testosterona resultando na involução da glândula (Metzger *et al.*, 1993; Singh *et al.* 2000; Miyata *et al.*, 2003; Singh *et al.*, 2006).

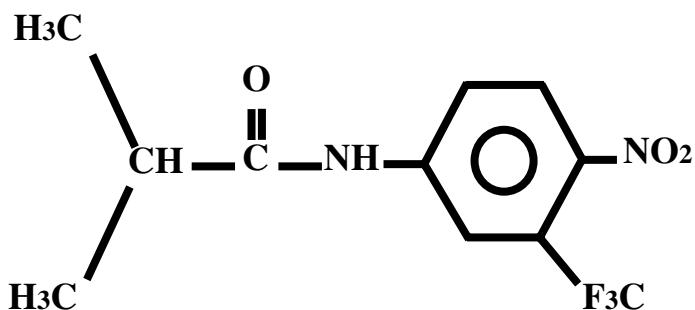


Figura 5: Estrutura química da Flutamida (Eulexin): *3α*-trifluoro-2-methyl-4'-nitro-m-propionotoluidide (Sufrin e Coffey, 1975).

Estudos mais recentes descrevem que o metabólito ativo da flutamida, ao se ligar ao RA provoca mudanças conformacionais, agindo como medida terapêutica no combate ao câncer de próstata. Esse anti-androgênico além de bloquear completamente a translocação do complexo receptor-DHT para interior do núcleo, também pode agir na translocação de RA, porém de forma mais lenta e incompleta, diminuindo assim a expressão de genes alvos (Farla *et al.*, 2005; Gao *et al.*, 2006).

A distribuição da flutamida nos tecidos foi examinada em ratos machos após a administração de 5 mg/kg de ^{14}C -flutamida. Enquanto a concentração da flutamida estava geralmente baixa em todos os tecidos examinados, o seu metabólito apresentava um aumento de 70 vezes após 6 horas da dosagem. O metabólito foi relativamente concentrado no lóbulo ventral prostático e na glândula seminal de ratos, demonstrando serem os órgãos alvos da atividade farmacológica da flutamida (Neri, 1989).

Vários trabalhos e ensaios experimentais demonstram que a administração diária de flutamida, em doses variando entre 1 a 50 mg/kg de peso corporal, reduz consideravelmente o peso e volume da próstata, e das glândulas seminais de ratos sem alterar a potência sexual (Neri, 1989; Sogani *et al.*, 1984; Srougi 1992). Narayan e colaboradores (1996), estudando 372 pacientes durante dois anos, demonstraram que 250mg de flutamida diária foi suficiente para reduzir de 14% a 29% o volume prostático.

A administração de 10 mg/kg/animal de flutamida durante 10 dias em cobaias macho ao longo do desenvolvimento pós-natal, promoveu alterações substanciais estruturais e ultra-estruturais nos componentes epiteliais e no padrão de distribuição e na

concentração dos componentes fibrilares, levando em conta a fase do desenvolvimento pós-natal (Cordeiro *et al.*, 2004).

Em camundongos transgênicos com adenocarcinoma (TRAMP), este anti-andrógeno em altas doses diminuiu a incidência de tumor e aumentou o período de latência, sugerindo ser uma medida preventiva viável (Raghaw *et al.*, 2000). Porém, a terapia prolongada com flutamida pode levar a seleção de células tumorais, as quais não sofrem a ação antiandrogênica e assim, contribuir para a progressão da lesão (Lee e Chang, 2003 e Foster e Harris, 2005). Isso poderia ocorrer provavelmente devido ao efeito estimulatório da Flutamida sobre a liberação de LH e consequente aumento dos níveis de andrógeno testicular, podendo haver dessa forma, um maior estímulo proliferativo destas células tumorais.

A depleção de andrógenos, seja por castração cirúrgica e/ou química, após a próstata ter atingido o seu tamanho final, causa à atrofia do órgão, com notável redução do seu tamanho e peso, inicialmente caracterizados por uma parada na síntese e uma acelerada liberação da secreção luminal, seguida pela diminuição do tamanho das células epiteliais por processos de morte e degeneração celular, resultando em lóbulos reduzidos e células epiteliais menores (Aumüller e Seitz, 1990). São observados ainda um declínio na síntese de DNA e de proteínas no conteúdo e na complexidade do RNA e uma diminuição do receptor de andrógeno. A castração na idade adulta leva à morte, predominantemente, as células epiteliais luminas (Hayward *et al.*, 1996; Carvalho *et al.*, 1997a,b; Vilamaior, 2000).

Uma diminuição de 48% no tamanho da glândula foi notada em homens nos seis primeiros meses de terapia com anti-andrógeno flutamida, enquanto que a combinação de castração cirúrgica com esta droga (bloqueio androgênico completo-BAC) promoveram uma diminuição prostática de 56% (Noldus *et al.*, 1996; Cordeiro *et al.*, 2004). Em ratos, a regressão do lóbulo ventral também foi mais acentuada após o tratamento combinado (Montalvo *et al.*, 2002). Assim, há uma redução de estimulação androgênica nas células anômalas uma vez que, estas apresentam dependência hormonal inicial (Raghaw *et al.*, 2000). Porém, essas terapias podem conduzir ao desenvolvimento de células insensíveis a andrógenos, as quais são resistentes aos sinais apoptóticos e resultam em doença

metastática (O'Neill, 2001). Tais eventos patológicos dificultam o encontro de uma solução terapêutica para a lesão e, equivalem a um ponto crucial na compreensão do potencial maligno do câncer de próstata (Shulz *et al.*, 2003).

Além disso, há ressaltar-se que a sinalização de RA é mantida ou regulada positivamente em tumores que retomam crescimento após falha de terapia androgênica e a ativação de genes reguladores de andrógenos é suficiente para facilitar a sobrevivência do tumor. Há evidências sugerindo que a seleção de mecanismos que facilitem a permanência da sinalização por RA, ou que ative novas vias de proliferação celular, podem depender da terapia utilizada durante o tratamento (Scher *et al.*, 2004).

Enquanto os eventos associados às modificações alcançadas pelas células epiteliais da parte glandular e no estroma foram intensamente investigados após a castração cirúrgica e química, as modificações morfológicas, bioquímicas e imuno-histoquímicas que ocorrem no epitélio e no estroma prostático, ainda necessitam de maiores investigações após efeito de aplicação de anti-androgênicos, principalmente frente à tentativa de melhor compreensão das relações andrógenos-receptores e epitélio-estroma.

Características do modelo experimental

Os estudos sobre a fisiologia da próstata têm despertado grande interesse para as Ciências Biomédicas, uma vez que este órgão ser alvo do desenvolvimento de graves lesões no homem, tanto na fase adulta quanto na senescência. Assim, é de fundamental importância que se tente estabelecer padrões comparativos com outros animais, para melhor compreender os mecanismos envolvidos em condições malignas nos homens.

Os gerbilos, também conhecidos com esquilos da Mongólia, são roedores da família Muridae, subfamília Gerbillinae (McKenna e Bell, 1997), provenientes das regiões áridas da China e Mongólia (Schwentker, 1963). Introduzidos por Vitor Schwentker, nas Américas em 1954 como nova proposta de animal experimental, ficaram durante muito tempo nas limitações dos Estados Unidos como animais de excelência para a pesquisa biomédica (Williams, 1974). Nas últimas décadas, esse animal vem sendo gradativamente introduzido nos biotérios das universidades brasileiras e têm assumido importante papel

nos experimentos biológicos e biomédicos, juntamente com outras espécies clássicas como *Rattus norvegicus* (rato) e *Mus musculus* (camundongo).

Na pesquisa científica é cada vez maior a utilização dos gerbilos na experimentação biomédica, principalmente nas áreas da imunologia (Jeffers *et al.*, 1984; Nawa *et al.*, 1994) e fisiologia (Muller e Nielsen, 1979) e também em estudos sobre o sistema genital (Santos *et al.*, 2003; Corradi *et al.*, 2004; Pinheiro *et al.*, 2004; Segatelli *et al.*, 2004). Os estudos a respeito do sistema genital masculino dessa espécie têm descrito as relações dos ductos das glândulas acessórias com a uretra (Williams, 1974; Pinheiro *et al.*, 2004) e os efeitos de bloqueios ou manipulações hormonais sobre a organização estrutural do lóbulo prostático ventral (Corradi *et al.*, 2004; Oliveira, 2005).

Nosso grupo de pesquisa adotou como modelo experimental para o estudo da próstata o gerbilo *Meriones unguiculatus*. De anatomia similar à do rato e camundongo, os gerbilos adultos de ambos os sexos variam entre 11,5 e 14,5 cm de comprimento corpóreo. Os machos adultos pesam em torno de 90-100g enquanto as fêmeas adultas pesam valores próximos a 85g (Kramer, 1964). Esses animais têm sido amplamente utilizados para estudos de natureza didático-científica, principalmente pelo fato de terem comportamento dócil em cativeiro. Outra característica importante a ser considerada sobre esses roedores é a facilidade de manutenção no biotério, pois, por serem de origem desértica, apresentam micção infreqüente, agilizando muito a limpeza das gaiolas no processo de manutenção, promovendo uma maior facilidade e plasticidade no manuseio e manutenção dos animais nos biotérios.

A glândula prostática dos gerbilos tem se mostrado semelhante à humana, no referente à compacidade e fusão de lóbulos. Entre as partes glandulares, encontra-se um estroma conjuntivo vascularizado, com poucas fibras conjuntivas e elásticas, além de abundantes células musculares lisas dispostas ao redor dos ductos prostáticos. Ultra-estruturalmente, o epitélio prostático apresenta heterogeneidade entre os seus tipos celulares e o estroma glandular mostra abundantes células musculares lisas entremeadas às camadas de colágeno (Williams, 1974; Zanetoni *et al.*, 2001). Além disso, o modelo vem apresentando respostas significativas em estudos sobre tratamentos hormonais (Santos *et al.*, 2003; Scarano *et al.*, 2004), drogas contra hiperplasia prostática humana (Corradi *et al.*,

2004; Cordeiro *et al.*, 2005), bem como, desenvolvimento de neoplasias espontâneas associadas ao envelhecimento (Zanetoni *et al.*, 2001). O surgimento dessas últimas está correlacionado com declínio nas concentrações séricas de testosterona nos animais senis (Zanetoni *et al.*, 2001). Assim, parece interessante a tentativa de se estabelecer novos modelos para estudos da próstata. E nestes, o gerbilo pode ser incluído como uma proposta valiosa na melhoria do conhecimento do comportamento morfofisiológico e patológico dessa importante glândula masculina.

Atualmente sabe-se que vários fatores hormonais são responsáveis pela manutenção da estrutura histofisiológica da próstata masculina (Reiter, 1999). No entanto, o conhecimento sobre o grau de expressão do RA na próstata normal e anormal, bem como após terapias de bloqueios androgênicos ainda é limitado. Desta forma, investigar as diferentes interações dos andrógenos com seus receptores nos compartimentos prostáticos ao longo do desenvolvimento pós-natal e o comportamento desses receptores em diferentes ambientes de ablação androgênica utilizados em tratamento de lesões prostáticas, permitirá um melhor entendimento da biologia prostática e possibilitará condutas terapêuticas no tratamento do CaP.

IV. OBJETIVOS

O presente estudo teve como objetivo avaliar o grau de expressão do receptor androgênico no lóbulo ventral da próstata do gerbilo *Meriones unguiculatus* (Muridae, Gerbillinae) nas diferentes idades do desenvolvimento pós-natal (jovens, adultos e senis) mediante diferentes terapias de bloqueios androgênicos. Além disso, foi realizado a dosagem hormonal da concentração sérica de testosterona e avaliação estereológica dos volumes ocupados pelo compartimento epitelial, luminal e estromal visto sua abordagem comparativa em relação à expressão do receptor de andrógeno.

V. ARTIGO

Título: “Androgen receptor expression in the Mongolian Gerbil ventral prostate: Evaluation during different phases of post-natal development and following androgen blockage”

Manuscript ID: 5224817051478325

Journal: Reproductive Biology and Endocrinology

Androgen receptors expression in the Mongolian Gerbil ventral prostate: Evaluation during different phases of post-natal development and following androgen blockage

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Abstract

Background: The normal growth, differentiation and maintenance of the morphofunctional integrity of the prostate gland are dependent on the interaction of constant levels of androgens with their receptors. The need to study the responses to hormones under several conditions and the effect of their blockage is due to the fact that the human prostate is the site of a great number of age-related diseases, and the ones with a major medical importance are prostate cancer and benign prostatic hyperplasia, which can both be treated with androgen suppression. The aim of this study was to analyze immunohistochemical degree of expression of androgen receptor (AR) of the ventral lobe of the gerbil prostate during different phases of the postnatal development employing different treatments for androgen blocking.

Methods: Seventy-five male gerbils were divided, randomly, into 3 groups of 25 animals, where each group corresponded to one phase of postnatal development: young, adult and old. In each phase, it was possible to morphologically and stereologically analyze the compartments prostatics, as well as to immunohistochemically analyze the degree of expression of AR after the androgen blockage therapies. In addition, it was possible to establish the hormonal dosage of serum testosterone concentration given the comparative approach of the expression of AR.

Results: There is a heterogenic pattern of AR distribution in the ventral lobe throughout postnatal development, in which the younger animal was the higher the interaction of circulating androgens that stimulate the AR expression in the compartments prostatics. The androgen blockage therapies decreased AR expression in the prostatic compartments, but the androgen reposition after these blockages was not sufficient to recover the glandular structure or stimulate the AR expression up to normal physiological conditions.

Conclusions: Both the regulation and distribution of androgen receptors along the gerbil prostatic tissues are complex mechanisms that are likely to be genetically regulated by androgens prenatally or by other factors that are still unknown. This rodent species seems to be a valuable model in the attempt to improve the understanding of the morphophysiological and pathological behavior of this important gland in humans throughout aging and to stimulate new therapeutic ideas to fight prostate cancer.

Background

Androgens regulate the growth, differentiation and programmed death of cells in the prostate gland and other male reproductive organs via interaction with the androgen receptor (AR). The main androgen is testosterone, which, inside the prostate, is converted by the action of type 2 5 α -reductase enzyme to 5 α -dihydrotestosterone (DHT), which is a more active androgen form that presents a higher affinity for AR than testosterone [1-6]. If there is a blockage in the conversion of testosterone to DHT during the prostatic development, an intense reduction in the morphogenesis and growth of the organ occurs or there are even important changes in the stroma of adult individuals when they are treated with the inhibitor of this enzyme [7, 8].

The AR, a member of the steroid receptor family that is activated by androgens, is the major regulatory transcription factor in normal development of the prostate and in the growth of androgen-dependent prostate cancer. Thus, AR is assumed to contribute to prostate cancer development during its recurrence in the androgen-deprived patient, but the function of AR in the development of androgen-independent and dependent prostate cancers is still unknown [5, 9, 10-12].

The need to study the responses to hormones under several conditions and the effect of their blockage is due to the fact that the human prostate is the site of a great number of age-related diseases, and the ones with a major medical importance are prostate cancer (PC) and Benign Prostatic Hyperplasia (BPH). Although PC is resistant to attempts at androgen blockage, AR remains a critical factor for the growth and survival of most of the tumors and, therefore, the hormonal axis of AR expression constitutes a valuable target for antiandrogenic therapy [4, 13-15]. These lesions, whether malignant or not, can be treated with androgen removal strategies [8, 16, 17]. The suppression of testosterone and DHT concentrations in the serum can be obtained through orchectomy (surgical castration) and/or treatment with antiandrogenic drugs [8, 17, 18].

Flutamide is one of the most well-known non-steroidal antiandrogens that act as competitive inhibitors of testosterone and DHT in the binding of AR with the prostatic tissue. This drug, once absorbed by the gastrointestinal tract, undergoes conformational

changes, forming the most active metabolite, 2-hydroxyflutamide, which is the most potent antiandrogenic agent *in vivo*, with higher AR binding affinity than flutamide [19, 20-23].

Androgen blockers cause a reduction of androgenic stimulation in the anomalous cells since they initially present hormonal dependency [24]. However, these therapies can cause the development of androgen-insensitive cells that are resistant to apoptotic signaling, resulting in metastatic disease [25]. Such pathological events make it difficult to find a therapeutic solution to the lesion and they constitute a key factor in understanding the malignant potential of prostate cancer [26].

Although the events associated with the changes undergone by epithelial cells in the glandular portion and in the stroma have been intensely investigated after either surgical or chemical castration, the morphological, biochemical and immunohistochemical changes that occur in both the epithelium and the prostatic stroma still require further investigation after the effects of antiandrogenic treatment, mainly in terms of trying to better understand the relationships between androgen and receptors as well as between epithelium and stroma.

Thus, the objective of this work is to evaluate, based on immunohistochemical and stereologic studies, the expression of androgen receptors in both epithelial and stromal compartments of the prostatic ventral lobe of the gerbil *Meriones unguiculatus* (Muridae, Gerbillinae) during different ages of postnatal development (young, adult, and old) after the androgen blockage therapies.

Methods

Animals

Seventy-five male gerbils (*Meriones unguiculatus*, Gerbilinae: Muridae), supplied from the Universidade de São Paulo State UNESP (Botucatu, SP, Brazil) were housed under adequate conditions of luminosity (12-12 cicle) and temperature (24°C) and were fed rodent ration (Labina®, Purina, Agribrands do Brasil Ltda.) and water *ad libitum*.

Experimental protocol

The animals were distributed, randomly, into 3 groups of 25 animals each, where each group corresponded to one of the following postnatal development phases: young (with mean age of 48 ± 15.9 days), adults (112 ± 27.7 days), and old (18 ± 5.4 months). In each postnatal development phase 5 groups were formed of 5 animals each, according to the following experimental protocol of androgen blockade: **Group I - Control:** Received the pharmacological vehicle (0.9% saline solution) for 7 days; **Group II - Surgical Castration:** The animals were submitted to bilateral orchectomy by abdominal surgical incision after anesthesia with intra-muscular injections of ketamine-chlorohydrate (50 mg/kg body weight). After surgery the animals of this group were placed in individual boxes and submitted to the experiments for 7 days; **Group III - Chemical Castration:** Daily subcutaneous injections of 0.3 ml of antiandrogenic flutamide 10 mg/Kg/body weight (Sigma Chemical Co, St. Louis, Missouri-EUA) for 7 days [17]; **Group IV - Blockade Androgen Completed (BAC):** Surgical Castration associated with Chemical Castration for 7 days; **Group V - BAC** for 7 days, following received intradermic testosterone injections (1 mg/kg testosterone cypionate [Novaquimica/Sigma Pharma, Hortolândia, São Paulo, Brazil] in 0.25 ml corn oil) for 7 additional days [27].

Morphological and Stereological Analysis

After the respective treatment periods, the five groups of animals at each age were anesthetized lightly by CO₂ inhalation and weighed. After this procedure, the animals

were sacrificed by decapitated and blood collected for serological analysis. For analysis under light microscopy, the entire prostatic complex was excised, weighed and cut into fragments. The samples were fixed for 24h in formaldehyde 4% just prepared in phosphate buffer, pH 7.2. After fixation, the material was dehydrated in graded ethanol series and the embedding was performed in Paraplast (Histosec-Merck). The 4 μ m histological sections were submitted to hematoxylin-eosin (HE) staining for general morphological and stereological analysis of the tissue compartments. The microscopy analyses were performed with an Olympus photomicroscope. The Image-Pro Plus computer software, version 4.5 (©Media Cybernetics) for Windows®, was used to digitize the histological images the each phase of the postnatal development.

The stereological analyses of the tissue compartments was determined according to the procedure of Weibel using a 168-point grid test system [28], as applied to the rat male prostate [29]. The data were obtained from 20 random microscopic fields per experimental group in the ages studied at 100x objective. The relative volume (%) was calculated after counting the number of points that coincided with each of the tissue compartments (epithelium, lumen of ducts, stroma and non-muscle stroma). The absolute volume (mm³) of each of these compartments was determined by multiplying the volume relative by the mean prostatic weight based on the determination that 1 mg of fresh rat ventral tissue had a volume of approximately 1 mm³ [30].

Animal handling and experiments were done according to the ethical guidelines of the State University of Campinas, following the Guide for Care and Use of Laboratory Animals (process Nr. 1214-1). The large sample size used in this work was justified by the minute size of the organ and the large number of analytical procedures employed.

Immunohistochemistry (IHC)

Tissue sections (4 μ m) of formalin-fixed, paraffin-embedded gerbil prostatic ventral lobe were immunostained as described previously. Antigen retrieval was performed in citrate buffer (pH 6.0) by steamer for 45 min. and endogenous peroxidase activity was quenched by incubation in 3% hydrogen peroxide in methanol for 10 min. Thereafter, the

sections were incubated overnight at 4°C in a humidified chamber in rabbit polyclonal AR (N-20) primary antibody (Santa Cruz Biotechnology, CA, USA), diluted 1:100 in PBS containing 3% goat serum. The reaction sites were visualized by incubating the tissue sections with biotinylated second antibody, avindin-biotin-peroxidase complex, and diaminobenzidine reagent. In the negative control sections, PBS containing 3% goat serum replaced the primary antibody and sections were stained with Hematoxylin as a blue nuclear counterstain for 8 sec.

Immunostaining for AR was assessed by visual estimation under light microscopy at x 1000 magnification (oil immersion). Immunostaining for AR was additionally assessed by visual counting of immunopositive and immunonegative nuclei and expressed as a percentage of the total of 30 acini and 30 area stromal nuclei counted [31-33].

Hormonal serum concentrations

After blood collection the serum was separated by centrifugation and stored at -20°C for subsequent hormone assay. The determination of serum concentration of testosterone was performed by luminescence-immunoassay (mouse antibodies anti-testosterone – Johnson & Johnson ®, USA) in an automatic analyzer. The sensitivity was 0.1–150 ng/ml for testosterone. The intra-assay and inter-assay variations were 4.6 and 4.3%, respectively.

Statistical Analysis

All statistical analyses were performed with Statistica 6.0 software (StatSoft). The ANOVA and Tukey honest significant difference (HSD) tests were applied and $p \leq 0.05$ was considered statistically significant.

Results

Morphological and stereologic aspects of the prostatic ventral lobe

The analysis of histological sections shows that the ventral prostate lobe of the gerbil is formed by tubular alveolar glands, lined with a prismatic epithelium which is simple and active in terms of production of secretion (depending on the phase of postnatal development), and with the nuclei of these cells chiefly disposed in a polarized way in the basal region (Fig. 1).

The prostatic ducts are surrounded throughout their extent by layers of smooth muscle fibers (muscular stroma – MS) which connect themselves by closely forming a concentric cluster and by means of a loose connective tissue, non-muscular stroma (NMS), detectable between the epithelium and the muscular stroma. The NMS is also present between the ducts, in the form of loose connective fibrovascular tissues containing fibroblasts and low-caliber blood vessels, each with its important and specific role in the viability and secretory function of the tissue (Fig. 1).

According to the age of the animal, it has been observed that the prostatic ducts have three different types of architectural differentiation, conferring a higher or lower diameter and luminal structure. In the glandular lumen, there is a secretion of an array of proteins since it is stained intensely by eosin (Fig. 1).

Young phase

The control group animals reached an average body weight of 38g and their prostatic complex weighed an average of 0.05g. The ventral lobe is composed of small (or developing) secretory ducts constituted by a simple epithelium with folds in their extension and with mainly oval nuclei. The ducts are lined with a thick layer of concentric smooth muscle fibers and with a large NMS between the epithelium and the smooth muscle layer (Fig. 1). The analysis of the absolute volume of the prostatic compartments have shown that the epithelium occupies 0.83 mm^3 of the organ, the lumen 0.45 mm^3 , the MS 1.1 mm^3 , and the NMS 2.6 mm^3 (Table 1).

The biometrical results show that the group treated with testosterone after BAC presented a significant 221% increase in relative prostatic weight in comparison with the control group, and they have also demonstrated that this growth was reflected in the prostatic compartments after the stereologic analysis, presenting highly significant values. The other treatments, despite decreasing the values of the biometrical parameters, did not present a statistically significant reduction.

Adult phase

In this phase the animals reached an average body weight of 70.4g, and their prostatic complex weighed an average of 0.73g. The morphological analysis associated with stereological data show that the prostatic ducts appeared wider, with a luminal volume of 32.0 mm³, resulting in an epithelium of 11.0 mm³ that displays small epithelial cells, without any type of folding, probably due to its distention during the secretory process. Besides, the ducts are surrounded by only one layer of smooth muscle fiber, in which this muscular stroma occupies a volume of 8.5 mm³, compared with a volume of 22.0 mm³ for non-muscular stroma (Fig. 1 and Table 2).

The biometric data for descriptive statistics presented significant changes only the BAC group, showed a significant decrease in relation to the prostatic complex weight, resulting in a decrease of 39% in relative weight. The animals treated with flutamide and BAC displayed significant reduction in absolute volumes of the epithelium, lumen and non-muscular stroma, when compared to the other groups (Table 2).

Old phase

After completing one year of life, the animals were sacrificed. By then, their average body weight had reached 93.0g and their prostatic complex weighed an average of 0.92g. The ventral prostate region of these animals is composed of prostatic ducts with a luminal volume of 46.0 mm³ and it contains an epithelium of 11.0 mm³ with small folds whose cells can be columnar or short. An interesting feature was the presence of ducts lined with a thick smooth muscle layer with a volume of 11.6 mm³ and a large region of

non-muscular stroma measuring 23.5 mm³, which differs, mainly, the young phase (Fig. 1 and Table 3).

Among the treatments, the castrated group presented a statistically significant reduction in prostatic complex weight, associated with a reduction of the absolute volume of the epithelium, lumen and the non-muscular stroma. In addition, the group treated with testosterone after the androgen blockage showed a significant growth of 22% in prostate relative weight (Table 3).

The morphological variations presented in the muscular and non-muscular stroma of the control animals along the three ages analyzed appear to be related to the extensive proliferation of the secretory compartment of the epithelium during sexual maturity (Fig. 1). After the analyses of the relative volumes (%) of the prostatic compartments, it was possible to notice that the proportion of MS and NMS had apparently decreased from the young age to adulthood and it had also shown a luminal increase (Tables 1, 2 and 3).

Hormonal profile

The hormonal dosage of serum testosterone presented a particular characteristic for each phase of postnatal development. The young control group presented 3.2 ng/ml of serum testosterone, 3.8 ng/ml in the adult group, and 2.7 ng/ml in the old group. Among the different treatments to which the animals were subjected, the groups treated only with antiandrogenic flutamide and those treated with testosterone after BAC were the ones that showed statistically significant results. The treatment with flutamide for 7 days was sufficient to increase the testosterone concentrations in the 3 ages researched. Hence, the treatment with testosterone after BAC for 7 days caused the levels to exceed 300 ng/ml at all ages (Tables 1, 2, and 3).

Immunohistochemical analysis

The immunohistochemical analyses allowed the standardization of different staining intensity degrees related to AR in the ventral lobe of the gerbil prostate as degrees of intensity ranging from Strong and Moderate to Negative (no stain).

The immunohistochemical evaluation associated with stereology demonstrates that in the control group there is a heterogeneous pattern of positive responses for AR in the stromal region and mainly in the epithelial region of each phase of gerbil postnatal development. The AR expression was detected in the nuclei of basal and luminal epithelial cells and of some stromal cells (nuclei of smooth muscle cells and fibroblasts) and the epithelial region has presented a greater intensity of expression to AR than the stromal region (Fig. 2).

The epithelial basal cells, discontinuously located along the basal membrane and in lesser frequency than the secretory luminal cells, showed apparently the same degree of stain intensity for each phase and, therefore, they can also be direct targets of androgenic action (Fig.2). In the negative control of immunohistochemical reaction, no AR expression was detected in the nuclei of either stromal or epithelial cells during the 3 ages evaluated (Fig. 2A).

In addition, it could be noted that all the types of androgen blockage therapies used in the 3 phases of postnatal development significantly reduced the intensity degree of AR expression compared to the control group. In the each group experimental was observed an increase in the AR-positive stromal cells when compared to the epithelium after the androgen blockage therapies (Fig. 3 and 4).

Young phase

In the control group 49% of the epithelial cells (luminal + basal) presented strong staining intensity, 32% stained moderately and 22% were AR negative. The same analysis of the stromal region demonstrated a balance between strong staining intensity corresponding to 33%, whereas moderate and negative staining corresponded to 36% and 31%, respectively (Fig. 2, 3 and 4). Thus the morphostereological characteristics

associated with the hormonal profile during the young age apparently contributed to stimulating a major expression of AR in the stromal region (69% AR-positive cells and 31% AR-negative cells) and even more so in the epithelial region (78% AR-positive cells and 22% AR-negative cells), in comparison with the other phases of postnatal development (Fig. 2, 3 and 4).

The stereological results of AR expression in the prostate ventral lobe show a significant decrease, mainly the high degree of staining, in all the experimental groups compared to the control group. The nuclei of both the epithelial and stromal cells responded positively to AR expression, but with very heterogeneous degrees of staining intensity (Fig. 2, 3 e 4). Moreover, after the androgen blockage therapy was possible to observe an increase in the stromal region AR-positive cells associated an increase the AR-negative epithelial cells.

When one compares the possible efficiency of androgen blockage during this developmental phase, it is possible to observe that orchietomy and antiandrogenic action, as well as both associated can considerably decrease the AR expression in the prostatic ventral region (Fig. 3 and 4). Among the 3 ages researched, the young phase presented the highest AR stain intensity, both in the epithelium and in the stroma, even after the androgen blockages.

Adult phase

Although the intensity of AR expression in comparison with the young phase was lower, it was greater than that of the old phase. A highly heterogeneous pattern of AR-positive cells can be observed both in the epithelial and stromal compartments (Fig. 2).

Within the control group 42.5% of cells in the epithelial region presented strong staining intensity, 27% moderate staining, and 30.5% negative; but in the stroma, 23.0% of cells presented strong stain, 29% moderate, and 48% AR-negative. These data demonstrate that, although the adult phase presents the highest concentration of serum testosterone, there was a sharp decrease of AR expression mainly in the stromal region in comparison with the young phase (Figs. 3 and 4).

In addition, it was possible to observe that within all the experimental groups, mainly in groups II and IV which had been surgically castrated, there was a marked and significant decrease of AR expression in the prostatic compartments. In the animals that had been surgically castrated 7 days before, 92% of the epithelial cells and 84% of the stromal cells had been AR-negative (Fig. 3 and 4).

Hormonal reposition with testosterone after BAC resulted in a high concentrations of this androgen in the serum, but in contrast to the young phase, it did not cause a significant increase in the absolute and relative volumes of the different prostatic compartments analyzed and, in addition, a significant increase could be noted in the AR-negative cells (80% in the epithelium and 71.5% in the stroma) (Fig. 3 and 4).

Old Phase

The immunohistochemical analyses associated with AR stereology in the control group revealed that 40% of the epithelial cells were AR-negative, while 35% and 25% presented moderate and strong staining, respectively. The highly significant results were revealed in the stroma with strong staining intensity corresponding to only 17% of the cells, 19% moderate staining, and 64% of AR-negative cells. Among the 3 phases of postnatal development studied, the old phase presented the lowest degree of expression of androgen receptors in the ventral lobe of the gerbil prostate (Figs. 2, 3 and 4).

As for the 4 experimental groups, it was possible to notice highly significant differences of reduction of AR expression in comparison with the control group. Within the castrated group, 91.5% of the epithelial cells and 89.0% of the stromal cells were AR-negative while in the BAC group, 91% of epithelial cells and 87% of the stromal cells were AR-negative (Figs. 3 and 4).

Since the prostate of senile animals is an androgen-dependent gland, it was possible to observe a statistically significant increase of AR expression in the group V in comparison with the other androgen-blocked groups, that is, in the epithelial compartment, 32% of the cells were AR-positive and 68% were AR-negative, and in the stroma, the breakdown between AR-positive and AR-negative was 46% to 54%, respectively. Therefore, the gerbil old age provides the best evidence that the prostate is an androgen-

dependent organ and that the 7-day androgen treatment after androgen blockage resulted in the immunostained need normal conditions in comparison with both the adult and the young phases of this species (Fig. 2, 3 and 4).

Thus, the immunohistochemical analyses associated with the stereology of androgen receptor expression in the ventral lobe of the gerbil prostate have contributed to elucidating the fact that in this rodent species there is a heterogeneous pattern of positive responses to AR in each prostatic compartment and to each phase of postnatal development in the treated groups, compared to the control groups. Besides, the analysis of variance (ANOVA) shows that the different staining degrees related to androgen receptors at each age were highly significant, with $p \leq 0.05$.

Discussion

The immunohistochemical analyses of the ventral lobe of the gerbil prostate have shown that androgen receptor expression has been detected in the nuclei of luminal and basal epithelial cells as well as in some stromal cells.

The basal cells of the rat adult prostate and the prostatic ventral region of mice were AR negative and, for that matter, they do not seem to be direct targets of androgenic action [31, 34-37]. This absence of AR in the basal cells can be a common characteristic in other species of rodents [46].

In the gerbil, the basal cells express the androgen receptor protein in the 3 phases of postnatal development, but the greatest concentrations are apparently in the young animals. The AR staining intensity corresponded to that of luminal cells in all the experimental groups. Besides, this cluster of cells showed strong staining after hormonal reposition of testosterone in animals that had been subjected to BAC, probably by responding to the androgenic action with an increase in proliferation.

These data can be best explained based on studies of castrated rats, in which it is possible to observe that, after castration, the luminal cells undergo drastic atrophy while the basal cells remain unchanged and they also increase their proliferation rate in order to

reconstruct the glandular epithelium when there is androgen reposition [44, 45, 47]. The functional meaning of basal cells in rat prostates is not quite established, although it has been suggested that they can act as either reserves or stem cells of the luminal epithelium as well as phagocytic cells that regulate the number of secretory cells or as potential mediators between the activities of luminal cells related to regulation of the transportation of materials between the stromal and epithelial compartments [31, 48].

More recent immunohistochemical analyses have shown that these cells express the cytokeratins CK5 and CK14, as seen in undifferentiated epithelial-cell staining (in proliferation activity) [49]. Based on these findings, this cellular type has been assigned a role in the regeneration of the glandular epithelium and it has also been suggested that at least a part of this cell population has stem cell properties [45, 47]. This could explain the AR expression in the basal cells of the gerbil ventral prostate, even after the androgen blockages.

Other important components of the glandular epithelium include the luminal secretory epithelial cells since they are the most abundant and are responsible for the secretion of prostatic fluid. These cells, often cylindrical and differentiated, not only express activity specific to enzymes such as prostatic acid phosphatase [51] and prostate specific antigen-PSA, but also can be identified by their selective expression of cytokeratins CK8 and CK18 [49, 52].

In the gerbil, these cells have presented a high intensity of AR expression located in the nucleus, which indicates that they are direct targets of androgenic action. The androgen blockage caused a decrease in AR expression, which thus provoked the emergence of a heterogeneous pattern of AR positive and AR negative cells at all the ages evaluated. The intense AR staining observed in luminal epithelial cells contrasts with the abundance of 5 α -reductase enzyme present in the stroma, which converts testosterone into dihydrotestosterone (DHT) in the prostate and can also directly act on the epithelial region [38-41, 53, 54].

The populations of luminal and basal cells have been apparently resistant to the androgenic blockages used for a period of 7 days, that is, these cells activated during the gland recovery process after the exogenic androgen reposition and presented a strong

staining intensity for AR, which has also been observed in adult mice that had been castrated 3 weeks earlier and subjected to hormonal reposition [31]. It is important to observe that hormonal reposition with testosterone resulted an increase in AR expression in the ventral lobe of the gerbil prostate, though; this expression was not the same as the one observed in the control groups. Therefore, the androgenic reposition after some period of androgen blockage not showed the same comportament in AR expression for motives that are still unknown.

The according by [55], described the regional morphofunctional differences along the system of ducts of the rat prostate ventral lobe. These authors suggested that the epithelium responds in different ways to the same concentration of circulating androgen, with responses ranging from cellular proliferation to cellular death, and that the activity of epithelial cells is determined by the stromal component.

At first, the epithelium was regarded as the main target of the action of androgens in the prostate. However, in recent decades, several findings indicate that the androgens act primarily on the prostatic stromal cells and the epithelial activity is, for the most part, mediated by stromal factors [38, 43, 55-57].

Although there are some variations among the species, the prostatic stroma in mammals is composed of smooth muscle cells and fibroblasts spread among the elements of the extracellular matrix. The stromal cells produce a complex gamut of components of the extracellular matrix including types I and III fibrillar collagen as well as type VI microfibrillar collagens [58, 59]. A series of studies have attempted to clarify the participation of these macromolecules in controlling the prostatic function based on the phenomenon of prostatic regression caused by castration [58-62].

In the gerbil, it was possible to observe that the stromal cells are AR-positive, mainly the smooth muscle cells around the ducts, but they present a significantly lesser staining intensity in relation to the epithelial layer, which suggests that the epithelial cells present a greater amount of stroma-related androgen receptor protein. However, after hormonal treatments, it was possible to notice an increase in stromal AR expression in relation to the epithelium in the all phase of gerbil postnatal development. Therefore, in this species of rodents, there is a homeostasis regulated by both the direct and the indirect

action of androgen receptors and that both the epithelial and stromal compartments seems to be the mains targets of the androgenic and antiandrogenic action and the emergence of prostatic age-related lesions [62].

Among the 3 ages studied, the young phase has been the one that presented the highest staining intensity for AR, even after the androgen blockers. The according by [63], observed that the presence of AR in the prostate after castration and in the absence of endogenous ligands cannot prevent receptor transcriptional activity by the cells, and they have also stated that several molecular signaling pathways of the growth factors can influence the transcriptional activity of androgen receptors.

The concentrations of serum testosterone produced by the testicles in the young phase apparently contributed to stimulating a major expression of AR in the prostatic compartments, which is necessary for the development and functionality of this organ during this phase of postnatal development until it reaches adulthood [3, 42]. Besides, the effective response to the concentrations of circulating androgens caused the prostate of the surgically and / or chemically castrated animals not to present significant decrease in its relative prostate weight as well as in the expression of androgen receptors in comparison with the other ages of postnatal development.

These data are based on the works of [64] and [7], about the rat prostate, where it was possible to observe that the serum testosterone concentrations are very low during the first postnatal weeks and that, during puberty (young phase), there is a significant increase in serum testosterone, which induces prostatic growth. Moreover, it is important to observe that the total cellular content for AR is influenced by the presence or absence of androgens, a process called autoregulation [10, 11, 65].

Statistically, the androgenic blockage reduces AR expression much more in the epithelial region than in the stromal region, and all of this is associated with a decrease in absolute and relative volumes of the epithelial compartment and with an increase in the stromal compartment during the young phase. Exogenous hormonal reposition after BAC resulted in a high concentration of serum testosterone, which stimulated a significant increase in the absolute volume of different prostatic compartments, resulting in an increase in the relative weight of the prostate. But this hormonal reposition was not

sufficient to stimulate a recovery or an AR major expression after the androgenic blockage, that is, it was possible to notice a significant increase of AR-negative cells (78% in the epithelium and 74% in the stroma). Therefore, it was not possible to observe a close relationship between the size of the prostatic complex and the intensity of AR expression during the young phase.

In the adult phase, once the prostatic gland had reached its normal size, an increase was observed in the luminal diameter associated with an apparent decrease in epithelial height and a reduction in the stromal area in comparison with the young phase. In addition, the concentrations of circulating androgens in this period are responsible for the morphofunctional maintenance of the organ, which indicates a heterogeneous pattern of AR expression in the epithelial and stromal cells probably due to the balance between the proliferation and the death of cells in the prostatic compartments. Statistical analyses after the androgenic blockage therapies showed a significant reduction in the AR expression degree.

Moreover, in the adult phase, AR expression does not seem to be directly related to the weight of the prostatic complex and even less to the detected concentration of circulating androgen. Therefore, there must be an intrinsic androgen-dependent autoregulatory mechanism in AR expression in the gerbil prostate, as shown in experiments carried out by [3], with Brown Norway rats subjected to orchectomy. A series of partial studies developed by [42, 46, 66, 67], with young and adult Sprague Dawley rat prostates reported that there is a specific autoregulation in each prostatic lobe for the androgen receptor.

When the animals reached old age, a highly significant decrease in the serum testosterone concentrations associated with the high rate of AR-negative cells was observed not only in the control group but also in the experimental groups. Therefore, this phase of development, in comparison with the others, seems to be much more susceptible to variations and hormonal treatments. One of the hypotheses for explaining this phenomenon is the fact that there are changes, whether molecular or not, in the nuclear AR expressions due to the decrease of testicular androgen production with age, causing the epithelial cells

to contribute to the evolution of cellular androgen-dependent hyperplasia and, in more serious cases, they can become androgen-independent [3, 7, 68, 69].

After the moment when the prostate has reached its highest level of normal growth, cellular hyperplasia or prostatic carcinoma appears in some species, including humans, dogs and some species of rats. These changes are related to the decrease of testicular androgen production and to peripheral concentrations of androgens acting on the prostate. Besides, AR mutations can occur with a certain frequency, especially in advanced prostatic tumors, which develop resistance to hormonal therapy. But the molecular mechanisms involved in the processes of proliferation, differentiation and apoptosis have not yet been completely established, not to mention the mechanisms of neoplastic transformation and carcinogenesis [40, 41, 70-72].

The androgens, whether from testicular or adrenal origins, are not necessary for the development of epithelial receptors in the postnatal rat prostate and that the AR expression in the epithelial region is genetically determined by androgens prenatally or by other factors that remain unknown [73]. This could explain the heterogeneous pattern of AR expression in the prostatic compartments in each phase of gerbil postnatal development in the control groups and especially in the experimental groups.

The influence of the androgens on both the structural organization and physiology of the prostate is a well-known fact. It is known that androgen suppression leads to a process of involution known as prostatic regression, mainly characterized by its epithelial atrophy and progressive decrease in glandular volume [16]. Thus the androgen blockage therapies dispensed during the 3 phases of gerbil postnatal development significantly influenced the decrease of AR expression in the prostatic compartments.

According to [74], described the effect of castration on the adult gerbil prostatic lobes. The results demonstrate that the decrease of androgens caused a progressive reduction of the wet weight and the relative weight of the set of prostatic lobes. Three weeks after castration, the lobes had lost more than a half of their weight. This process of involution resembles the one described for the rat [57, 61, 75] and it is characterized by reduction of acinar size and by epithelial atrophy, which in the species studied had become clearer two weeks after castration. In addition, the comparison of the

involution process between the lobes of the Mongolian gerbil shows that the ventral lobe decreased in epithelial height and volume during the first week after castration, which was not detected in the dorsal lobe. Such an observation may suggest a greater androgenic dependence on the epithelial cells of the ventral lobe in comparison with the dorsal lobe for the maintenance of secretory activity, as observed in other rodents [3].

According to [75], androgen depletion through chemical or surgical castration causes an important prostatic involution and, although about 5% of residual circulating testosterone remains after orchectomy, peripherally produced by the adrenal gland, this testosterone is not enough to maintain the organ homeostasis, which could explain the significant changes in the prostatic compartments.

Flutamide is a potent non-steroidal antiandrogenic agent functionally specific for the sexual androgen-dependent accessory structures, which is the case in the prostate gland. Although this drug is efficient in treating BPH by inhibiting the union of androgens to their respective cellular receptors, it also stimulates the release of LH and, consequently, the increase of testosterone concentrations and, for that reason, many patients remain potent during the treatment [19, 21, 22, 76]. These physiological results have also been observed in the gerbil, in which the testosterone concentrations significantly increased in association with the decrease of AR expression in comparison with the control group.

In the lateral prostate of male guinea pigs, androgen blockage with flutamide has provoked distinct morphological and structural changes in some tissue components taking into account the phase of postnatal development. This differentiated action of flutamide in several phases of the postnatal development seems to be directly connected to hormonal influence that is sometimes absolute and sometimes relative or non-existent in rodents [17, 64]. It could explain the heterogeneous pattern of AR expression in the gerbil prostate ventral lobe.

Several studies have demonstrated that the combination of flutamide with surgical castration in the treatment of prostate cancer increased the degree of responses to the treatment and, most importantly, it increased survival by an average of 7 months compared to the utilization of only LHRH agonist and orchectomy. Besides, these studies

state that, for best efficiency, the anti-androgen must be combined with surgical castration during the initial phases of the treatment [19, 23, 64, 76].

The combination of flutamide with surgical castration for 7 days resulted in the decrease of the serum testosterone concentrations associated with a significant decrease of AR expression, mainly in the old phase where the concentration of endogenous androgen is lower in comparison with the other ages. Therefore, this therapeutic combination dispensed to this rodent species seems to be the most effective means of blocking circulating androgen, which in turn will be able to be used for treatment of age-related prostatic lesions.

The vast scope of research in the area of steroid receptors in the last 3 decades has provided considerable information on the physiological and pathologic functions of steroid receptors. The androgen receptor is the most recently cloned steroid receptor and it has been extensively researched due to its involvement in the male reproductive system and in prostate cancer [16, 42, 77-79]. The analyses of each modulation stage of AR activity constitute a prerequisite for better understanding the of androgen receptor expression in the target tissues. Molecular and clinical studies have provided important information on how the androgen-receptor signaling pathways can act on these diseases. However, the evaluation of other molecular mechanisms is necessary to understand the physiological and pathological functions of these androgen-receptor signaling pathways, as well as the development of therapeutic drugs to treat the patients.

Conclusions

In the Mongolian gerbil there is a distribution pattern of AR in the prostatic ventral lobe through postnatal development in which the younger animal the greater the interaction of circulating androgens that stimulate the AR expression in the epithelial and stromal compartments. Besides, in this species of rodent, the basal cells express AR and it is possible to assign this cell population property of stem cells that play an important role in the regeneration of the glandular epithelium.

The androgen blockage therapies reduced AR expression in the prostatic compartments but the androgenic reposition after these blockages was not showed the same the degree of expression of androgen receptor near normal physiological conditions. In general, the complete androgen blockage therapy seems to be the most efficient means of androgen depletion in the gerbil, which, in turn, can be used to treat prostatic age-related lesions.

It is possible to conclude that the regulation and distribution of androgen receptors in the gerbil prostatic compartments are complex mechanisms that probably are genetically regulated by prenatal androgens or other factors that are still unknown. The gerbil appears to be a valuable experimental model in the attempt to improve the knowledge of the morphophysiological and pathological behavior of the human prostate throughout aging and the formulation of new therapeutic ideas to prevention prostate cancer.

Authors' contributions

RSC carried out the sampling, tissue processing, developed the methods and wrote the manuscript. SGPC helped immunohistochemical assays, supervised analysis and participated in discussing the results. WRS carried out the hormonal immunoassay and participated in discussing the results, and helped write the manuscript. FCAS made the statistical analysis, participated in the discussion of the results, and helped write the manuscript. SRT and PSLV participated in the planning of the experiments and, the discussion of the results, and helped write the manuscript.

Acknowledgments

This paper is part of the thesis presented by RSC to the Institute of Biology, UNICAMP, in partial fulfillment of the requirement for a PhD degree, and was supported by grants from the Brazilian agencies CAPES – Coordinating Body for Training University-level Personnel (fellowship to RSC), FAPESP – São Paulo Research Foundation (Proc. Nr. 02/12942-6) and CNPq - Brazilian National Research and Development Council (Proc. Nr. 301111/05-7 research fellowship to SRT). The authors wish to thank to Mr. Luiz Roberto Falleiros Júnior for their technical assistance, as well as all other researchers at the Microscopy and Microanalysis Laboratory. Acknowledgement is also due to Mr. James Welsh and Mr. Ricardo S. Sobreira for English-language revision of this paper. Special tanks to Dr. Ana Maria G. Custódio for suggesting and advice.

References

1. Cunha GR, Hayward SW, Dahiya R, Foster BA: Smooth muscle-epithelial interactions in normal and neoplastic prostatic development. *Acta anat* 1996, 155: 63-72.
2. Thomson AA, Foster BA, Cunha GR: Analyses of growth factor and receptor mRNA levels during development of the rat seminal vesicle and prostate. *Development* 1997, 124: 2431-2439.
3. Banerjee PP, Banerjee S, Brown TR: Increased Androgen Receptor Expression Correlates with Development of Age-Dependent, Lobe-Specific Spontaneous Hyperplasia of the Brown Norway Rat Prostate. *Endocrinology* 2001, 142(9): 4066-4075.
4. Taplin ME, Ho SM: The endocrinology of prostate cancer. *J Clin Endocr Metabo*. 2001, 86(8): 3467-3477.
5. Dehm SM, Tindall DJ: Molecular regulation of androgen action in prostate cancer. *Journal of Cellular Biochemistry* 2006, 99: 333-344.
6. Brum IS, Spritzer PM, Brentani MM: Biologia Molecular das Neoplasias de Próstata. *Arq Bras Endocrinol Metab* 2005 49(5): 797-804.
7. Marker PC, Donjacour AA, Dahiya R, Cunha GR: Hormonal, cellular, and molecular control of prostatic development. *Developmental Biology* 2003, 253: 165-174
8. Corradi LS, Góes RM, Carvalho HF, Taboga SR: Inhibition of 5- α -reductase activity induces stromal remodeling and smooth muscle de-differentiation in adult gerbil ventral prostate. *Differentiation* 2004, 72(5): 198-208.
9. Hughes IA, Lim HN, Martin H, Mongan NP, Dovey L, Ahmed SF, Hawkins JR: Developmental aspects of androgen action. *Molecular and Cellular Endocrinology* 2001, 185: 33-41.
10. Heinlein CA, Chang C: Androgen receptor in prostate cancer. *Endocr Rev* 2004, 25(2): 276-308.
11. Farla P, Hersmus R, Trapman J, Houtsmuller AB: Antiandrogens prevent stable DNA-binding of the androgen receptor. *Journal of Cell Science* 2005, 118: 4187-4198.
12. Gao W, Kim J, Dalton JT: Pharmacokinetics and pharmacodynamics of nonsteroidal androgen receptor ligands. *Pharmaceutical Research* 2006, 23(8):1641-1658

13. Buchanan G, Irvine RA, Coetzee GA, Tilley WD: Contribution of the androgen receptor to prostate cancer predisposition and progression. *Cancer Metastasis Rev* 2001, 20: 207–223.
14. Feldman BJ, Feldman D: The development of androgen-independent prostate cancer. *Nature Rev Cancer* 2001, 1: 34–45.
15. Leav I, Schelling KH, Adams JY, Merck FB, Alroy J: Role of canine basal cells in postnatal prostatic development, induction of hyperplasia, and sex-hormone-stimulated growth, and ductal origin of carcinoma. *Prostate* 2001, 48: 210-224.
16. Lee DK, Chang C: Expression and Degradation of Androgen Receptor: Mechanism and Clinical Implication. *The Journal of Clinical Endocrinology & Metabolism* 2003, 88(9): 4043–4054.
17. Cordeiro RS, Scarano WR, Góes RM, Taboga SR: Tissue Alterations in the Guinea Pig Lateral Prostate Following Antiandrogen Flutamide Therapy. *BioCell* 2004, 28(1): 21-30.
18. Ruijter E, Van De Kaa C, Miller G, Ruiter D, Debruyne F, Schalken J: Molecular genetics and epidemiology of prostate carcinoma. *Endocr Rev* 1999, 20(1): 22-45.
19. Singh SM, Gauthier S, Labrie F: Androgen Receptor Antagonists (Antiandrogens): Structure-Activity Relationships. *Current Medicinal Chemistry* 2000, 7:211-247.
20. Miyata K, Yabushita S, Sano M, Miyashita K, Okuno Y, Matsuo M: Effects of perinatal exposure to flutamide on sex hormona responsiveness in F1 male rats. *The Journal of Toxicological Sciences* 2003, 28(3): 149-163.
21. Goto K, Koizumi K, Takauri H, Fojii Y, Furuyama Y, Saika H, Suzuki H, Saito K, Suzuki K: Effects of flutamide on sex maturation behavior of offspring born to male rats treated during late pregnancy. *The Journal of Toxicological Sciences* 2004, 29(5): 517-534.
22. Foster PMD, Harris MW: Changes in androgen-mediated reproductive development in male rat offspring following exposure to a single oral dose of flutamide at different gestational ages. *Toxicological Sciences* 2005, 85: 1024–1032.
23. Singh P, Uzgare A, Litvinov I, Denmeade SR, Isaacs JT: Combinatorial androgen receptor targeted therapy for prostate cancer. *Endocrine-Related Cancer* 2006, 13: 653–666.
24. Raghow S, Kuliyev E, Steakley M, Greenberg N, Steiner MS: Efficacious chemoprevention of primary prostate cancer by flutamide in an autochthonous transgenic model. *Cancer Res* 2000, 60: 4093-4097.

25. O`neill AJ, Boran SA, O`keane C, Coffey RNT, Hegarty NJ, Hegarty P, Gaffney EF, Fitzpatrick JM, Watson RWG: Capase 3 expression in benign prostatic hyperplasia and prostate carcinoma. *Prostate* 2001, 47: 183-188.
26. Shulz WA, BurchardT M, Cronauer MV: Molecular biology of prostate cancer. *Mol Hum Reprod* 2003, 9(8): 437-448.
27. Santos FCA, Carvalho HF, Góes RM, Taboga SR: Ultrastructural characterization of secretory cell in the gerbil female prostate. *Tissue Cell* 2003, 34: 273-282.
28. Weibel ER. 1963. Principles and methods for the morphometric study of the lung and other organs. *Lab. Invest.* 12: 131-155.
29. Huttunen E, Romppanen T, Helminen HJ: A histoquantitative study on the effects of castration on the rat ventral prostate lobe. *Jounal of Anatomy* 1981, 132: 357-370.
30. Vilamaior PSL, Taboga SR, Carvalho HF: Postnatal growth of the ventral prostate in Wistar rats: a stereological and morphometrical study. *Anat Rec* 2006, 288:885–892.
31. Mirosevich J, Bentel JM, Zeps N, Redmound SL, D'antuono MF, Dawkins HJS: Androgen receptor expression of proliferating basal and luminal cells in adult murine ventral prostate. *Journal of Endocrinology* 1999, 162: 341-350.
32. Sweat SD, Pacelli A, Bergstrahl EJ, Slezak JM, Bostwick DG: Androgen receptor expression in prostatic intraepithelial neoplasia and cancer. *The Journal of Urology* 1999, 161: 1229-1232.
33. Chan TY, Mikolajczyk SD, Lecksell K, Shue MJ, Rittenhouse HG, Partin AW, Epstein JI: Immunohistochemical staining of prostate cancer with monoclonal antibodies to the precursor of prostate-specific antigen. *Urology* 2003, 62 (1): 177-181
34. Hayward SW, Rosen MA, Cunha GR: Stromal-epithelial interactions in the normal and neoplastic prostate. *Br J Urol* 1997, 79 (Suppl.2): 18-26.
35. Galbraith SM, Duchesne GM: Androgens and prostate cancer: Biology, pathology and hormonal therapy. *European Journal of Cancer* 1997, 33(4): 545-554.
36. Titus MA, Schell MJ, Lih FB Tomer KB, Mohler JL: Testosterone and Dihydrotestosterone Tissue Levels in Recurrent Prostate Cancer. *Clin Cancer Res* 2005, 11(13): 4653-4657.
37. Berthold DR, Moore MJ: Novel targets in prostate cancer. *Expert Opin Ther Target* 2006, 10(5): 777-80.

38. Takeda H, Chodak G, Mutchnik S, Nakamoto T, Chang C: Immunohistochemical localization of androgen receptors with mono and polyclonal antibodies to androgen receptor. *J of Endocrinology* 1990, 126: 17-25.
39. Steers WD: 5alpha-reductase activity in the prostate. *Urology* 2001, 58(6 Suppl 1):17-24.
40. Powell SM, Brooke GN, Whitaker HC, Reebye V, Gamble SC, Chotai D, Dart DA, Belandia B, Bevan CL: Mechanisms of androgen receptor repression in prostate cancer. *Biochem Soc Trans* 2006, 34(Pt 6): 1124-7.
41. Sanchez D, Rosell D, Honorato B, Lopez J, Arocena J, Sanz G: Androgen receptor mutations are associated with Gleason score in localized prostate cancer. *BJU Int* 2006, 98(6):1320-5.
42. Prins GS, Birch L, Greene G: Androgen receptor localization in different cell type of the adult rat prostate. *Endocrinology* 1991, 129(6): 3187-3199.
43. Tilley WD, Horsfall DJ, Mcgee MA, Henderson DW, Marshall VR: Distribution of oestrogen and androgen receptors between the stroma and epithelium of the guinea-pig prostate. *J. Steroid Biochem* 1985, 22(6): 713-9.
44. Bonkhoff H, Remberger K: Differentiation pathways and histogenetic aspects of normal and abnormal prostatic growth: a stem cell model. *Prostate* 1996, 28: 98-106.
45. Signoretti S, Waltengny D, Dilks J, Isaac B, Lin D, Garraway L, Yang A, Montironi R, Mckenon F, Loda M: P63 is a prostate basal cell marker and is required for prostate development. *Am J Pathol* 2000., 157(6): 1769-1775.
46. Prins GS, Birch L: Immunocytochemical analysis of androgen receptor along the ducts of the separate rat prostate lobes after androgen withdrawal and replacement. *Endocrinology* 1993, 132:169-178.
47. Peehl DM: Cellular biology of prostatic growth factors. *The Prostate* 1996, supplement 6: 74-78.
48. Cunha GR, Ricke W, Thomson A, Marker PC, Risbridger G, Haywar SW, Wang YZ, Donjacour AA, Kurita T: Hormonal, cellular, and molecular regulation of normal and neoplastic prostatic development. *J. Ster. Bioch. & Molec. Biol.* 2004, 92:221-236.
49. Hudson DL, Guy AT, Fry P, O'hare MJ, Watt FM, Masters JNR: Epithelial cell differentiation pathways in the human prostate: identification of intermediate phenotypes by heratin expression. *J Histoche. Cytoche* 2001., 49(2): 271-278.

50. Custódio AMG, Góes RM, Taboga SR: Acid phosphatase activity in gerbil (*Meriones unguiculatus*) prostate: comparative study in female and male glands during postnatal development. *Cell Biol Int* 2004, 28(5): 335-344.
51. Van Leenders GJLH, Schalken JA: Epithelial cell differentiation in the human prostate epithelium: implications for the pathogenesis and therapy of prostate cancer. Critical Review in Oncology/Hematology 2003, 46(1): 3-10.
52. Wilson RP, Bruchovsky N, Shnitka TK, Rennie PS, Comeau TL: Stromal 5 α -reductase activity is elevated in benign prostatic hyperplasia. *Acta Endocrinologica* 1980, 94: 284-288.
53. Grossmann ME, Huang H, Tindall DJ: Androgen Receptor Signaling in Androgen-Refractory Prostate Cancer. *Journal of the National Cancer Institute* 2001, 93(22): 1687-1697.
54. Nemeth JA, Lee C: Prostatic ductal system in rats: regional variation in stromal organization. *Prostate* 1996, 28: 124-128.
55. Cunha GR, Bigsby RM, Coopple PS, Sugimura Y: Stromal-epithelial interactions in adult organs. *Cell Diff* 1985, 17: 137-148.
56. Sugimura Y, Cunha GR, Donjacour AA: Morphogenesis of ductal networks in the mouse prostate. *Biol Reprod* 1986, 34: 961-971.
57. Carvalho HF, Taboga SR, Vilamaior PSL: Collagen type VI is a component of the extracellular matrix microfibril network of the prostatic stroma. *Tissue Cell* 1997a, 29: 163-170.
58. Carvalho HF, Line SRP: Basement membrane associated changes in the rat ventral prostate following castration. *Cell Biol Int* 1996, 20: 809-819.
59. Carvalho HF, Vilamaior PSL, Taboga SR: Elastic system of the rat ventral prostate and its modifications following orchietomy. *Prostate* 1997b, 32: 27-34.
60. Vilamaior PSL, Felisbino SR, Taboga SR, Carvalho HF: Collagen fiber reorganization in the rat ventral prostate following androgen deprivation: An possible role for the smooth muscle cells. *Prostate* 2000, 45: 253-258.
61. Zanetoni C, Taboga SR: Age-related modifications in stromal and epithelial compartments of the male prostate of *Meriones unguiculatus*. *Acta Microsc* 2001, 3(c): 203-204.

62. Culing Z, Hobisch A, Cronauer MV: Androgen receptor activation in prostatic tumor cell lines by insulin-like growth factor-I, keratinocyte growth factor, and epidermal growth factor. *Cancer Res* 1994, 54: 5474–5478.
63. Dounjacour AA, Cunha GR: The effect of androgen deprivation on branching morphogenesis in the mouse prostate. *Dev Biol* 1988, 128: 1-14.
64. Paris F, Weinbauer GF, Blüm V, Nieschlag E: The effect of androgen and antiandrogens on the immunohistochemical localization of the androgen receptor in accessory reproductive organs of male rats. *J Steroid Biochem Molec Biol* 1994, 48: 129-137.
65. Prins GS, Cooke PS, Birch L: Androgen receptor expression and 5 α -reductase activity along the proximal-distal axis of the rat prostatic duct. *Endocrinology* 1992, 130:3066–3073.
66. Prins GS, Woodham C, 1995. Autologous regulation of androgen receptor messenger ribonucleic acid in the separate lobes of the rat prostate gland. *Biol. Reprod.*, 53:609–619.
67. Zhao GQ, Holterhus PM, Dammshauser I, Hoffbauer G, Aümuller G: Estrogen-induced morphological and immunohistochemical changes in stroma and epithelium of rat ventral prostate. *Prostate* 1994, 21: 183-199.
68. Horsfall DJ, Mayne K, Ricciardelli C, Rao M, Skinner JM, Henderson DW, Marshall VR, Tilley WD: Fase-related in guinea pig prostatic stroma. *Lab Invest* 1994, 70: 753- 763.
69. Wernet N, Gerdes J, Loy V, Seitz G, Scherr O, Dhom G: Investigations of the estrogen (ER-ICA-test) and progesterone receptor in the prostate and prostatic carcinoma on immunohistochemical basis. *Virchows Archiv Pathol* 1988, 412: 387-391.
70. Nevalainen MT, Valve EM, Anhonen T, Yagi A, 1997. Androgen-dependent expression of prolactin in rat prostate epithelium in vivo and organ culture. *FASEB J.*, 11: 1297-1307.
71. Brinkmann AO, Blok LJ, Ruiter PE, Doesburg P, Steketee K, Berrevoets CA, Trapman J: Mechanisms of androgen receptor activation and function. *J Ster Bioch and Mol Biol* 1999, 69: 301-313.
72. Husmann DA, Mcphaul MJ, Wilson JD: Androgen receptor expression in the developing rat prostate is not altered by, castration, flutamide, or suppression of adrenal axis. *Endocrinology* 1991, 128(4):1902-1905.
73. Rochel SS: Variações regionais e efeitos da castração nos lóbulos dorsal e ventral da próstata do gerbilo da mongólia (*Meriones unguiculatus*). Dissertação de mestrado, 2006, Unesp/Ibilce.

74. Antoniolli E, Della-Colleta HHM, Carvalho HF: Smooth muscle cell behavior in the ventral prostate of castrated rats. *Journal of Andrology* 2004, 25(1): 50-56.
75. Pompeo ACL, Damião R, Aguinaga AS: Tumores Prostáticos - I Consenso Brasileiro: Câncer de Próstata 1998. São Paulo, 1998.
76. Hayashi N, Sugimura Y, Kamamura J, Doncajour AA, Cunha GR: Morphological and functional heterogeneity in the rat prostatic gland. *Biol Reprod* 1991, 45: 308-321.
77. Jesik CJ, Holland JM, Lee C: An anatomic and histologic study of the rat prostate. *Prostate* 1982, 3: 81-97.
78. Chow PH, Chan CW, Cheng LY: Contents of fructose, citric acid, acid phosphatase, proteins and electrolytes in secretions of the accessory sex glands of the male golden hamster. *Int J Androl* 1992, 16: 41-45.
79. Reznik G, Hamlim MH, Ward JM, Stinson SF: Prostatic hyperplasia and neoplasia in aging F344 rats. *Prostate* 1981, 2(3): 261-268.

Figures Legends

Figure 1: Histological sections stained with HE of young, adult and old gerbil lobe ventral prostate. Legends: **ep** - epithelium; **L** - lumen; **SMC** – smooth muscle cell; * - non-muscular stroma. Bars: 50 μm .

Figure 2: Histologic sections submitted to AR IHC. Brown stain means positive demarcation. Legends: **arrow-stron** – stained stronger; **arrow-mod** – stained moderate; **arrow-neg** – stained negative; **arrowheads** – basal cell. Bars: 20 μm . **Figure 2A:** Negative Control for IHC.

Figure 3. Graphic representation of epithelial cells counting with different staining degrees of intensity for the AR expression in the different phases of the postnatal development. *Statistically significant differences between groups ($p \leq 0.05$).

Figure 4. Graphic representation stromal cells counting with different staining degrees of intensity for the AR expression in the different postnatal development phases. *Statistically significant differences between groups ($p \leq 0.05$).

TABLE 1: Quantitative analysis of experimental groups in the young phase (mean \pm SEM).

Quantitative analysis (g)	Control	Castrated	Flutamide	BAC	BAC+Testosterone
Body weight *	38.0 ^a \pm 0.73	42.5 ^a \pm 0.9	43.3 ^a \pm 2.0	38.0 ^a \pm 2.0	58.4 ^b \pm 1.5
Prostatic complex weight *	0.05 ^a \pm 0.005	0.05 ^a \pm 0.003	0.04 ^a \pm 0.003	0.04 ^a \pm 0.003	0.23 ^b \pm 0.02
Relative prostate weight † *	0.001 ^a \pm 0.00012	0.0009 ^a \pm 0.0002	0.0009 ^a \pm 0.00008	0.001 ^a \pm 0.00007	0.004 ^b \pm 0.003
Relative weight variation ‡ (%)	—	- 7.7	- 23.5	- 14	+ 221
Testosterone concentrations ng/ml *	3.2 ^a \pm 0.38	1.2 ^a \pm 0.12	13.0 ^b \pm 2.5	3.6 ^a \pm 0.8	318 ^c \pm 11.6
Stereological Data					
Relative Volume (%)					
Epithelium *	17.0 ^a \pm 0.56	11.0 ^b \pm 0.8	20.0 ^c \pm 0.6	11.0 ^b \pm 0.56	20.0 ^c \pm 0.9
Lumen *	9.0 ^a \pm 1.4	5.0 ^a \pm 0.5	8.0 ^a \pm 0.5	3.5 ^b \pm 0.5	45.0 ^e \pm 1.75
Muscular Stroma *	23.0 ^a \pm 1.4	32.0 ^b \pm 2.3	21.0 ^a \pm 1.0	30.0 ^b \pm 1.9	16.0 ^c \pm 0.8
Non-Muscular Stroma *	51.0 ^a \pm 1.82	52.0 ^a \pm 2.4	51.0 ^a \pm 0.9	55.5 ^a \pm 1.95	19.0 ^b \pm 1.0
Absolute Volume (mm³)					
Epithelium *	0.83 ^a \pm 0.02	0.6 ^a \pm 0.04	0.8 ^a \pm 0.025	0.45 ^b \pm 0.02	4.6 ^c \pm 0.21
Lumen *	0.45 ^a \pm 0.7	0.24 ^a \pm 0.025	0.34 ^a \pm 0.02	0.14 ^a \pm 0.02	10.3 ^b \pm 0.4
Muscular Stroma *	1.1 ^a \pm 0.07	1.6 ^b \pm 0.12	0.8 ^a \pm 0.04	1.2 ^a \pm 0.07	3.6 ^c \pm 0.2
Non-Muscular Stroma *	2.6 ^a \pm 0.09	2.6 ^a \pm 0.12	2.0 ^b \pm 0.04	2.2 ^a \pm 0.08	4.5 ^c \pm 0.24

* Statistically significant differences between control and treatments; superscript letters (a, b, c) represent statistically significant differences between the experimental groups, *p \leq 0.05.

† Relative weight corresponds to the ratio between the weight of the prostate and that of the whole body.

‡ Relative weight variation is shown with respect to the control, which was taken as 100%.

TABLE 2: Quantitative analysis from experimental groups in the adult phase (mean ±SEM).

Quantitative analysis (g)	Control	Castrated	Flutamide	BAC	BAC+Testosterone
Body weight	70.4 ± 1.3	68.0 ± 4.3	65.2 ± 4.0	61.0 ± 2.6	70.0 ± 3.2
Prostatic complex weight *	0.73 ^a ± 0.025	0.6 ^a ± 0.1	0.52 ^a ± 0.1	0.4 ^b ± 0.1	0.7 ^a ± 0.05
Relative prostate weight †	0.01 ± 0.0002	0.008 ± 0.0013	0.008 ± 0.0009	0.006 ± 0.0015	0.009 ± 0.0004
Relative weight variation ‡ (%)	—	- 17	- 16	- 39	- 4
Testosterone concentrations ng/ml *	3.8^a ± 0.15	9.7^a ± 0.66	258.4^b ± 47.0	8.0^a ± 0.96	328^c ± 5.2
Stereological Data					
Relative Volume (%)	Epithelium *	15.0 ^a ± 0.75	15.0 ^a ± 0.9	12.4 ^a ± 0.9	11.0 ^b ± 0.8
	Lumen	43.0 ± 2.4	41.5 ± 2.8	44.5 ± 2.8	46.0 ± 2.5
	Muscular Stroma *	11.5 ^a ± 0.9	12.0 ^a ± 0.8	14.5 ^a ± 1.2	17.5 ^b ± 1.0
	Non-Muscular Stroma	30.5 ^a ± 1.9	31.5 ± 1.8	28.6 ± 2.3	25.5 ± 1.8
Absolute Volume (mm³)	Epithelium *	11.0 ^a ± 0.54	9.0 ^a ± 0.55	6.4 ^b ± 0.45	4.5 ^c ± 0.32
	Lumen *	32.0 ^a ± 1.75	25.0 ^b ± 1.7	23.0 ^b ± 1.5	18.4 ^b ± 1.0
	Muscular Stroma *	8.5 ^a ± 0.7	7.5 ^a ± 0.5	7.5 ^a ± 0.62	7.0 ^a ± 0.4
	Non-Muscular Stroma *	22.0 ^a ± 1.4	19.0 ^a ± 1.1	15.0 ^b ± 1.2	10.2 ^c ± 0.7

* Statistically significant differences between control and treatments; superscript letters (a, b, c) represent statistically significant differences between the experimental groups, *p ≤ 0.05.

† Relative weight corresponds to the ratio between the weight of the prostate and that of the whole body.

‡ Relative weight variation is shown with respect to the control, which was taken as 100%.

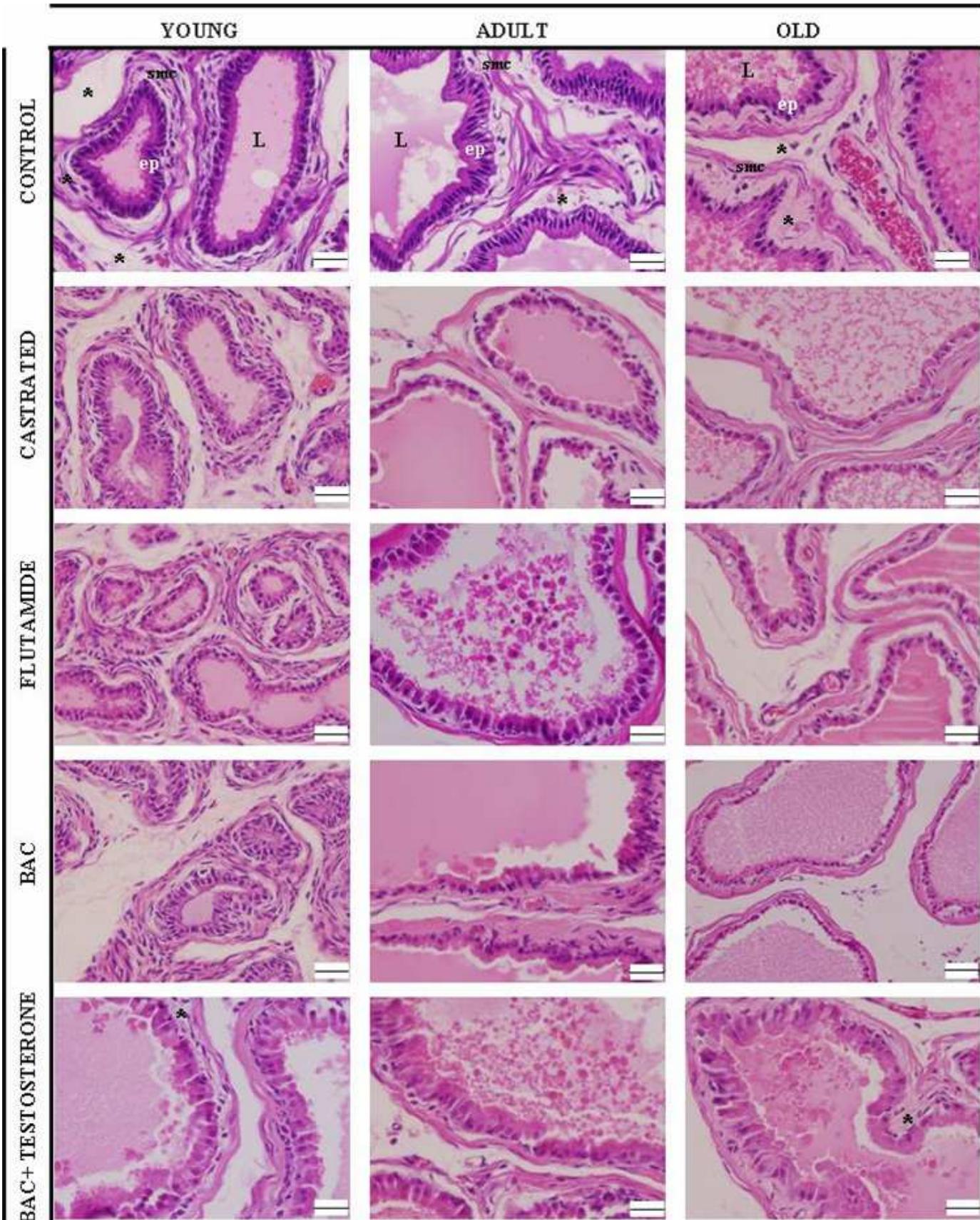
TABLE 3: Quantitative analysis from experimental groups in the old phase (mean \pm SEM).

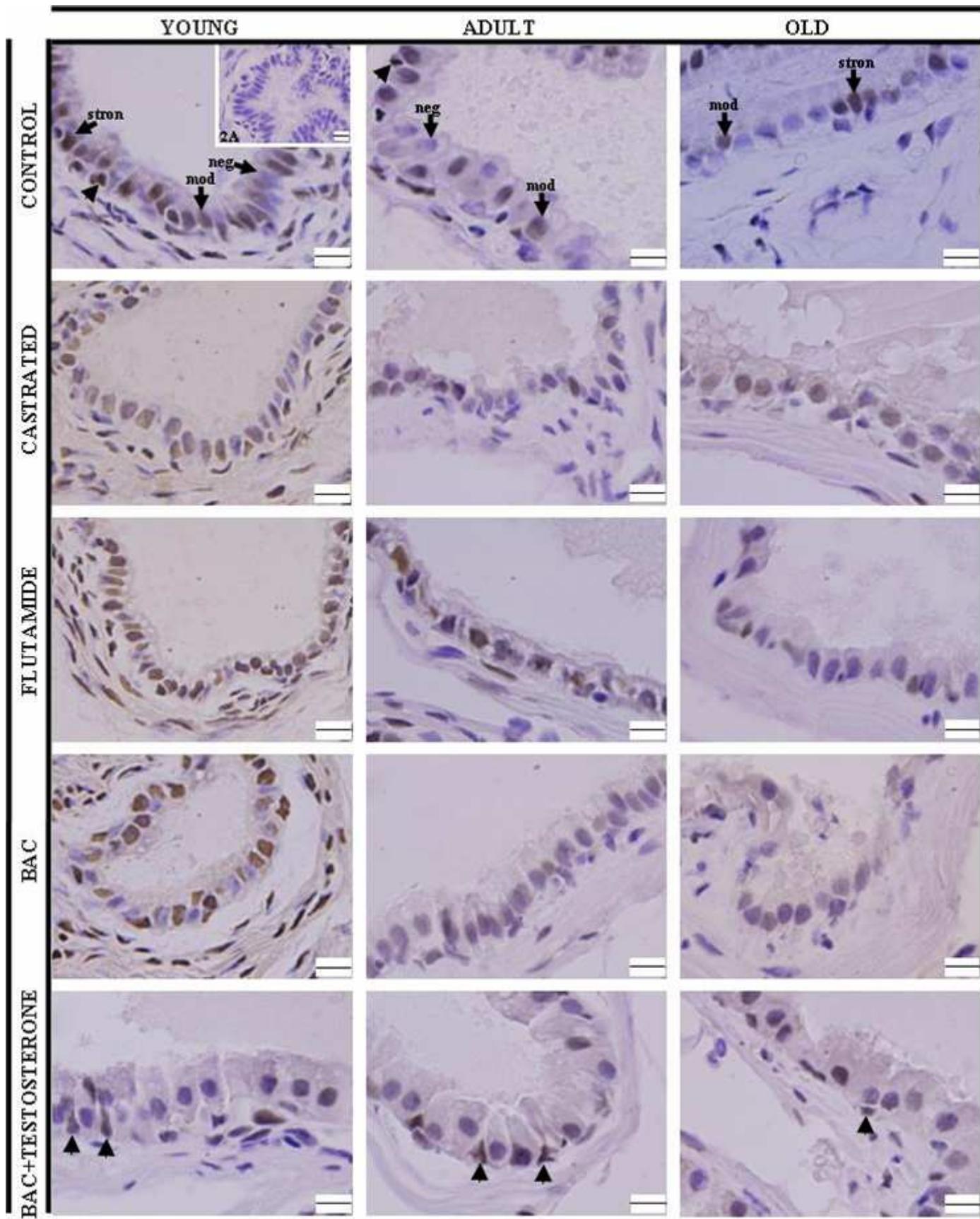
Quantitative analysis (g)	Control	Castrated	Flutamide	BAC	BAC+Testosterone
Body weight	93.0 \pm 2.6	96.0 \pm 12.3	100.5 \pm 3.5	80.0 \pm 2..3	72.2 \pm 4.3
Prostatic complex weight *	0.92 ^a \pm 0.06	0.7 ^b \pm 0.05	0.94 ^{ac} \pm 0.04	0.8 ^a \pm 0.06	0.85 ^a \pm 0.03
Relative prostate weight † *	0.01 ^a \pm 0.0003	0.007 ^a \pm 0.0006	0.009 ^a \pm 0.0005	0.009 ^a \pm 0.0008	0.013 ^b \pm 0.0012
Relative weight variation ‡ (%)	—	- 25	- 2.6	+ 2	+ 22
Testosterone concentrations ng/ml *	2.7 ^a \pm 0.25	1.3 ^a \pm 0.2	55.2 ^b \pm 11	2.5 ^a \pm 0.65	347 ^c \pm 9.0
Stereological Data					
Relative Volume (%)	Epithelium *	12.0 ^a \pm 0.8	11.0 ^a \pm 0.5	13.5 ^a \pm 0.9	10.5 ^a \pm 0.5
	Lumen *	50.0 ^a \pm 1.6	51.5 ^a \pm 2.0	48.0 ^a \pm 1.5	45.0 ^a \pm 2.0
	Muscular Stroma	13.0 \pm 0.7	13.0 \pm 1.5	13.5 \pm 0.85	14.0 \pm 0.9
	Non-Muscular Stroma	25.0 \pm 1.8	24.5 \pm 1.7	25.0 \pm 1.2	30.5 \pm 1.6
Absolute Volume (mm³)	Epithelium *	11.0 ^a \pm 0.74	7.6 ^b \pm 0.33	12.6 ^a \pm 0.86	8.3 ^a \pm 0.44
	Lumen *	46.0 ^a \pm 1.5	36.0 ^b \pm 1.5	45.3 ^a \pm 1.4	36.3 ^b \pm 1.6
	Muscular Stroma	11.6 ^a \pm 0.62	9.2 \pm 1.1	12.6 \pm 0.8	11.0 \pm 0.75
	Non-Muscular Stroma *	23.5 ^a \pm 1.6	17.0 ^b \pm 1.2	23.5 ^a \pm 1.1	24.5 ^a \pm 1.3

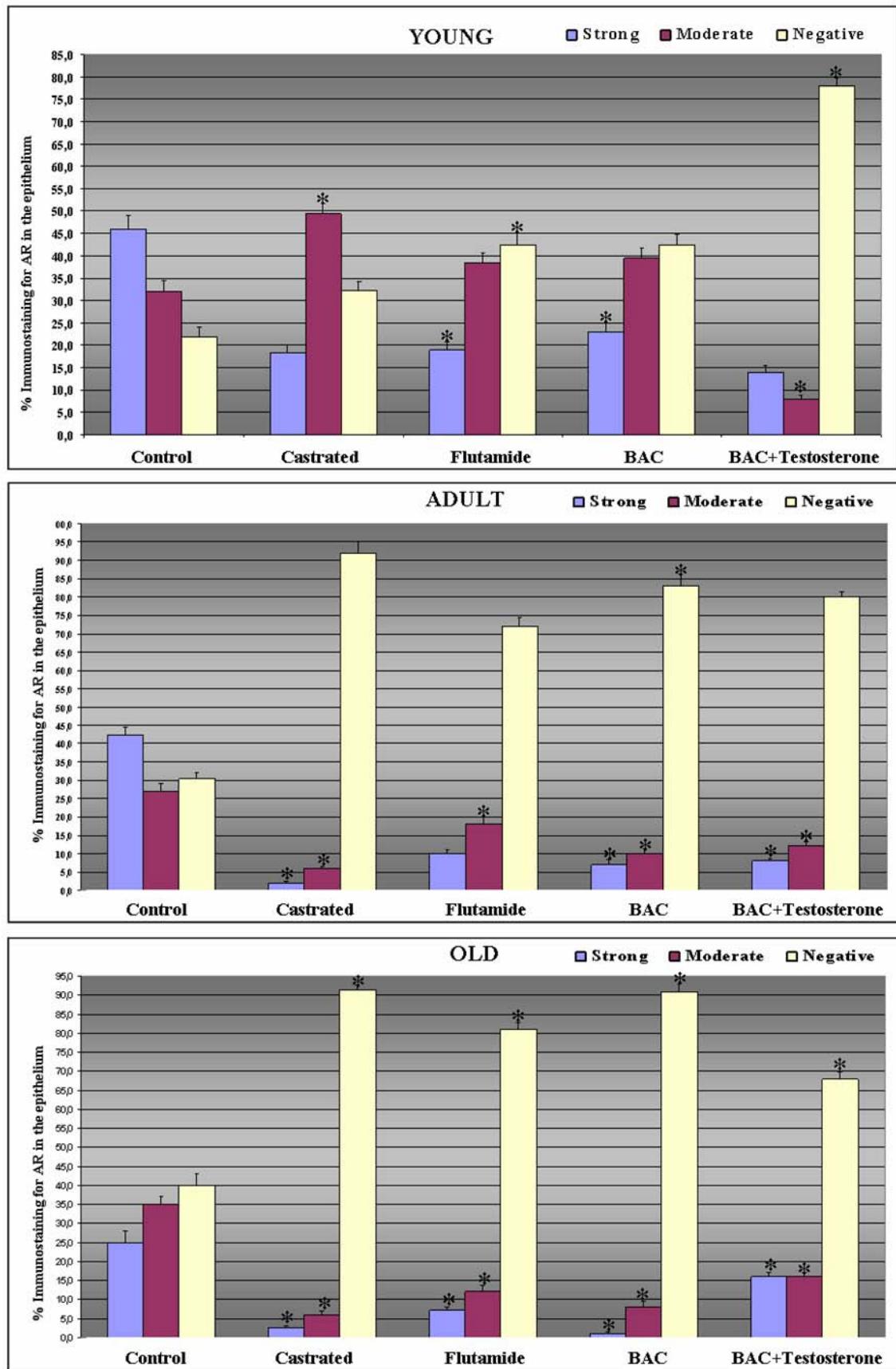
* Statistically significant differences between control and treatments; superscript letters (a, b, c) represent statistically significant differences between the experimental groups, * $p \leq 0.05$.

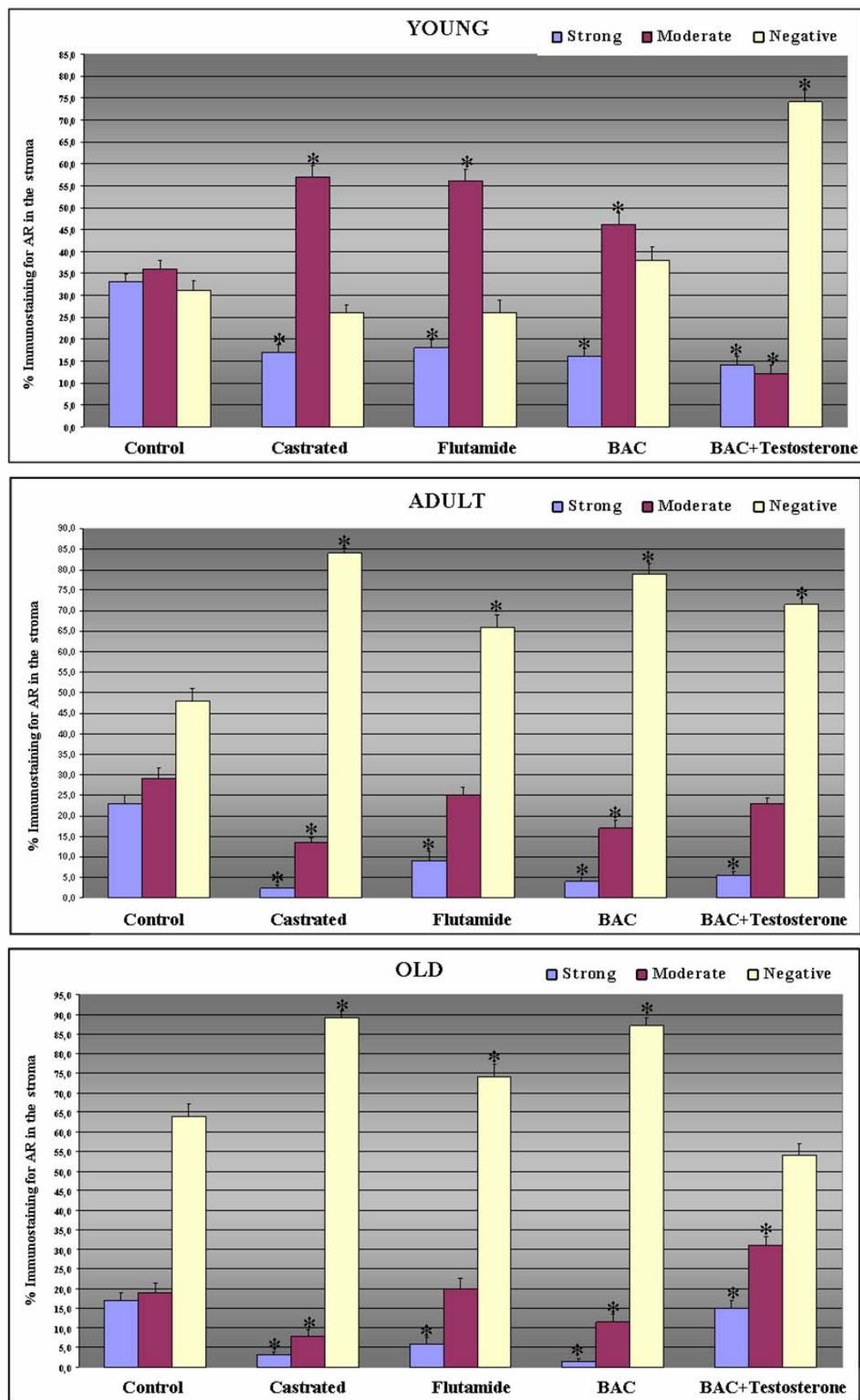
† Relative weight corresponds to the ratio between the weight of the prostate and that of the whole body.

‡ Relative weight variation is shown with respect to the control, which was taken as 100%.









VI. CONCLUSÕES GERAIS

No gerbilo da Mongólia há um padrão de distribuição do RA no lóbulo ventral prostático ao longo do desenvolvimento pós-natal, sendo que quanto mais jovem for o animal maior a interação de andrógenos circulantes estimulando a expressão de RA nos compartimentos epitelial e estromal. Além disso, nessa espécie de roedor as células basais expressaram RA, podendo atribuir a essa população de célula propriedades de células-tronco exercendo um papel importante na regeneração do epitélio glandular.

As terapias de bloqueios androgênicos diminuíram a expressão de RA nos compartimentos prostáticos e a reposição androgênica após esses bloqueios não apresentou o mesmo grau de intensidade de expressão de RA próximo às condições fisiológicas normais. De maneira geral, a terapia de bloqueio androgênico completo parece ser a maneira eficaz de depleção androgênica no gerbilo, e que por sua vez, poderá ser utilizado no tratamento de lesões prostáticas relacionadas à idade.

Conclui-se que a regulação e a distribuição do RA nos compartimentos prostáticos do gerbilo são mecanismos complexos que, provavelmente, são geneticamente regulados por andrógenos antes do nascimento ou por outros fatores ainda desconhecidos. O gerbilo parece ser um modelo experimental valioso na tentativa de melhorar o conhecimento do comportamento morfofisiológico e patológico da próstata humana ao longo do envelhecimento e da formulação de novas idéias de terapias de combate ao câncer de próstata

VII. REFERÊNCIAS BIBLIOGRÁFICAS

- Abate-Shen C, Shen MM. 2000. Molecular genetics of prostate cancer. *Genes & Dev* 14: 2410-2434.
- Aumüller G, Seitz J. 1990. Protein secretion and secretory process in male accessory sex glands. *Int Rev Cytol*, 121: 127-231.
- Balk SP. 2002. Androgen receptor as a target in androgen-independent prostate cancer. *Urology*, 60 (Suppl. 3A): 132-138.
- Banerjee PP, Banerjee S, Brown TR. 2001. Increased Androgen Receptor Expression Correlates with Development of Age-Dependent, Lobe-Specific Spontaneous Hyperplasia of the Brown Norway Rat Prostate. *Endocrinology*, 142(9): 4066–4075.
- Berges RR, Vulkanovic J, Epstein JI, Carmichel M, Cisek L, Johnson DE, Veltri RW, Walsh PC, Isaacs JT. 1995. Implication of cell kinetic changes during the progression of human prostatic cancer. *Clin Cancer Res*, 1(5): 473-480.
- Bonkhoff H, Remberger K. 1996. Differentiation pathways and histogenetic aspects of normal and abnormal prostatic growth: a stem cell model. *Prostate*, 28: 98-106.
- Bostwick DG, Ramnani D, Qian J. 2000. Prostatic intraepithelial neoplasia: Animal models 2000. *Prostate*, 43: 286-294.
- Brinkmann AO, Blok LJ, Ruiter PE, Doesburg P, Steketee K, Berrevoets CA, Trapman J. 1999. Mechanisms of androgen receptor activation and function. *J Ster Bioch and Mol Biol*, 69: 301-313.
- Brooks JR, Berman C, Nguyen H, Prahalada S, Primka RL, Rasmusson GH, Slater EE. 1991. Effet of castration, DES, Flutamide, and 5-alpha-redutase inhibitor, MK-906, on the growth of the Dunning rat prostatic carcinoma, R-3327. *Prostate*, 18(3): 215-227.
- Brum IS, Spritzer PM, Brentani MM. 2005. Biologia Molecular das Neoplasias de Próstata. *Arq Bras Endocrinol Metab*, 49(5): 797-804.
- Bruyninx M, Hennuy B, Cornet A, Houssa P, Daukandt M, Reiter E, Poncin J, Closset J, Hennen G. 1999. A novel gene overexpressed in the prostate of castrated rats: hormonal regulation, relationship to apoptosis and to acquired prostatic cell androgen independence. *Endocrinol*, 140(10): 4778-4799.
- Buchanan G, Irvine RA, Coetzee GA, Tilley WD. 2001. Contribution of the androgen receptor to prostate cancer predisposition and progression. *Cancer Metastasis Rev*, 20: 207–223.

- Carvalho HF, Line SRP. 1996. Basement membrane associated changes in the rat ventral prostate following castration. *Cell Biol Int*, 20: 809-819.
- Carvalho HF, Taboga SR, Vilamaior PSL. 1997a. Collagen type VI is a component of the extracellular matrix microfibril network of the prostatic stroma. *Tissue Cell*, 29: 163-170.
- Carvalho HF, Vilamaior PSL, Taboga SR. 1997b. Elastic system of the rat ventral prostate and its modifications following orchietomy. *Prostate*, 32: 27-34.
- Cooke PS, Young P, Cunha GR. 1991. Androgen receptor expression in developing male reproductive organs. *Endocrinology* 128: 2867-2873.
- Cordeiro RS, Scarano WR, Góes RM, Taboga SR. 2004. Tissue Alterations in the Guinea Pig Lateral Prostate Following Antiandrogen Flutamide Therapy. *BioCell* 28(1): 21-30.
- Cordeiro RS, Scarano WR, Taboga SR. 2005. Immunocytochemical Analysis of Androgen Receptor of the Gerbil Ventral Prostate. *Braz J Morphol Sci*, Supplement, p.36.
- Corradi LS, Góes RM, Carvalho HF, Taboga SR. 2004. Inhibition of 5- α -reductase activity induces stromal remodeling and smooth muscle de-differentiation in adult gerbil ventral prostate. *Differentiation*, 72(5): 198-208.
- Cunha GR. 1976. Epithelial-stromal interactions in development of the urogenital tract. *Int Rev Cytol*, 47: 137-194.
- Cunha GR, Donjacour AA, Sugimura Y. 1986. Stromal-epitelial interactions and heterogeneity of proliferative activity within the prostate. *Biochem Cell Biol*, 64: 608-614.
- Cunha GR, Hayward SW, Dahiya R, Foster BA. 1996. Smooth muscle-epithelial interactions in normal and neoplastic prostatic development. *Acta anat*, 155: 63-72.
- Debes JD, Tindall DJ. 2002. The role of androgens and the androgen receptor in prostate cancer. *Cancer Lett*, 187: 1-7.
- Dehm SM, Tindall DJ. 2006. Molecular regulation of androgen action in prostate cancer. *Journal of Cellular Biochemistry*, 99: 333–344.
- Dounjacour AA, Cunha GR. 1988. The effect of androgen deprivation on branching morphogenesis in the mouse prostate. *Dev Biol*, 128: 1-14.
- Droller MJ. 1997. Medical approaches in the management of prostatic disease. *Br J Urol*, 79 (Suppl.) 2: 42-52.
- Farla P, Hersmus R, Trapman J, Houtsmuller AB. 2005. Antiandrogens prevent stable DNA-binding of the androgen receptor. *Journal of Cell Science*, 118: 4187-4198.
- Feldman BJ, Feldman D. 2001. The development of androgen-independent prostate cancer. *Nature Rev Cancer*, 1: 34-45.

- Foster PMD, Harris MW. 2005. Changes in androgen-mediated reproductive development in male rat offspring following exposure to a single oral dose of flutamide at different gestational ages. *Toxicological Sciences*, 85: 1024–1032.
- Galbraith SM, Duchesne GM. 1997. Androgens and prostate cancer: Biology, pathology and hormonal therapy. *European Journal of Cancer*, 33(4): 545-554.
- Gao W, KIM J, Dalton JT. 2006. Pharmacokinetics and pharmacodynamics of nonsteroidal androgen receptor ligands. *Pharmaceutical Research*, 23(8):1641-1658
- Garraway LA, Lin D, Signoretti S, Waltregny D, Dilks J, Bhattacharya N, Loda M. 2003. Intermediate basal cells of the prostate: in vitro and in vivo characterization. *Prostate*, 55: 206-218.
- Goto K, Koizumi K, Takaori H, Fojii Y, Furuyama Y, Saika H, Suzuki H, Saito K, Suzuki K. 2004. Effects of flutamide on sex maturation behavior of offspring born to male rats treated during late pregnancy. *The Journal of Toxicological Sciences*, 29(5): 517-534.
- Hayward SW, Baskin LS, Haughney PC, Cunha AR, Foster BA, Dahiya R, Prins GS, Cunha GR. 1996. Epithelial development in the rat ventral prostate, anterior prostate and seminal vesicle. *Acta Anat*, 155: 81-93.
- Hayward SW, Rosen MA, Cunha GR. 1997. Stromal-epithelial interactions in the normal and neoplastic prostate. *Br J Urol*, 79 (Suppl.2): 18-26.
- Heinlein CA, Chang C. 2004. Androgen receptor in prostate cancer. *Endocr Rev*, 25(2): 276-308.
- Hsing AW, Reichardt JKV, Stanczyk FZ. 2002. Hormones and prostate cancer: current perspectives and future directions. *Prostate*, 52: 213-235.
- Hughes IA, Lim HN, Martin H, Mongan NP, Dovey L, Ahmed SF, Hawkins JR. 2001. Developmental aspects of androgen action. *Molecular and Cellular Endocrinology*, 185: 33–41.
- Jeffers GW, Klei TR, Enright FM. 1984. Activation of the jird (*Meriones unguiculatus*) macrophages by the filarial parasite Brugia paghangi. *Infect Immun*, 43(1): 43-48.
- Jenster G, Trapman J, Brinkmann AO. 1993. Nuclear import of the androgen receptor. *Biochem J*, 293: 761-768.
- Kramer AWJr. 1964. Body and organ weights and linear measurements of the Mongolian Gerbil. *Anatomical Records*, 150(4): 343-347.
- Krieg M, Nass R, Tunn S. 1993. Effect of aging on endogenous level of 5 α -Dihydrotestosterone, testosterone, estradiol, and estrone in epithelium and stroma of normal and hyperplastic human prostate. *J Clin Endocrinol Metab*, 77(2): 375-381.

- Leav I, Schelling KH, Adams JY, Merck FB, Alroy J. 2001. Role of canine basal cells in postnatal prostatic development, induction of hyperplasia, and sex-hormone-stimulated growth, and ductal origin of carcinoma. *Prostate*, 48: 210-224.
- Lee DK, Chang C. 2003. Expression and Degradation of Androgen Receptor: Mechanism and Clinical Implication. *The Journal of Clinical Endocrinology & Metabolism*, 88(9): 4043-4054.
- Lucia MS, Bostwick DG, Bosland M, Cockett ATK, Knapp DW, Leav I, Pollard M, Rinker-Schaeffer C, Shirai T, Watkins BA. 1998. Workgroup I: Rodent models of prostate cancer. *Prostate*, 36: 49-55.
- Marker PC, Donjacour AA, Dahiya R, Cunha GR. 2003. Hormonal, cellular, and molecular control of prostatic development. *Developmental Biology*, 253: 165-174
- McEntee MF, Epstein JI, Syring R, Tierney LA, Strandberg JD. 1996. Characterization of prostatic basal cell hyperplasia and neoplasia in aged macaques: comparative pathology in human and nonhuman primates. *Prostate*, 29: 51-59.
- McKenna N, Bell SK. 1997. Classification of mammals above the species level. Columbia University Press. New York. 631p.
- Meeker AK, Sommerfeld H J, Coffey DS. 1996. Telomerase is activated in the prostate and seminal vesicles of the castrated rat. *Endocrinology*, 137(12): 5743-6.
- Metzger DL, Kerrigan JR. 1993. Androgen receptor blockade with flutamide enhances growth hormone secretion in late pubertal males: Evidence for independent actions of estrogen and androgen. *Journal of Clinical Endocrinology and Metabolism*, 76(5): 1147-1152.
- Miyata K, Yabushita S, Sano M, Miyashita K, Okuno Y, Matsuo M. 2003. Effects of perinatal exposure to flutamide on sex hormone responsiveness in F1 male rats. *The Journal of Toxicological Sciences*, 28(3): 149-163.
- Mohler JL, Gregory C W, Ford OH, Kim D, Weaver CM, Petrusz P, Wilson EM, French FS. 2004. The androgen axis in recurrent prostate cancer. *Clin Cancer Res*, 10: 440-448.
- Montalvo L, Sánchez-Chapado M, Prieto JC, Carmena MJ. 2002. Regulation of the expression of protein kinase C isoforms in rat ventral prostate: effects of age, castration and flutamide treatment. *Life Sci*, 71: 2257-2266.
- Muller M, Nielsen JT. 1979. Effect of superior cervical ganglionectomy on monoamine content in the epithalamic area of the Mongolian Gerbil – a fluorescence histochemical study. *Cell Tissue Res*, 201(1): 1-9.

- Narayan P, Trachtenberg J, Lepor H, Dedruyne EMJ, Tewari A, Stone N, Das S, Cruz-Jimenez JE, Shearer R, Klimberg I, Schellhammer PF, Costello AJ. 1996. A dose-response study of the effect of flutamide on benign prostatic hyperplasia: Results of multicenter study. *Urology*, 47(4): 497-504.
- Nawa Y, Horii Y, Okada M, Arizono N. 1994. Histochemical and cytochemical characterizations of mucosal and connective tissue mast cells of Mongolian gerbils (*Meriones unguiculatus*). *Int Arch Allergy Immunol*, 104(3): 249-254.
- Nemeth JA, Lee C. 1996. Prostatic ductal system in rats: regional variation in stromal organization. *Prostate*, 28: 124-128.
- Neri R. 1989. Pharmacology and pharmacokinetic of flutamide. *Urology*, 34(4): 46-56.
- Nevalainen MT, Valve EM, Anhonen T, Yagi A. 1997. Androgen-dependent expression of prolactin in rat prostate epithelium in vivo and organ culture. *Faseb J*, 11: 1297-1307.
- Nishiyama T, Hashimoto Y, Takahashi K. 2004. The Influence of Androgen Deprivation Therapy on Dihydrotestosterone Levels in the Prostatic Tissue of patients with prostate cancer. *Clin Cancer Res*, 10: 7121-7126.
- Noldus J, Ferrari M, Prestigiacomo A, Stamey TA. 1996. Effect of flutamide and flutamide plus castration on prostate size in patients with previously untreated prostate cancer. *Urology*, 47: 713-718.
- O'Neill AJ, Boran SA, O'keane C, Coffey RNT, Hegarty NJ, Hegarty P, Gaffney EF, Fitzpatrick JM, Watson RWG. 2001. Capase 3 expression in benign prostatic hyperplasia and prostate carcinoma. *Prostate*, 47: 183-188.
- Oliveira SM. 2005. A próstata ventral do gerbilo frente às diferentes formas de castração e subsequente reposição hormonal por testosterona. 65f. Tese (Mestrado em Biologia Celular e Estrutural) – Instituto de Biologia, Universidade Estadual de Campinas, Campinas.
- Pinheiro PFF, Almeida CCD, Segatelli TM, Martinez M, Padovani CB, Martinez FE. 2003. Structure of the pelvic and penile urethra – relationship with the ducts of the sex accessory glands of the Mongolian gerbil (*Meriones unguiculatus*). *J Anat*, 202: 431-444.
- Pollard M, Luckert PH. 1987. Autochthonous prostate adenocarcinomas in Lobund-Wistar rats: a model system. *Prostate*, 11: 219-227.
- Pompeo ACL, Damião R, Aguinaga AS. 1998. Tumores Prostáticos - I Consenso Brasileiro: Câncer de Próstata. São Paulo.
- Raghaw S, Kuliyev E, Steakley M, Greenberg N, Steiner MS. 2000. Efficacious chemoprevention of primary prostate cancer by flutamide in an autochthonous transgenic model. *Cancer Res*, 60: 4093-4097.

- Rauch F, Polzar B, Stephan H, Zanotti S, Paddenberg R, Mannherz HG. 1997. Androgen ablation leads to an upregulation and intranuclear accumulation of deoxyribonuclease I in rat prostate epithelial cells paralleling their apoptotic elimination. *J Cell Biol*, 137: 909-923.
- Reese JH, McNeal JE, Redwine EA, Samloff IM, Stamey TA. 1986. Differential distribution of pepsinogen II between the zones of the human prostate and seminal vesicle. *J Urol*, 136: 1148-1152.
- Reiter E, Hennuy B, Bruyninx M, Cornet A. 1999. Effects of pituitary hormones on the prostate. *Prostate*, 38:159-165.
- Ruijter E, Van De Kaa C, Miller G, Ruiter D, Debruyne F, Schalken J. 1999. Molecular genetics and epidemiology of prostate carcinoma. *Endocr Rev*, 20(1): 22-45.
- Santos AF, Huang H, Tindall DJ. 2004. The androgen receptor: a potential target for therapy of prostate cancer. *Steroids*, 69: 79–85
- Santos FCA, Carvalho HF, Góes RM, Taboga SR. 2003. Ultrastructural characterization of secretory cell in the gerbil female prostate. *Tissue Cell*, 34: 273-282.
- Scarano WR, Vilamaior PSL, Taboga SR. 2006. Tissue evidence of the testosterone role on the abnormal growth and aging effects reversion in the Gerbil (*Meriones unguiculatus*) prostate. *The Anatomical Record Part A* 288a:1190–1200.
- Scarano WR, Cordeiro RS, Góes RM, Taboga SR. 2004. Intraepithelial alterations in the Guinea pig lateral prostate at different ages after estradiol treatment. *Journal of Submicroscopy Cytology and Pathology*, 36(2): 141-148.
- Scher HI, Buchanan G, Gerald W, Butler LM, Tilley WD. 2004. Targeting the androgen receptor: improving outcomes for castration-resistant prostate cancer. *Endocr Relat Cancer*, 11: 459-476.
- Schwentker V. 1963. The gerbil - a new laboratory animal. *Veterinarian*, 6(4): 5-9.
- Segatelli TM, França LR, Pinheiro PFE, Almeida CCD, Martinez M, Martinez FE. 2004. Spermatogenic cycle length and spermatogenic efficiency in the gerbil (*Meriones unguiculatus*). *Journal of Andrology*, 25(6): 872-880.
- Shulz WA, Burchardt M, Cronauer MV. 2003. Molecular biology of prostate cancer. *Mol Hum Reprod*, 9(8): 437-448.
- Singh P, Uzgare A, Litvinov I, Denmeade SR, Isaacs JT. 2006. Combinatorial androgen receptor targeted therapy for prostate cancer. *Endocrine Related Cancer*, 13: 653–666.
- Singh SM, Gauthier S, Labrie F. 2000. Androgen Receptor Antagonists (Antiandrogens): Structure-Activity Relationships. *Current Medicinal Chemistry*, 7:211-247.

- Slayter MV, Anzano MA, Kadomatsu K, Smith JM, Sporin MB. 1994. Histogenesis of induced prostate and seminal vesicle carcinoma in Lobund-Wistar rats: a system for histological scoring and grading. *Cancer Res*, 54: 1440-1445.
- Sogani PC, Vagaiwala MR, Ehitmore Jr WF. 1984. Experience with flutamide in patients with advanced prostatic cancer without prior endocrine therapy. *Cancer*, 54: 744-750.
- Srougi M. 1992. Câncer de próstata. A revista da clínica médica, Nov –Dez. pp. 3-18.
- Sufrin G, Coffey DS. 1975. Flutamide: Mechanism of action of a new nonsteroidal antiandrogen. *Investigate Urology*, 13(6): 429-434.
- Sugimura Y, Cunha GR, Donjacour AA. 1986. Morphogenesis of ductal networks in the mouse prostate. *Biol Reprod*, 34: 961-971.
- Tang DG, Porter AT. 1997. Target to apoptosis: a hopeful weapon for prostate cancer. *Prostate*, 32: 284-293.
- Taplin ME, Ho SM. 2001. The endocrinology of prostate cancer. *J Clin Endocr Metabol*, 86(8): 3467-3477.
- Thomson AA, Foster BA, Cunha GR. 1997. Analyses of growth factor and receptor mRNA levels during development of the rat seminal vesicle and prostate. *Development*, 124: 2431-2439.
- Tuxhorn JA, Ayala GE, Rowley DR. 2001. Reactive stroma in prostate cancer progression. *J Urol*, 166: 2472-2483.
- Untergasser G, Madersbacher S, Berger P. 2005. Benign prostatic hyperplasia: age-related tissue-remodeling. *Exp Gerontol*, 40: 121-128.
- Vilamaior PSL, Felisbino SR, Taboga SR, Carvalho HF. 2000. Collagen fiber reorganization in the rat ventral prostate following androgen deprivation: An possible role for the smooth muscle cells. *Prostate*, 45: 253-258.
- Wells LJ, Cavanova MW, Maxwell EL. 1954. Genital abnormalities in castrated rats and their prevention by means of testosterone propionate. *Anat Rec*, 118: 109-133.
- Williams WM. 1974. The Anatomy of the Mongolian Gerbil. Tumblebrook Fram, Inc, USA. 107p.
- Zanettoni C, Taboga SR. 2001. Age-related modifications in stromal and epithelial compartments of the male prostate of *Meriones unguiculatus*. *Acta Microsc*, 3(c): 203-204.

VIII. ANEXOS

DECLARAÇÃO

Declaro para os devidos fins que o conteúdo de minha dissertação/tese de mestrado/doutorado intitulada “Expressão de Receptores Androgênicos no Lóbulo Ventral da Próstata do Gerbilo da Mongólia”

() não se enquadra no Artigo 1º, § 3º da Informação CCPG 002/06, referente a bioética e biossegurança.

() está inserido no Projeto CIBio (Protocolo nº _____), intitulado

(X) tem autorização da Comissão de Ética em Experimentação Animal (Protocolo nº 1214 – 1).

() tem autorização do Comitê de Ética para Pesquisa com Seres Humanos (?) (Protocolo nº _____).

Aluno(a)

Orientador(a)

Para uso da Comissão ou Comitê pertinente:

(X) Deferido () Indeferido

Nome: Prof. Dra. ANA MARIA A. GUARALDO
Função: Presidente

Comissão de Ética na Experimentação Animal
CEEA/IB - UNICAMP